

Novel Antibacterial Agents Targeting Drug Resisitance Strains

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ABSTRACT: The growing threat of multidrug resistant bacteria has made the discovery of new antimicrobials more urgent than ever. The overuse and misuse of antibiotics, often available over the counter in many countries, have fuelled this resistance. The World Health Organization (WHO) has highlighted 12 bacterial families that pose the greatest danger to human health, where the effectiveness of current antibiotic treatments is severely limited.This paper explores new strategies for developing novel antibiotics that target human bacterial pathogens without any existing resistance. By leveraging advancements in research and technology, such as nanotechnology and computational methods (like in silico modelling and Fragment Based Drug Design), we have seen improvements in how antimicrobials work and their ability to target specific sites.Beyond traditional antibiotics, alternative approaches are emerging, including antimicrobial peptides, essential oils, agents that disrupt bacterial communication (anti quorum sensing), and other innovative compounds like darobactins, vitamin B6, bacteriophages, and cannabinoids.Prodrugs, particularly siderophores, have also shown great potential in creating a new generation of antimicrobials with enhanced effectiveness against multidrug-resistant bacteria. To effectively combat these resistant bacteria and prevent the spread of resistant infections, it's crucial to combine research and technological innovation with stricter regulations and public education on the responsible use of antibiotics, both in healthcare and agriculture.

I. INTRODUCTION:

Before the dawn of the 20th century, infectious diseases were the primary cause of high illness and death rates across the globe $[1]$. This trend began to shift with the discovery of antibiotics, a period that started with Alexander Fleming's groundbreaking discovery of penicillin in 1929. The advent of antibiotics revolutionized medicine, providing a powerful tool to combat bacterial infections that had previously been lifethreatening^[2,3]. However, this medical breakthrough came with an unintended consequencethe emergence of antibiotic resistance. Over time, the widespread and often careless use of antibiotics led to the development of resistant bacterial strains, diminishing the effectiveness of these once-powerful drugs.

The World Health Organization (WHO) has identified twelve families of bacteria that pose the greatest threat to human health. These bacteria have been classified into three priority groups based on the level of threat they represent: critical, high, and medium priority pathogens^[4]. The critical group includes bacteria like Acinetobacter, Pseudomonas, and Enterobacteriaceae, which are notorious for their resistance to multiple antibiotics. The high-priority group encompasses pathogens such as Enterococcus faecium and Staphylococcus aureus, while the medium priority group includes bacteria like Streptococcus pneumoniae and Shigella spp. Among these, the ESKAPE pathogens (Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumannii, Pseudomonas aeruginosa, and Enterobacter species) are particularly concerning due to their multidrug resistant (MDR) and extensively drug-resistant (XDR) characteristics $[5-$ ^{8]}. These pathogens have rendered even the most advanced antibiotics ineffective, creating a significant challenge for healthcare providers.

Given the escalating threat of AMR, there is an urgent need to develop novel strategies and approaches to combat this growing problem $[9-10]$. This review explores various innovative methods in the design and development of new antimicrobial agents. Among these strategies, the utilization of advanced technologies like nanotechnology and computational method such as in silico modelling and Fragment-Based Drug Design (FBDD) has led

to significant improvements in antimicrobial efficacy and enhanced selectivity towards specific bacterial targets. These technological advancements are paving the way for the development of more effective and targeted antimicrobial therapies.

Furthermore, the development of prodrugs, particularly siderophores, has shown great promise. These compounds can be engineered to improve drug delivery and efficacy, especially against multidrug-resistant bacteria. Combinatorial treatments, which involve the use of multiple drugs or therapeutic agents in tandem, are also being explored as a means to enhance the effectiveness of antimicrobial therapies and reduce the likelihood of resistance development.

In conclusion, addressing the challenge of antimicrobial resistance requires a multifaceted approach that combines innovative research, technological advancements, and public health initiatives $^{[11]}$. . The development of new antimicrobial agents, alternative treatments, and drug repurposing strategies, along with the responsible use of antibiotics, will be crucial in overcoming the threat posed by multidrug-resistant and extensively drug-resistant bacteria. By continuing to invest in these efforts, we can hope to preserve the effectiveness of antimicrobial therapies and protect global public health for future generations.

ANTIMICROBIAL RESISTANCE:

Antibiotics have played a crucial role in treating bacterial infections and diseases, greatly improving our health and life expectancy. Thanks to antibiotics, many illnesses that used to be fatal are now manageable. However, some bacteria have developed the ability to resist these drugs, leading to what we call antibiotic resistance, or antimicrobial resistance.When bacteria become resistant, it means that antibiotics can no longer effectively control or kill them $[12]$. These resilient bacteria can survive and even thrive despite the presence of antibiotics. Many types of bacteria that cause infections have the potential to develop resistance to at least some antibiotics. When bacteria become resistant to multiple antibiotics, they are known as multi-resistant organisms (MRO).Some bacteria are naturally resistant to certain antibiotics. For instance, benzylpenicillin doesn't have much effect on most bacteria in the human digestive system.Antimicrobial resistance is a serious public health concern. However, we can help prevent the spread of antibiotic resistance by avoiding unnecessary prescriptions, using antibiotics correctly when prescribed, and practicing good hygiene and infection control measures.

Bacteria resistant to antibiotics

Over time, some bacteria that were once easily defeated by common antibiotics have evolved and developed resistance. For example, Staphylococcus aureus (often known as "golden staph") and Neisseria gonorrhoeae (the bacteria responsible for gonorrhea) are now almost always resistant to benzyl penicillin. In the past, these infections were routinely treated with penicillin, but today, the rise in antimicrobial resistance is becoming a global issue.

Antimicrobial resistance poses a serious threat to public health. The most worrisome aspect is that some bacteria have become resistant to almost all antibiotics that are currently available $^{[13]}$. This makes treating infections far more complicated, turning once-manageable conditions into serious and costly health challenges. On top of that, these resistant bacteria can easily spread from person to person, leading to potential outbreaks that are difficult to control.

Here are some notable examples of bacteria that have developed significant resistance:

- **Methicillin-Resistant Staphylococcus Aureus (MRSA):** A type of staph bacteria that is resistant to several widely used antibiotics, making it harder to treat.
- **Vancomycin-Resistant Enterococcus (VRE):** This bacteria resists vancomycin, one of the antibiotics often used when other treatments fail.
- **Multi-Drug-Resistant Mycobacterium Tuberculosis (MDR-TB):** A strain of the bacteria that causes tuberculosis, resistant to the most potent TB drugs.
- **Carbapenemase-Producing Enterobacterales (CPE):** A group of bacteria that have developed resistance to carbapenems, a class of last-resort antibiotics.

These examples illustrate the growing challenge of antimicrobial resistance. As these bacteria continue to adapt and outsmart our current treatments, infections become increasingly difficult to manage. The spread of these resistant bacteria means that diseases we once controlled easily can now pose severe threats to our health, requiring

more complex and costly treatments. This underscores the urgent need for responsible antibiotic use, continued research, and strong public health measures to prevent the spread of resistant bacteria.

ANTIMICROBIAL PEPTIDES:

Antimicrobial peptides (AMPs) are small molecules made up of a chain of amino acids, usually ranging from 10 to 60 in number. These peptides have a unique feature called amphipathic properties, which allows them to easily interact with cell membranes or penetrate into the cytosol of microorganisms $^{[14]}$. The effectiveness of AMPs against bacteria and other microbes depends on several factors, including their charge, structure, sequence length, concentration, hydrophobicity (how they interact with water), and the composition of the target membrane $^{[15]}$.

AMPs that are produced by ribosomes and contain only natural amino acids can be classified into different types based on their structure. Some examples include:

- Linear, α-helical peptides: These have a spiral shape and include peptides like cecropins, magainins, and melittins.
- Peptides rich in specific amino acids: For example, PR-39, which is rich in proline and arginine, or indolidin, another type of specialized peptide.
- Peptides with disulfide bonds: These bonds add stability to the peptide's structure, with defensins and protegrin being examples.

Overall, AMPs are a diverse group of molecules with powerful antimicrobial properties, making them a key area of interest in the development of new treatments for infections^[16-19].

Mechanism of Action of Antimicrobial Peptides:

Research has shown that antimicrobial peptides (AMPs) work in a way that's quite different from traditional antibiotics used to treat infections. To improve how we design and develop these peptides, it's important to understand how they interact with microbial membranes.AMPs interact with bacterial membranes in a unique way that depends on the specific lipids in those membranes^[20]. When AMPs come into contact with bacterial cell membranes, they can disrupt the structure of both the inner and outer membranes, leading to cell death $[21]$. This happens because AMPs are attracted to the negatively charged

surfaces of the bacterial membranes^[22]. They can damage the membrane directly, interfere with the production of essential proteins, DNA, and RNA, or target specific components inside the cell.The key to how AMPs work lies in the electrostatic forces between the positively charged AMPs and the negatively charged bacterial membranes $^{[23-26]}$. Both Gram-positive and Gram-negative bacteria have membranes rich in phospholipids like phosphatidylglycerol and cardiolipin, which attract these positively charged peptides.One common type of AMP is the cationic amphipathic α-helix. For these peptides, there are several models that explain their mechanism. One such model is the barrel-stave model, where the α-helix forms vertical pores in the bacterial membrane. In this model, the AMPs cluster together to form barrelshaped structures that create water-permeable pores across the membrane, leading to the disruption and death of the bacterial cell^[27,28].

Current antibacterial therapies and limitations:

Antibacterial therapies have been a cornerstone of modern medicine since the discovery of antibiotics in the early 20th century. The antibiotics in clinical use today are predominantly based on a few key mechanisms that target essential bacterial processes. These processes, however, are becoming less susceptible to current treatments due to the emergence of resistance^[29]. Let's delve deeper into the antibiotic classes, their mechanisms of action, and the growing limitations in their usage.

1. β-lactams (Penicillins, Cephalosporins, and Carbapenems): The β-lactam class of antibiotics revolutionized healthcare by effectively treating a wide variety of bacterial infections^[30]. These antibiotics work by inhibiting bacterial cell wall synthesis. However, the widespread overuse and misuse of β-lactams over the decades have led to the development of β-lactamases, enzymes that hydrolyze the β-lactam ring, rendering the antibiotic ineffective. This is particularly problematic in organisms such as E. coli and K. pneumoniae, where β-lactamase-mediated resistance is common^[31]. The development of extended-spectrum β-lactamases (ESBLs) and carbapenemases has further complicated the clinical landscape, limiting treatment options even in hospital settings where infections tend to be more severe and resistant^[32].

2. Quinolones (Ciprofloxacin, Levofloxacin): Quinolones are potent antibiotics that target bacterial DNA replication machinery by inhibiting key enzymes such as DNA gyrase and topoisomerase $IV^{[33]}$. These enzymes play crucial roles in maintaining the supercoiled state of bacterial DNA, which is necessary for replication and transcription. Furthermore, efflux pumps, especially in Gram-negative bacteria, actively transport quinolones out of the cell, lowering their intracellular concentration and allowing bacteria to survive even in the presence of the drug.

3. Macrolides (Erythromycin, Azithromycin): These antibiotics inhibit protein synthesis by binding to the 50S ribosomal subunit, a component of the bacterial ribosome responsible for protein production^[34].One of the most prevalent is the methylation of ribosomal RNA (encoded by the erm gene), which prevents macrolides from binding to the ribosome. In addition, bacteria have evolved efflux pumps that expel macrolides from the cell, reducing their efficacy.

4. Tetracyclines (Doxycycline, Minocycline): Like macrolides, tetracyclines target protein synthesis, but they bind to the 30S ribosomal subunit instead. This prevents the attachment of tRNA molecules, which carry amino acids necessary for protein synthesis, thereby inhibiting bacterial growth. Despite these challenges, tetracyclines remain a vital part of the antibiotic arsenal, especially for treating atypical bacteria such as Chlamydia and Rickettsia^[35].

Limitations and the growing threat of resistance:

The effectiveness of these therapies is increasingly compromised by the ability of bacteria to develop and transmit resistance mechanisms. These mechanisms include:

Enzymatic degradation: The production of enzymes like β-lactamases and carbapenemases is one of the most widespread and clinically significant forms of antibiotic resistance. These enzymes break down the core structure of βlactams and other antibiotics, nullifying their antibacterial activity. Some bacterial species can even acquire genes for these enzymes through horizontal gene transfer, spreading resistance across different species and environments^[36].

Efflux pumps: Many bacteria, particularly Gram-negative organisms, have developed multidrug efflux pumps that expel a wide range of antibiotics from the cell. The AcrAB-TolC system in E. coli is one example of an efflux pump that contributes to resistance against multiple classes of antibiotics, including β-lactams, quinolones, and tetracyclines^[37].

Target modification: Bacteria can evolve mutations in the targets of antibiotics, such as PBPs, ribosomal subunits, or DNA gyrase, making these targets less susceptible to drug binding. For example, mutations in PBPs are a key factor in methicillin resistant Staphylococcus aureus (MRSA), while changes in DNA gyrase are commonly seen in quinolone-resistant bacteria^[38].

Mechanisms of resistance to antibiotics

Antibiotic resistance is a complex issue, and bacteria can resist treatment through several different mechanisms. Here's a breakdown of the main ways bacteria develop resistance:

Alterations in Drug Targets: Antibiotics work by targeting specific areas in bacteria, such as ribosomes (which make proteins) or enzymes. For instance, many bacteria resist macrolide antibiotics by altering their ribosomal targets. Similarly, bacteria like Staphylococcus aureus or Streptococcus pneumoniae can become resistant to penicillin by changing the penicillin-binding proteins. This resistance is often due to mutations that affect the target sites of various antibiotics, including beta-lactams, quinolones, glycopeptides, macrolides, tetracyclines, and rifampicin^[39].

Enzymatic Inactivation: Some bacteria produce enzymes that break down antibiotics, rendering them ineffective. This is a major resistance mechanism. For example, beta-lactamase enzymes can destroy beta-lactam antibiotics like penicillin. Other enzymes can modify antibiotics, such as acetylases and phosphotransferases, which affect drugs like chloramphenicol and erythromycin^[40].

Reduced Permeability: Bacteria can become resistant by altering their cell membranes to prevent antibiotics from entering. They might also actively pump the drugs out before they can do any harm. For example, Pseudomonas aeruginosa can develop carbapenem resistance through changes in a specific membrane protein called OprD. Reduced membrane permeability is a key factor in resistance to quinolones and aminoglycosides

Active Efflux Pumps: Some bacteria have active pumping systems that expel antibiotics from their cells. This is particularly common with tetracyclines, which are pumped out of the cell before they can be effective. These pumps can also

help bacteria resist other types of antibiotics, including quinolones, certain macrolides, and betalactams. These pumps can be controlled by both plasmids (small DNA molecules) and chromosomal $\mathrm{genes}^{[41,42]}.$

Alternative Metabolic Pathways: Bacteria can develop resistance by using different metabolic pathways. For example, some bacteria can avoid the effects of sulfonamides and trimethoprim by obtaining folate from their environment instead of synthesizing it themselves. This alternative approach helps them survive even in the presence of these antibiotics $^{[43]}$.

Novel targets, discovery approaches, and sources

Novel Antibiotic Targets:Antibiotics resistance is pushing researchers to find new ways to fight bacterial infections, and one promising approach is targeting specific parts of bacterial biology that are different from human biology. Here's a closer look at some of these new targets:

1.Amino Acid Biosynthesis: Bacteria make their own amino acids, which are crucial for protein synthesis and other functions $[44]$. Unlike humans, bacteria need to synthesize nine essential amino acids: leucine, isoleucine, valine, threonine, methionine, tryptophan, phenylalanine, histidine, and lysine. Since humans don"t produce these amino acids, targeting their synthesis in bacteria could be an effective strategy. For instance, lysine is important not only for protein synthesis but also for building bacterial cell walls^[45]. A newly discovered enzyme involved in lysine synthesis, called L,L-diaminopimelate aminotransferase (DapL), is found in certain pathogens like Chlamydia and Leptospira^[46]. Inhibiting DapL could lead to new, narrow-spectrum antibiotics.

2.Shikimate Pathway: This pathway, essential for making aromatic amino acids (like tryptophan), is another key target. Enzymes involved in this pathway, such as shikimate kinase and type II dehydroquinase, are critical for pathogens like Helicobacter pylori and Mycobacterium tuberculosis^[47] . Researchers are developing inhibitors for these enzymes. This pathway is also vital for some parasites, which opens up the possibility of creating selective treatments for diseases caused by protozoa like Toxoplasma gondii and Plasmodium falciparum^[48].

3.Cardiolipin Biosynthesis: Studies of antibioticresistant strains of Staphylococcus aureus and Bacillus subtilis have revealed changes in genes related to cardiolipin, a type of phospholipid $[49]$. This suggests that targeting cardiolipin biosynthesis might offer a new way to fight infections^[50].

4.Teixobactin: A groundbreaking antibiotic called teixobactin targets cell-wall synthesis by binding to specific molecules (Lipid II and Lipid III) that are essential for the cell walls of Gram-positive bacteria^[51]. Since these molecules don't change much through mutations, teixobactin might be less likely to encounter resistance $^{[52]}$.

5.LptD Inhibitors: Another promising approach involves targeting LptD, a protein in Pseudomonas aeruginosa involved in inserting lipopolysaccharides into the bacterial outer membrane^[53]. Inhibitors like POL70780 are being developed for Pseudomonas aeruginosa and could potentially be adapted for other bacteria because LptD is highly conserved $^{[54]}$.

6.Staphylopine: A peptide called staphylopine found in Staphylococcus aureus binds to metal ions like nickel, cobalt, zinc, copper, and iron. The genes involved in making staphylopine are also found in other pathogens like Yersinia pestis and Pseudomonas aeruginosa. Targeting staphylopine biosynthesis could lead to broad-spectrum antibiotics^[55].

Novel Discovery Approaches:

Finding new antibiotics has traditionally involved a lot of trial and error, often by growing bacteria and testing their extracts for new compounds^[56]. . However, newer genomic approaches are starting to take over this old "Grind and Find" method. Here's a look at how modern tools and techniques are revolutionizing the search for antibiotics:

1.AntiSMASH: This tool allows scientists to search for antibiotic-producing gene clusters in microbes by comparing them to known examples from plants, fungi, and bacteria. Essentially, it helps researchers identify which genes in a microbe might be responsible for making antibiotics by looking at similarities with already known antibiotic genes^[57].

2.Targeted Gene Mining: Another approach involves focusing on genes responsible for producing specific natural products. By understanding these genes, scientists can see if other strains of bacteria might also produce important intermediates of these products^[58]. This method also involves focusing on the final steps of the biosynthetic pathway, which can help in

developing a variety of antibiotic analogues from a single study^[59].

3.Engineering Microbes: Genomic analysis can uncover genes that regulate the production of antibiotics, such as repressors or activators $[60]$. This knowledge can be used to engineer microbes that produce antibiotics in higher amounts.

4.Challenges with Current Methods: Despite these advances, only a small fraction of bacterial diversity is regularly cultivated, which means we're missing out on many potential antibiotics. Many existing bioinformatics tools are designed with known molecules in mind, which can lead to repetitive discoveries. For example, Actinomycetes, particularly the Streptomyces genus, are overrepresented due to sampling $\overline{\text{biases}}^{\left[61,62\right]}$.

5.Improving Soil Metagenomics: Soil metagenomics, which involves sequencing DNA from environmental samples, has been challenging for discovering new antibiotics. Sequencing often doesn"t go deep enough to provide useful information. A new barcoding approach, using degenerate PCR primers, helps identify and sort biosynthetic gene clusters from environmental samples more effectively. This method, combined with the eSNaPD server, led to the discovery of malacidins, a new family of antibiotics that doesn"t develop resistance easily in lab conditions $[63]$.

6.Antibioticome: This public web application provides a comprehensive analysis of all known antibiotics, identifying their chemical origins, families, and targets. It uses a "retrobiosynthetic" algorithm to trace the building blocks of each antibiotic, which helps in discovering new candidates with unknown modes of action.

7.Predicting Resistance: A new web application correlates self-resistance in antibiotic-producing organisms with known resistance mechanisms^[64]. This tool can scan biosynthetic gene clusters for similarities to resistance genes, helping predict how new antibiotics might work and how bacteria might resist them.

8.Detecting Resistance Genes: Tools like DeepARG and the Resistance Gene Identifier (RGI) from the Comprehensive Antibiotic Resistance Database (CARD) help detect antibiotic resistance genes in both genomes and environmental samples^[65].

Novel Antimicrobial Sources:Researchers are turning to some pretty surprising sources to find new antibiotics, and it's leading to exciting discoveries. Here"s a rundown of how scientists are exploring unconventional sources for potential new drugs:

1.Marine Life: Marine organisms, like sponges, are proving to be rich sources of antibiotics. For example, a species of bacteria living in a marine sponge, Entotheonella, was found to produce over ten different antibiotics. This finding highlights how the unique environments of marine life can harbour novel antibiotic compounds^[66].

2.Insects and Their Microbes: Insects might also be a treasure trove for new antibiotics. The gut bacteria of the cotton leaf-worm, a common pest, produce an antibiotic peptide called mundticin. Similarly, bacteria from the guts of ants and other insects have been shown to make several new antibiotics, which could have applications in food safety and antimicrobial coatings^[67].

3.Amphibians and Reptiles: Amphibians like rainforest frogs have skin that resists bacterial and viral infections, thanks to antimicrobial peptides they produce. Researchers have tested these peptides against HIV and found promising results^[68]. The Komodo dragon, a large reptile known for its robust immune system, has also been found to produce a peptide with strong antibiotic properties, which has been used as a basis for developing a synthetic peptide with enhanced effects^[69-72].

4.Fungi and Bacteria Co-cultures: In the world of fungi and bacteria, combining different species can lead to the production of unique antibiotics not seen in isolated cultures. For example, co-cultures of bacteria and fungi have led to the discovery of new compounds, such as the antibiotic pestalone. This technique is proving to be a powerful method for uncovering novel antibiotics $^{[73-77]}$.

5.Microfluidic Chambers: Traditional methods of growing bacteria often miss out on many potential strains because they can"t simulate their natural environments well. New technologies like microfluidic chambers help by separating individual bacterial cells and allowing them to grow in conditions that better mimic their natural surroundings $^{[78,79]}$. This approach has already led to the cultivation of previously unculturable bacteria, such as Eleftheria terrae, which produces the antibiotic teixobactin^[80-87].

6.Venoms: Animal venoms are another rich source of antibiotics. For instance, bee venom contains a peptide called melittin that can kill various pathogens. Snake venoms, like those from the Agkistrodon halys and Bungarusfaciatus, have

been found to disrupt bacterial cell membranes and inhibit bacterial growth. Additionally, combining venom peptides with existing antibiotics has shown potential in fighting resistant bacteria^[88].

Inhibition of pathogenesis - anti virulence strategies:

Bacteria influence our bodies by using various tools called virulence factors. These factors help bacteria invade, infect, and evade our immune system^[89]. The virulence of a bacterium depends on its species and how many there are at the initial infection stage. Once inside, bacteria quickly activate their genetic machinery to produce these factors, which are essential for their survival and ability to cause disease^[90].

Some key virulence factors include:

- Adherence Factors: These help bacteria stick to host tissues.
- **Invasion Factors:** These allow bacteria to penetrate and spread within the host.
- **Polysaccharide Capsules:** These protect bacteria from being engulfed by immune cells.
- **Lipopolysaccharides:** These are components of the bacterial cell wall that can trigger strong immune responses.
- **Toxins:** These are harmful substances produced by bacteria that can damage host tissues.
- **Siderophores:** These help bacteria gather iron from the host, which is crucial for their growth.

One promising approach to fighting bacterial infections is to target and inhibit these virulence factors rather than trying to kill the bacteria directly. This method reduces the chance of bacteria developing resistance, as it doesn't put the same evolutionary pressure on them. Some key strategies in this approach include:

- **Blocking Quorum-Sensing:** Preventing bacteria from communicating with each other to coordinate their attack.
- **Inhibiting Toxin Production:** Reducing the ability of bacteria to produce harmful toxins.
- **Disrupting Adhesion:** Interfering with the bacteria's ability to stick to host cells.
- **Targeting Specific Virulence Genes:** Focusing on the specific genes that enable bacteria to cause disease

Anti-toxins:

Toxins are harmful substances produced by bacteria that can target and damage our cells. When these toxins enter our body, they act like foreign invaders and trigger an immune response. Our body produces specific antibodies, known as antitoxins, to neutralize these toxins^[91].

To help combat bacterial infections, scientists can create special antibodies, called monoclonal antibodies (mAbs), which are designed to target either bacterial surface structures or the toxins they produce^[92]. These anti-exotoxin antibodies work in several ways to reduce the harmful effects of bacteria:

- **Neutralizing Toxins:** The antibodies can bind to the toxins and prevent them from affecting our cells.
- **Promoting Phagocytosis:** The antibodies can mark the bacteria or toxins for destruction by immune cells, a process known as antibodydependent phagocytosis.
- **Enhancing Bacterial Killing:** The antibodies can activate the complement system, which helps to destroy bacteria more effectively.
- **Independent Killing:** The antibodies can directly impact bacteria through mechanisms that don"t rely on the immune system"s usual pathways.
- By targeting bacterial toxins and surface structures, these monoclonal antibodies help reduce the severity of infections and support our body's natural defenses^[93].

Biofilm formation inhibitor:

Bacteria can form dense clusters known as biofilms, which are responsible for about twothirds of all infections. These biofilms protect the bacteria and help them survive in tough conditions, making them up to 1,000 times more resistant to antibiotics[94] . This means standard antimicrobial treatments often fail to eliminate biofilms from infection sites, sometimes causing additional harm due to higher drug concentrations needed to try and penetrate the biofilm^[95].

Recent studies have shown that an increase in a molecule called cyclic di-GMP (c-di-GMP) promotes biofilm formation and enhances bacterial virulence^[96]. Given how biofilms contribute to both the persistence of infections and the resistance to drugs, it's crucial to find new treatments that target biofilm formation in novel $ways^{[97]}$.

Here are some promising strategies for combating biofilms:

1.Modifying c-di-GMP Levels: One approach involves changing the levels of c-di-GMP to prevent or reduce biofilm formation. By doing so, we can potentially disrupt the biofilm formation process and make bacteria more susceptible to treatment.

2.Inhibiting Quorum Sensing: Another strategy is to interfere with quorum sensing, the bacteria's way of communicating to coordinate their behaviour and biofilm formation. Nitric oxide (NO), a compound involved in cell signalling, has been found to help disperse biofilms. When NO is applied externally, it stimulates enzymes that break down c-di-GMP, causing bacteria to revert to a free-floating state rather than forming biofilms.

3.Using Synthetic Peptides: Researchers are also exploring synthetic peptides derived from natural antimicrobial peptides. For example, peptides like human cathelicidin LL-37, indolicidin, and cathuitamycins have been shown to inhibit biofilm formation. One particularly promising peptide, Antimicrobial Peptide 1018, has broad-spectrum anti-biofilm activity. It works by binding to and degrading certain signalling molecules crucial for biofilm formation and maintenance^[98].

Teichoic acids inhibitors:

Teichoic acids are important molecules found in the cell walls of certain bacteria. They play several crucial roles, including helping bacteria survive in harsh environments, resisting antibiotics, and evading our immune system. They are involved in sticking to surfaces, cell division and growth, maintaining cell shape, and regulating the balance of ions inside the cell. Importantly, humans don"t have teichoic acids, which makes them a promising target for developing new antibacterial treatments^[99].

Research has shown that targeting the synthesis of teichoic acids can make methicillin resistant Staphylococcus aureus (MRSA) more sensitive to β-lactam antibiotics, which are typically used to treat infections. Scientists are exploring ways to inhibit teichoic acid production to fight resistant bacterial infections [100].

One approach involves targeting the D-alanylation pathway, which is a key process in how bacteria modify their cell walls. By interfering with this pathway, researchers hope to disrupt teichoic acid synthesis and make bacteria more vulnerable to antibiotics^[101].

Several compounds are being studied for their ability to inhibit teichoic acid production, including:

- Targocil: A compound that targets the synthesis of Teichoic acids [102]
- **Tarocins:** A group of substances with potential to disrupt Teichoic acid formation [103]
- Tunicamycin: Another molecule that can inhibit the synthesis of Teichoic acids.

II. CONCLUSION

The synthesis and design of novel antibacterial agents targeting drug-resistant strains are of paramount importance in the ongoing battle against infectious diseases. The rise of resistant bacterial pathogens such as MRSA, VRE, and MDR-TB has rendered many traditional antibiotics ineffective, posing a serious threat to global public health. This review has explored various cuttingedge strategies in antibacterial drug design, including the development of peptide-based antibiotics, nanotechnology-enhanced drug delivery systems, and natural product-inspired drug discovery. These approaches offer new hope for overcoming the limitations of existing treatments. Furthermore, natural products continue to be a rich source of inspiration for novel antibacterial agents, providing diverse chemical scaffolds that can be optimized through synthetic modifications.Despite these promising developments, several challenges remain. The rapid mutation rates of bacteria and the complex mechanisms of resistance, such as biofilm formation and efflux pumps, complicate the development of universally effective
antibiotics.Future directions in antibacterial $antibiotics. Future$ directions in research should focus on a holistic approach that combines traditional and innovative strategies. This includes exploring novel drug targets, such as bacterial virulence factors and resistance mechanisms, as well as the development of new drug delivery systems that can bypass resistance mechanisms. The use of combination therapies, where multiple drugs with complementary mechanisms of action are used together, offers a promising strategy to prevent the emergence of resistance and enhance treatment efficacy.Moreover, ongoing research into the use of bacteriophages, CRISPR-Cas systems, and other biological tools for precision targeting of drugresistant bacteria could revolutionize the field of antibacterial therapy.

REFRENCES:

- [1]. Adedeji, W.A. The Treasure Called Antibiotics. Ann. Ib. Postgrad. Med. 2016, 14, 56–57.
- [2]. Breijyeh, Z.; et al.Resistance of Gram-Negative Bacteria to Current Antibacterial Agents and Approaches toResolve It. Molecules 2020, 25, 1340.
- [3]. Karaman, R.; et al.Resistance of Gram-Positive Bacteria to Current Antibacterial Agents and OvercomingApproaches. Molecules 2020, 25, 2888.
- [4]. Antimicrobial Resistance, C. Global burden of bacterial antimicrobial resistance in 2019: A systematic analysis. Lancet 2022,399, 629–655.
- [5]. Asokan, G.V.; et al.WHO Global Priority Pathogens List: A Bibliometric Analysis of Medline-PubMed for Knowledge Mobilization to Infection Prevention and Control Practices in Bahrain. Oman Med. J. 2019, 34, 184–193
- [6]. WHO. Priority Pathogens List of Antibiotic-Resistant Bacteria for Which New Antibiotics are Urgently Needed; WHO: Geneva, Switzer-land, 2017.
- [7]. Shrivastava, S.; et al.World health organization releases global priority list of antibiotic-resistant bacteriato guide research, discovery, and development of new antibiotics. J. Med. Soc. 2018, 32, 76.
- [8]. Mancuso, G.; et al.Bacterial Antibiotic Resistance: The Most Critical Pathogens. Pathogens 2021,10, 1310.
- [9]. Mulani, M.S.; et al.Emerging Strategies to Combat ESKAPE Pathogens in theEra of Antimicrobial Resistance: A Review. Front. Microbiol. 2019, 10, 539.
- [10]. Qadri, H.; et al.Novel Strategies to Combat the Emerging Drug Resistance in Human Pathogenic Microbes. Curr.Drug Targets 2021, 22, 1424–1436.
- [11]. De Oliveira, D.M.P.; et al.Antimicrobial Resistance in ESKAPE Pathogens. Clin. Microbiol. Rev. 2020, 33, e00181-19.
- [12]. Merrett GLB (2013) Tackling antibiotic resistance for greater global health security. Chatham House.
- [13]. Holmes AH, et al. (2016) Understanding the mechanisms and drivers of antimicrobial resistance. Lancet 387(10014): 176-187.
- [14]. Huan, Y.; et al.Antimicrobial Peptides: Classification, Design, Application and Research Progress in MultipleFields. Front. Microbiol. 2020, 11, 582779.
- [15]. Lachowicz, J.I.; et al.The Best PeptidomimeticStrategies to Undercover Antibacterial Peptides. Int. J. Mol. Sci. 2020, 21, 7349.
- [16]. Kapil, S.; et al.d-Amino acids in antimicrobial peptides: A potential approach to treat and combat antimicrobial resistance.Can. J. Microbiol. 2021, 67, 119–137.
- [17]. Moretta, A.; et al.Antimicrobial Peptides: A New Hope in Biomedical and Pharmaceutical Fields. Front. Cell. Infect. Microbiol. 2021, 11,668632.
- [18]. Mishra, A.K.; et al. Tryptophan-Rich and Proline-Rich Antimicrobial Peptides. Molecules 2018, 23, 815.
- [19]. Sarkar, T.; et al.Antimicrobial Peptides and Proteins: From Nature"s Reservoir to the Laboratory and Beyond.Front. Chem. 2021, 9, 691532.
- [20]. Mahlapuu, M.; et al.Antimicrobial peptides: An emerging category of therapeutic agents. Front.Cell. Infect. Microbiol. 2016, 6, 194.
- [21]. Rima, M.; et al. Antimicrobial Peptides: A Potent Alternative to Antibiotics.Antibiotics 2021, 10, 1095.
- [22]. Zhang, Q.Y.; et al. Antimicrobial peptides:Mechanism of action, activity and clinical potential. Mil. Med. Res. 2021, 8, 48. Strandberg, E.; Bentz, D.; Wadhwani, P.; Ulrich, A.S. Chiral supramolecular architecture of stable transmembrane pores formedby an αhelical antibiotic peptide in the presence of lyso-lipids. Sci. Rep. 2020, 10, 4710.
- [23]. Klubthawee, N.; et al.A novel, rationally designed, hybrid antimicrobialpeptide, inspired by cathelicidin and aurein, exhibits membrane-active mechanisms against Pseudomonas aeruginosa. Sci. Rep.2020, 10, 9117.
- [24]. Le, C.F.; et al.Intracellular Targeting Mechanisms by Antimicrobial Peptides. Antimicrob. Agents Chemother.2017, 61, e02340-16.
- [25]. Sinha, S.; et al. Structure and Interactions of A Host Defense Antimicrobial PeptideThanatin in Lipopolysaccharide

Micelles Reveal Mechanism of Bacterial Cell Agglutination. Sci. Rep. 2017, 7, 17795.

- [26]. Zainal Baharin, N.H.; et al. The characteristics and roles of antimicrobial peptides as potentialtreatment for antibiotic-resistant pathogens: A review. PeerJ 2021, 9, e12193.
- [27]. Mink, C.; et al.OverlappingProperties of the Short Membrane-Active Peptide BP100 with (i) Polycationic
- [28]. Bush, K.; et al. beta-Lactams and beta-Lactamase Inhibitors: An Overview. Cold Spring Harb. Perspect. Med. 2016, 6,a025247.
- [29]. Eyler, R.F.; et al. Clinical Pharmacology of Antibiotics. Clin. J. Am. Soc. Nephrol. Cjasn 2019, 14, 1080–1090.
- [30]. Patel, P.H.; et al.Macrolides; StatPearls: Treasure Island, FL, USA, 2022.
- [31]. Vazquez-Laslop, N.; et al.How Macrolide Antibiotics Work. Trends Biochem. Sci. 2018, 43, 668–684.
- [32]. Rusu, A.; et al. The Development of Third-Generation Tetracycline Antibiotics and New Perspectives. Pharmaceutics 2021, 13,2085
- [33]. Nguyen, F.; et al.Tetracycline antibiotics and resistance mechanisms.Biol. Chem. 2014, 395, 559–575.
- [34]. Pham, T.D.M.; et al.Quinolone antibiotics. Medchemcomm 2019, 10, 1719–1739.
- [35]. Visalli MA.:et al.2003. AcrAB multi-drug efflux pump is associated with reduced levels of susceptibility totigecycline (GAR-936) in Proteus mirabilis. Antimicrob Agents Chemother47:665–669
- [36]. E.P. Abraham.;et al. An enzyme from bacteria able todestroy penicillin, Nature 146 (1940) 837.
- [37]. Mangwani N, et al.Bacterial biofilms and quorum sensing: Fidelity in bioremediation technology. Biotechnology and Genetic Engineering Reviews. 2016;32(1-2):43-73
- [38]. Ayliffe GA. The progressive intercontinental spread ofmethicillinresistant Staphylococcus aureus. Clin Infect Dis 1997;24:S74-79.
- [39]. Yüce A. Antimicrobiallaçlarakaniz-maları. KlimikDergisi 2001; 14:41-46
- [40]. NikaidoH. Prevention of drug access to bacterial targets:permeability barriers and active efflux. Science 1994; 264:382-388.
- [41]. NikaidoH. Prevention of drug access to bacterial targets:permeability barriers and active efflux. Science 1994; 264:382-388.
- [42]. Jawetz Eet al.Medical Microbiology.East Norwalk, CT: Appleton & Lange, pp 137- 167, 1995.
- [43]. Oppenheim BA. Antibiotic resistance in Neisseria meningitidis. Clin Infect Dis 1997; 24:S98-101.
- [44]. Hudson, A.O.; et al.Biochemical and Phylogenetic Characterization of a NovelDiaminopimelate Biosynthesis Pathway in Prokaryotes Identifies a Diverged Form of Ll-DiaminopimelateAminotransferase. J. Bacteriol. 2008, 190, 3256–3263.
- [45]. Triassi, A.J.; et al.L,L-DiaminopimelateAminotransferase (Dapl): A Putative Target for the Development of Narrow-Spectrum AntibacterialCompounds. Front. Microbiol. 2014, 5, 509.
- [46]. Gonzalez-Bello, C. Inhibition of Shikimate Kinase and Type II Dehydroquinase for Antibiotic Discovery:Structure-Based Design and Simulation Studies. Curr. Top. Med. Chem. 2016, 16, 960–977.
- [47]. Lee, C.E.; et al. The Crystal Structure of Trpd, a MetabolicEnzyme Essential for Lung Colonization by Mycobacterium Tuberculosis, in Complex with Its SubstratePhosphoribosylpyrophosphate. J. Mol. Biol. 2006, 355, 784–797.
- [48]. Czekster, C.M.; et al. Steady-State Kinetics ofIndole-3-Glycerol Phosphate Synthase from Mycobacterium tuberculosis. Arch. Biochem. Biophys. 2009, 486,19–26.
- [49]. Shen, H.; et al. Characterization of the Putative TryptophanSynthase Beta-Subunit from Mycobacterium tuberculosis. Acta Biochim. Biophys. Sin. 2009, 41, 379–388
- [50]. Roberts, C.W.;et al.The Shikimate Pathway and Its Branches in Apicomplexan Parasites. J. Infect.Dis. 2002, 185 (Suppl. S1), S25–S36.
- [51]. Campbell, S.A.; et al.A CompleteShikimate Pathway in

Toxoplasma Gondii: An Ancient Eukaryotic Innovation. Int. J. Parasitol. 2004, 34, 5–13.

- [52]. Johnston, C.W.; et al. Assembly and Clustering of Natural Antibiotics Guides Target Identification. Nat.Chem. Biol. 2016, 12, 233–239.
- [53]. Ling, L.L.; et al.A New Antibiotic Kills Pathogens without Detectable Resistance. Nature2015, 517, 455–459.
- [54]. Srinivas, N.; et al. Peptidomimetic Antibiotics Target Outer-Membrane Biogenesisin Pseudomonas aeruginosa. Science 2010, 327, 1010–1013.
- [55]. Ghssein, G.; et al.Biosynthesis of a Broad-Spectrum Nicotianamine-Like Metallophore in Staphylococcusaureus. Science 2016, 352, 1105–1109.
- [56]. Weber, T.; et al.Antismash 3.0-a Comprehensive Resource for the Genome Mining of Biosynthetic GeneClusters. Nucleic Acids Res. 2015, 43, W237– W243.
- [57]. Blin, K.; et al.Antismash 4.0- Improvements in Chemistry Prediction and GeneCluster Boundary Identification. Nucleic Acids Res. 2017, 45, W36–W41.
- [58]. Smanski, M.J.; et al.Synthetic Biology to Access andExpand Nature"s Chemical Diversity. Nat. Rev. Microbiol. 2016, 14, 135–149.
- [59]. Rudolf, J.D.; et al. A Genetically Amenable Platensimycin- andPlatencin-Overproducer as a Platform for Biosynthetic Explorations: A Showcaseof Ptmo4, a Long-ChainAcyl-Coa Dehydrogenase. Mol. Biosyst. 2015, 11, 2717–2726.
- [60]. Smanski, M.J.; et al. Leveraging Ecological Theory to Guide Natural Product Discovery.J. Ind. Microbiol. Biotechnol. 2016, 43, 115–128.
- [61]. Katz, M.; et al.Culture-Independent Discovery of Natural Products from SoilMetagenomes. J. Ind. Microbiol. Biotechnol. 2016, 43, 129–141.
- [62]. Reddy, B.V.; et al. Versatile, Web-Based BioinformaticsPlatform for Surveying and Mining Natural Product Biosynthetic Diversity from Metagenomes. Chem. Biol.2014, 21, 1023–1033.
- [63]. Hover, B.M.; et al.Culture-Independent Discovery of the Malacidins as Calcium-

DependentAntibiotics with Activity against Multidrug-Resistant Gram-Positive Pathogens. Nat. Microbiol. 2018, 3,415–422.

- [64]. Arango-Argoty, G.; et al.A Deep LearningApproach for Predicting Antibiotic Resistance Genes from Metagenomic Data. Microbiome 2018, 6, $23.$
- [65]. Jia, B.; et al.Card 2017: Expansion and Model-Centric Curation of the Comprehensive AntibioticResistance Database. Nucleic Acids Res. 2017, 45, D566–D573.
- [66]. Johnston, C.W.; et al. Assembly and Clustering of Natural Antibiotics Guides Target Identification. Nat.Chem. Biol. 2016, 12, 233–239.
- [67]. Wilson, M.C.; et al.An Environmental Bacterial Taxon with a Large and Distinct Metabolic Repertoire. Nature2014, 506, 58–62.
- [68]. Shao, Y.; et al. Symbiont-Derived Antimicrobials Contributeto the Control of the Lepidopteran Gut Microbiota. Cell Chem. Biol. 2017, 24, 66–75.
- [69]. Vancompernolle, S.E.; et al.Antimicrobial Peptides from Amphibian Skin Potently InhibitHuman Immunodeficiency Virus Infection and Transfer of Virus from Dendritic Cells to T Cells. J Virol 2005,79, 11598–11606.
- [70]. M C Chung, E.; et al.Komodo Dragon-Inspired SyntheticPeptide Drgn-1 Promotes Wound-Healing of a Mixed-Biofilm Infected Wound. NPJ Biofilms Microbiomes2017, 3, 9.
- [71]. Zipperer, A.; et al.Human Commensals Producing a Novel Antibiotic Impair PathogenColonization. Nature 2016, 535, 511–516.
- [72]. Qin, Z.; et al.Hutchings, M.I.Formicamycins, Antibacterial Polyketides Produced By Streptomyces formicae isolated from African Tetraponeraplantants. Chem. Sci. 2017, 8, 3218–3227.
- [73]. Oh, D.C.; et al.Dentigerumycin: A Bacterial Mediator of an Ant-FungusSymbiosis. Nat. Chem. Biol. 2009, 5, 391–393.
- [74]. Smanski, M.J.; et al. Leveraging Ecological Theory to Guide Natural

Product Discovery.J. Ind. Microbiol. Biotechnol. 2016, 43, 115–128.

- [75]. Lu, C.; et al.A Novel Ansamycin, Naphthomycin K from Streptomyces sp. J. Antibiot. 2007, 60, 649–653.
- [76]. Chagas, M.B.O.; et al. Antimicrobial Activity of Cultivable Endophytic Fungi Associated WithHancornia Speciosa Gomes Bark. Open Microbiol. J. 2017, 11, 179–188.
- [77]. Verma, V.C.; et al.Endophytic Actinomycetes fromAzadirachta indica A. Juss.: Isolation, Diversity, and Anti-Microbial Activity. Microb. Ecol. 2009, 57, 749–756.
- [78]. Parthasarathy, A.; et al.Isolationand Genomic Characterization of Six Endophytic Bacteria Isolated from Saccharum sp (Sugarcane): Insightsinto Antibiotic, Secondary Metabolite and Quorum Sensing Metabolism. J. Genom. 2018, 6, 117–121.
- [79]. Bertrand, Set al.Metabolite Induction ViaMicroorganism Co-Culture: A Potential Way to Enhance Chemical Diversity for Drug Discovery. Biotechnol.Adv. 2014, 32, 1180–1204
- [80]. Tyc, O.; et al.Impact of Interspecific Interactions on Antimicrobial Activity among Soil Bacteria. Front. Microbiol. 2014, 5, 567.
- [81]. Mearns-Spragg, A.; et al. Cross-Species Induction and Enhancement of Antimicrobial Activity Produced by Epibiotic Bacteria from Marine Algae and Invertebrates, after Exposure to Terrestrial Bacteria. Lett. Appl.Microbiol. 1998, 27, 142–146.
- [82]. Ahmed, S.; et al. A Synthetic, Species-Specific Activator of Secondary Metabolism and Sporulation in Streptomyces coelicolor. ChemBioChem 2013, 14, 83–91.
- [83]. Slavov, N.; et al.Total Synthesis of the Marine Antibiotic Pestalone and Its Surprisingly Facile Conversion into Pestalalactone and Pestalachloride A. Angew. Chem. Int. Ed. Engl. 2010, 49, 7588–7591.
- [84]. Lincke, T.; et al.Closthioamide: An Unprecedented Polythioamide Antibiotic from the Strictly Anaerobic Bacterium Clostridium cellulolyticum. Angew.

Chem. Int. Ed. Engl. 2010, 49, 2011– 2013.

- [85]. Schroeckh, V.; et al.Intimate Bacterial-Fungal Interaction Triggers Biosynthesis of Archetypal Polyketides in Aspergillus nidulans. Proc. Natl. Acad. Sci. USA 2009, 106, 14558–14563.
- [86]. Nichols, D.; et al. Use of Ichip for High-Throughput in Situ Cultivation Of "Uncultivable" Microbial Species. Appl.Environ. Microbiol. 2010, 76, 2445– 2450.
- [87]. Lubke, L.L.; Garon, C.F. The Antimicrobial Agent Melittin Exhibits Powerful In Vitro Inhibitory Effects onthe Lyme Disease Spirochete. Clin. Infect. Dis. 1997, 25 (Suppl. S1), S48–S51.
- [88]. Samy, R.P.; et al. Viper Metalloproteinase (Agkistrodon halys Pallas) with Antimicrobial Activity against Multi-Drug-Resistant Human Pathogens. J. Cell Physiol. 2008, 216, 54–68.
- [89]. Xu, C.; et al.A Bactericidal Homodimeric Phospholipases A2 from Bungarus fasciatus Venom. Peptides 2007, 28, 969– 973
- [90]. Ling, L.L.; et al.A New Antibiotic Kills Pathogens without Detectable Resistance. Nature 2015, 517, 455–459.
- [91]. Yuan Ribet.D.;et al.How pathogens colonize their hosts and invade deeper tissues. Microbes Infect 2015;17(3):173– 83.
- [92]. Rasko DA.; et al. Anti-virulence strategies to combat bacteria-mediated disease. Nat Rev Drug Discovery 2010;9(2):117.
- [93]. Cegelski L, et al.The biology and future pro-spects of antivirulence therapies. Nat Rev Microbiol 2008;6(1):17.
- [94]. Goeders N, et al.Toxin-antitoxin systems as multilevel interaction systems. Toxins 2014;6(1):304–24.
- [95]. DiGiandomenico A, et al.Anti monoclonal antibodies: the next generation? Curr OpinMicrobiol 2015;1(27):78–85.
- [96]. Simões M, et al. A review of current and emergent biofilm control strategies. LWT-Food Sci Technol 2010;43(4):573–83.
- [97]. Anderson GG, et al. Intracellular biofilmlike pods in urinary tract infections. Science 2003;301(5629):105-7.
- [98]. Ha DG, et al.c-di-GMP and its effects on biofilm formation and dispersion: a

Pseudomonas aeruginosa review. Microbiol Spectrum 2015;3(2).

- [99]. Park SR, et al. Discovery of cahuitamycins as biofilm inhibitors derived from a convergent biosynthetic pathway. Nat Commun 2016;16(7):10710.
- [100]. Mishra B, et al.Anti-staphylococcal biofilm effects of human cathelicidin peptides. ACS Med Chem Lett 2015;7(1):117–21.
- [101]. Pasquina LW, et al.Teichoic acid biosynthesis as an anti-bacterial agent target. Curr OpinMicrobiol 2013;16(5):531–7.
- [102]. Farha MA, et al.Inhibition of WTA synthesis blocks the cooperative action of PBPs and sensitizes MRSA to β-lactams. ACS Chem Biol 2012;8(1):226–33.