

Review on Sterility Testing

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ABSTRACT: All drug products and the equipment used to prepare drugs must undergo sterility testing of final product which is mandatory. The process of sterility testing or sterilization has different conditions, environment, temperatures for different methods and the products given by USP. Sterility testing plays major role in detecting the microorganisms and remove or kill them present on the substance, which helps the manufacturer's reputation and product safety. The process of sterility testing must be done at specific conditions given by USP by professionals. The prevention of product from microbial contamination also plays a vital role. This article describes the process, conditions environmental temperatures and suitable methods for the sterility testing of the given substance or product.

Keywords : microbial contamination, sterility testing, sterilization.

I. INTRODUCTION :

Sterility test is a set up strategy for identifying the presence of practical types of microorganisms in or on completed drug items. Sterility, in this sense, implies that an item is liberated from suitable microorganisms (albeit not really metabolic side-effects or on the other hand poisons). The exemplary structure sterility test analyzes a drug item in contact a culture medium, as a method of identifying the conceivable presence of suitable microorganisms. The test is required for all aseptically filled items. Lately various new innovation stages have arisen. This has been encouraged by an adjustment in arrangement by the U.S. Food and Drug Administration (FDA), making the way for options to the pharmacopeia strategies. This short audit evaluates a portion of these innovations.[1][2]

History :

Since the introduction of sterility testing in 1936, extraordinary upgrades have been made in our capacity to recognize microbial pollution in

drug mixes. At the point when sterility testing was presented, in USP-National Formulary (NF) 11, it was suggested uniquely for fluid arrangements and required just a 7-day hatching period and one kind of culture medium. Over the a long time since USP-NF 11 was distributed, USP sterility testing strategies and the media suggested have been changed habitually in continuous endeavors to improve the discovery of microbiological defilement. Today, USP Chapter <71> requires the utilization of two culture , Soybean-Casein Digest media Medium (SCDM) and Fluid Thioglycollate Medium (FTM), and a 14-day hatching period.[3]

Sterility testing:

USP Chapter <71> states that "... sterility testing is a demanding strategy, where asepsis of the method should be guaranteed for a right understanding of results... "[4] The USP likewise expresses that alternative strategies might be used as long as they are approved and "will yield results comparable to, or better than, the outcomes created by the regular method." [4] Albeit the compendial sterility testing is the most broadly acknowledged strategy, it has intrinsic limitations[5-9]. In a survey distributed in the Pharmacopeial Forum, the US Pharmacopeial (USP) Expert Board liable for USP Chapter <71> Sterility Testing inspected the media also, hatching conditions suggested for compendial sterility testing[7]. Through this survey, lacks were perceived also, suggestions made, some of which are talked about in this article.

Testing conditions:

USP Chapter <71> states that SCDM and FTM ought to be hatched at $22.5^{\circ} \pm 2.5^{\circ}\text{C}$ and $32.5^{\circ} \pm 2.5^{\circ}\text{C}$, respectively.[4] SCDM is utilized for advancing the development of high-impact microbes and organisms, while FTM is utilized essentially for anaerobes however will develop a few aerobes. Both the essential also, optional writing have appeared, in any case, that these are not exactly ideal development conditions for some

microorganisms and fungi.[5,7,9-11] While the suggested temperature of $22.5^{\circ} \pm 2.5^{\circ}\text{C}$ for SCDM might be pointed at identifying ecological contamination, it has been very much reported that most clinically huge microbes develop at temperatures somewhere in the range of 25°C and 40°C and growths at temperatures among 25°C and 30°C .^{7,8} The producers of the parasitic development medium suggested in the USP suggests brooding temperatures in the reach from 25°C to 30°C .⁹ One of the proposals made by the USP Master Committee for Sterility Testing was to expand the brooding temperature to $27.5^{\circ} \pm 2.5^{\circ}\text{C}$ to improve the recuperation of microscopic organisms, yeasts, and molds—a change that still can't seem to occur.

Precautions against sterility testing :

1. Tests for sterility are to be done via prepared work force utilizing procedures and hardware which limit the dangers of unplanned microbial pollution of the tests and of the testing climate.
2. Tests for sterility ought to be directed in a tidy up room climate that is identical to the norm of tidy up room needed for the aseptic assembling of drug items.
3. Work force possessing the aseptic testing territory during sterility testing or related aseptic controls should wear sanitized over garments. The utilization of cleaned articles of clothing may be satisfactory under specific conditions.
4. All gear, vessels and materials with which the sterile test media or the merchandise under test may come into contact over the span of the testing ought to be disinfected preceding use. Favored techniques are warming in an autoclave so that all surfaces are held at a temperature of 121°C and presented to immersed steam for in any event 15 minutes, or by warming to, and holding for at any rate 2 hours at a temperature of 160°C in a hot air broiler, or by openness to a base ingested radiation portion of 25 kGy.
5. All substances added to the products tried, and all substances added to sterile media or brought into sterile film filtration units (other than the arrangements under test) ought to be disinfected before use by heat. On the off chance that this diminishes the viability of the test the best elective technique ought to be utilized.
6. Preceding disinfection, all vessels, substances or external dress to be utilized for the execution of

tests for sterility, or brought into the testing territory, ought to be suitably bundled or shut, to forestall access of miniature life forms. Each bundle or thing being cleaned should bear a visual pointer proper to the technique for disinfection to show that it has been prepared however the suitable change in the presence of the marker ought not be taken as an assurance of the sterility of the substance. Each bundle or on the other hand thing ought to be dated with the date of sanitization to aid right stock pivot.

7. The external surfaces of all bundles of gear, vessels, and so forth, which are presented into the aseptic testing climate (counting vessels of media and bundles or compartments of products to be tried) ought to be liberated from defilement quickly preceding their presentation into the aseptic climate: they ought to be cleaned or sanitized by a proper technique which doesn't bias the suitability of miniature organic entities which might be available in the arrangements to be tried. A pass-through incubate (or move box) is viewed as a component of the testing climate. On the off chance that the bundles are twofold wrapped and disinfected the external wrapping ought to be taken out only before the presentation of the bundle into the testing. [12]

Major point affecting in sterility testing:- Test strategy, the climate wherein the test is directed, The nature of the way of life conditions gave, Sample size, Sampling method.

Factors effecting sterility testing :

- (1) Examining
- (2) Culture media for sterility testing
- (3) Technique for testing
- (4) Perception and result

Types of culture media:-

- (i) Fluid thioglycollate medium
- (ii) Soya bean casein digest medium.

Types of media:-

- (i) Fluid thioglycollate medium
- (ii) Soyabean casein digest medium.

Fluid thioglycollate medium:

Explicit job of certain fixings. Essential planned for the way of life of anaerobic microbes.

Hatching of the media for 14 days at $30-35^{\circ}\text{C}$.

THIOGLYCOLATE MEDIUM

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L-Cystine	0.5 g
Agar	0.75 g
Sodium chloride	2.5 g
Glucose monohydrate/anhydrous	5.5/5.0 g
Yeast extract (water-soluble)	5.0 g
Pancreatic digest of casein	15.0 g
Sodium thioglycollate or	0.5 g
Thioglycollic acid	0.3 ml
Resazurin sodium solution (1 g/l of resazurin sodium), freshly p	1.0 ml
Water R	Upto 1000 ml

Sterilise in autoclave at 121 C for 20 mins
 pH after sterilization 6.9 to 7.3.

Soya-bean casein digest medium :

Principally proposed for the way of life of the two organisms and high-impact microbes. Explicit part

of certain fixings. Brooding of the media at 20-25 degreecelcius for 14 days.

SOYBEAN CAESIN DIGEST MEDIUM

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Pancreatic digest of casein	17.0 g
Papaic digest of soya-bean meal	3.0 g
Sodium chloride	5.0 g
Dipotassium hydrogen phosphate	2.5 g
Glucose monohydrate/anhydrous	2.5/2.3 g
Water R	Upto 1000 ml

pH after sterilization 7.1 to 7.5.

Techniques for testing :

Membrane filtration method and direct inoculation method.

Test methodology:The test strategy ought to be as per the pharmacopoeial technique utilized. Layer filtration of the item, with either an open or a shut framework, is the favored sterility test strategy. The channel ought to be pre-wetted, especially when little volumes and antimicrobials are tried. Filtration of the item ought to be trailed by the base number of washes of the layer with a reasonable washing liquid set up during approval examines.

The layer ought not be allowed to dry out between filtration steps. If the item can't be sifted, at that point direct immunization, inundation, in-situ hatching or mix techniques as proper. [15]

Membrane filtration method :There are various sorts of film channel that can be utilized for the sterility test. Basically these are isolated by properties: hydrophobic or hydrophilic; and by the essential material of production: nylon, cellulose acetic acid derivation, cellulose nitrate or polycarbonate. [13]

All standard channels have a porosity of 0.45µm with low item restricting attributes. For a standard fluid based item, hydrophilic channels are the most ordinarily utilized, (for example, those made from blended esters of cellulose). Hydrophilic channels are effectively wet with water. Hydrophilic channels can be wetted with basically any fluid, permitting the fluid to go through the channel successfully. [13]

Direct inoculation method: In the wake of moving the substance of the holder or compartments to be tried (for catgut and other careful stitches for veterinary use: strands) to the way of life medium, add an inoculum of a modest number of reasonable microorganisms (not more than 100 cfu) to the medium. In the two cases utilize similar microorganisms as those depicted above under Growth Promotion Test of Aerobes, Anaerobes, and Fungi. Play out a development advancement test as a positive control. Hatch all the compartments containing mode for not over 5 days. [14] On the off chance that plainly obvious development of microorganisms is gotten after the brooding, outwardly practically identical to that in the control vessel without item, either the item has no antimicrobial movement under the states of the test or such movement has been acceptably killed. The test for sterility may at that point be done minus any additional alteration. On the off chance that unmistakably obvious development isn't gotten within the sight of the item to be tried, outwardly practically identical to that in the control vessels without item, the item has antimicrobial movement that has not been acceptably dispensed with under the states of the test. Change the conditions in request to dispense with the antimicrobial action, and rehash the Method Suitability Test. [14]

Sterilization methods:

British Pharmacopeia (1993) perceives five techniques for the sanitization of drug items. These are: (I) dry warmth; (ii) warming in an autoclave (steam sanitization); (iii) filtration; (iv) ethylene oxide gas; and (v) gamma or electron radiation. Likewise, different methodologies including steam and formaldehyde and bright (UV) light have developed for use in specific circumstances. For every strategy, the conceivable changes of openness conditions are various, yet experience and item solidness necessities have commonly served to restrict this decision. By the by, it ought to be recalled that even the suggested strategies and regimens don't really show

comparable biocidal potential, yet essentially offer elective methodologies for application to a wide assortment of item types. Hence, each ought to be approved in their application to show that the base required degree of sterility confirmation can be accomplished[16]

Heat sterilization:

Warmth is the most dependable and generally utilized methods for sanitization, bearing the cost of its antimicrobial movement through obliteration of chemicals and other fundamental cell constituents. These deadly occasions continue at their generally fast in a completely hydrated state, in this manner requiring a lower heat input (temperature and time) under states of high dampness where denaturation and hydrolysis responses prevail, as opposed to in the dry state where oxidative changes happen. This technique for disinfection is restricted to thermostable items, yet can be applied to both dampness touchy and dampness safe items for which the British Pharmacopeia (1993) suggests dry (160- 180°C) and clammy (121-134°C) heat sanitization, individually. Where warm corruption of an item may perhaps happen, it can generally be limited by choosing the higher temperature range since the more limited openness times utilized by and large outcome in a lower partial corruption.[16]

Moist heat sterilization:

Wet warmth has been perceived as a proficient biocidal specialist from the beginning of bacteriology, when it was chiefly produced for the sanitization of culture media. It presently finds far and wide application in the preparing of numerous thermostable items also, gadgets. In the drug and clinical circle it is utilized in the sanitization of dressings, sheets, careful and demonstrative hardware, holders and terminations, and watery infusions, ophthalmic arrangements and water system liquids, notwithstanding the handling of grimy and debased things.

Cleansing by wet warmth generally includes the utilization of steam at temperatures in the range 121-134°C, and keeping in mind that elective techniques are accessible for the handling of items flimsy at these high temperatures, they infrequently offer a similar level of sterility confirmation and ought to be kept away from assuming there is any chance of this happening. The raised temperatures for the most part related with damp warmth sanitization strategies must be

accomplished by the age of steam under tension.[16]

Dry heat sterilization:

The deadly impacts of dry warmth on microorganisms are expected to a great extent to oxidative cycles which are less successful than the hydrolytic harm which results from openness to steam. Subsequently, dry warmth disinfection ordinarily utilizes higher temperatures in the reach 160-180°C and requires openness seasons of as long as 2 hours relying on the temperature utilized (area 10). Once more, bacterial spores are significantly more safe than vegetative cells, and their recorded obstruction changes notably relying on their level of dryness. In numerous early investigations on dry warmth obstruction of spores their water content was not enough controlled, so clashing information emerged with respect to the openness conditions important to accomplish viable cleansing. This was incompletely answerable for varieties in suggested openness temperatures and times in various pharmacopeias. Its application is for the most part confined to crystal and metal careful instruments (where its great vulnerability and non-destructive nature are of advantage), non-fluid thermostable fluids and thermostable powders. Practically speaking, the reach of materials which are really exposed to dry warmth disinfection is very restricted, and comprises to a great extent of things utilized in medical clinics. The major mechanical application is in the cleansing of glass bottles which are to be filled aseptically, and here the fascination of the cycle is that it accomplishes a satisfactory sterility affirmation level, however that it additionally obliterates bacterial endotoxins (results of Gram-negative microbes, otherwise called pyrogens, that cause fever when infused into the body). These are hard to dispose of by different methods. For the motivations behind depyrogenation of glass, temperatures of roughly 250°C are utilized.[16]

Gaseous sterilisation:

The synthetically receptive gases ethylene oxide (CH₂)₂, and formaldehyde (methanal, H.CHO) have wide range biocidal action, and have discovered application in the sterilization of re-usable careful instruments, certain clinical, demonstrative and electrical gear, and the surface sanitization of powders.

Disinfection measures utilizing ethylene oxide disinfection are undeniably more generally utilized on a global premise than those utilizing formaldehyde.

Ethylene oxide therapy can likewise be considered as an option in contrast to radiation disinfection in the business creation of expendable clinical gadgets. These strategies don't, notwithstanding, offer a similar level of sterility affirmation as warmth strategies and are for the most part held for temperature-delicate things.[16]

Radiation sterilization:

A few kinds of radiation discover a cleaning application in the assembling of drug and clinical items, head among which are quickened electrons (particulate radiation), gamma-beams and bright (UV) light (both electromagnetic radiations). The significant objective for these radiations is accepted to be microbial DNA, with harm happening as a result of ionization and free extreme creation (gamma-beams and electrons) or excitation (UV light). This last interaction is less harming and less deadly than ionization, thus UV light isn't as productive a cleansing technique as electron or gamma-light. As referenced before (segment 2), vegetative microscopic organisms by and large end up being the most delicate to illumination (with striking special cases, for example *Deinococcus* {*Micrococcus*) radiodurans), trailed by molds and yeasts, with bacterial spores and infections as the most safe (with the exception of UV light where shape spores end up being generally safe). The degree of DNA harm needed to deliver cell passing can shift and this, along with the capacity to complete powerful fix, likely chooses the opposition of the organic entity to radiation. With ionizing radiations (gamma-beam and quickened electrons), microbial opposition diminishes with the presence of dampness or broken down oxygen (because of expanded free extreme creation) and furthermore with raised temperatures.[16]

Abbreviations :

FD : food and drug administration

NF : national formulary

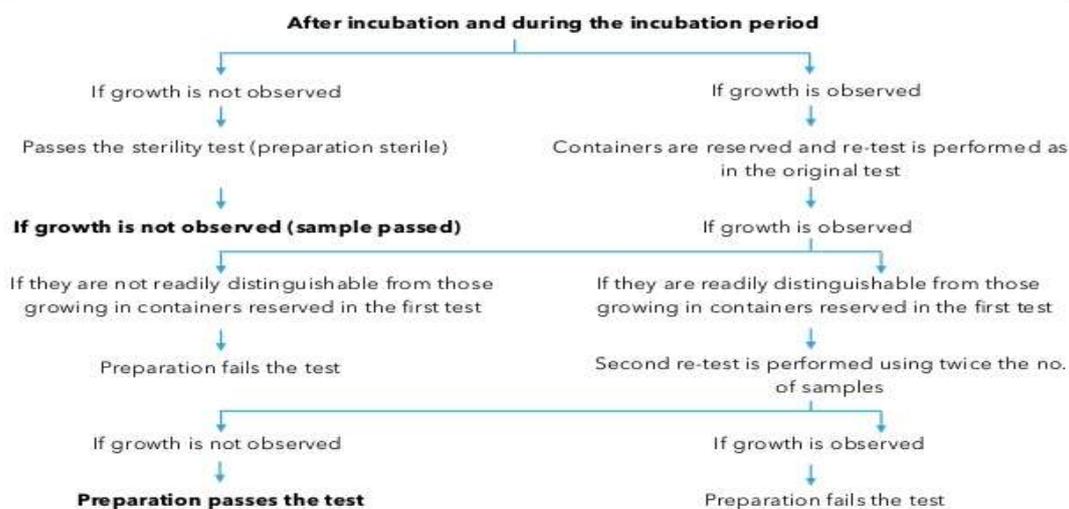
SCDM: Soybean-Casein Digest media Medium

FTM: Fluid Thioglycollate Medium

II. RESULT :

4. INTERPRETATION OF RESULTS

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III. CONCLUSION

This paper has inspected the Sterility Test. In doing so the two essential strategies have been delineated, along with a portion of the pragmatic parts of these strategies. The paper has additionally talked about the factual limits of the test and has accentuated the requirement for zeroing in on sterility affirmation, counting ecological controls during bunch fabricate, to build trust in the likelihood that the item is sterile.

REFERENCES :

- [1]. <http://dx.doi.org/10.4172/19485948.1000209> Sandle, J MicrobBiochem Technol 2015, 7:4
- [2]. Sykes G (1956) The technique of sterility testing. J Pharm Pharmacol 8: 573- 588
- [3]. Jason McGuire, BS Thomas C. Kupiec, PhD Analytical Research Laboratories Oklahoma City, Oklahoma
- [4]. United States Pharmacopeial Convention, Inc. United States Pharmacopeia 29–National Formulary 24. Rockville, MD: US Pharmacopeial Convention, Inc.; 2005: 2503–2513, 2521–2524, 2738–2742, 2744–2745, 2969–2976, 3041–3046, 3807–3808.
- [5]. Abdou MA. Comparative study of seven media for sterility testing. J Pharm Sci 1974; 63(1): 23–26.
- [6]. Bathgate H, Lazzari D, Cameron H et al. The incubation period in sterility testing. J Parenter Sci Technol 1993; 47(5): 254–257.
- [7]. Cundell AM. Review of the media selection and incubation conditions for the compendial sterility and microbial limit tests. Pharm Forum 2002; 28(6): 2034–2041.
- [8]. Ernst RR, West KL, Doyle JE. Problem areas in sterility testing. Bull Parenter Drug Assoc 1969; 23(1): 29–38.
- [9]. Moldenhauer J, Sutton SV. Towards an improved sterility test. PDA J Pharm Sci Technol 2004; 58(6): 284–286.
- [10]. Chapin KC, Murray PR. Media. In: Murray PR, Baron EJ, Pfaller MA et al, eds. Manual of Clinical Microbiology. 7th ed. Washington, DC: American Society for Microbiology; 1999: 1687–1707.
- [11]. Larone DH. Medically important fungi: A guide to identification. 4th ed. Washington, DC: American Society for Microbiology; 2002: 300–302.
- [12]. TGA guidelines for sterility testing of therapeutic goods September 2006 Page 10 of 36
- [13]. Baird, R. (1990): Monitoring microbiological quality: conventional testing methods in Denyer, S. and Baird, R. (Eds): Guide to Microbiological Control in Pharmaceuticals, Ellis Horwood: Sussex, pp125 145.



- [14]. 34(6) Sixth Interim Revision
Announcement: 71<71> STERILITY
TESTS
- [15]. website : <http://www.picscheme.org>
- [16]. blackwell science Ltd, pharmaceutical
microbiology, W.B.HUGO, A.D.RUSSELL,
sixth edition, pg:389-403