

Advances in Antimicrobial Resistance (AMR) — Novel Therapeutic Strategies and Diagnostics

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ABSTRACT: Advances in Antimicrobial Resistance (AMR) — Novel Therapeutic Strategies and Diagnostics Abstract Antimicrobial resistance (AMR) is an escalating global health crisis characterized by the ability of microorganisms to evade treatment with once-effective drugs. The emergence and spread of resistant pathogens such as MRSA, VRE, CRE, and MDR-TB threaten the efficacy of modern medicine. The development of AMR is driven by genetic mutations, horizontal gene transfer, misuse of antibiotics, and mechanisms such as efflux pumps, biofilm formation, and enzymatic degradation of antimicrobials. This review explores innovative strategies to address AMR, focusing on advancements in diagnostics, therapeutics, and genome editing technologies. Rapid molecular diagnostics, including PCR, WGS, and microfluidics, are improving pathogen detection and resistance profiling. Therapeutically, novel antibiotics, adjuvants, vaccine strategies, combination therapy, and non-traditional approaches like quorum sensing inhibitors, phage therapy, and nanotechnology are redefining AMR management. Genomics plays a crucial role in tracking resistance genes and guiding personalized medicine. CRISPR technology, in particular, holds immense promise for disrupting resistance genes, eliminating plasmids, and developing targeted antimicrobials. Future directions emphasize integrating genomics with artificial intelligence, exploring natural product drug discovery, and leveraging synthetic biology. A multidisciplinary approach is essential to curtail AMR's impact and preserve the efficacy of antimicrobial therapies for future generations.

KEYWORDS

Antimicrobial resistance (AMR); multidrug-resistant organisms; genomics; CRISPR; novel therapeutics; biofilms; phage therapy; nanoparticles; antibiotic

adjuvants; artificial intelligence (AI); diagnostics; efflux pump inhibitors; antibiotic stewardship.

I. INTRODUCTION

The capacity of microorganisms, such as bacteria, viruses, fungi, and parasites, to withstand the effects of therapies that were once successful against them is known as antimicrobial resistance (AMR) [1]. Because of this resistance, which arises from genetic alterations and gene transfer across species, infections are able to persist and proliferate in the presence of antimicrobial medications. AMR includes a wide range of drugs, including antibiotics, antivirals, antifungals, and antiparasitic; it is not restricted to any one class of microbes or drug type [2].

MR is a serious and growing worldwide health emergency. The World Health Organization (WHO) estimates that drug-resistant diseases cause around 700,000 fatalities each year, and that figure will increase significantly in the absence of effective measures [4]. AMR has a crippling financial impact as well, including higher medical expenses, longer hospital stays, and lost productivity [6]. The potential for mild infections and routine operations to become life-threatening in the absence of adequate antibiotic therapies highlights the urgency of tackling AMR [7].

The fast development of resistance microorganisms is making traditional antibiotics, which have been the mainstay of treating infectious diseases for decades, less and less effective [9]. This problem has been made worse by the overuse and abuse of antibiotics in human medicine, agriculture, and animal husbandry, which has resulted in the rise of strains that are extensively drug-resistant (XDR) and multidrug-resistant (MDR). Often called "superbugs," these bacteria are resistant to several

different types of antibiotics, making routine treatments ineffective and making infection control more difficult [10].

With new resistance strains appearing at an alarming rate, the landscape of resistant diseases is constantly changing. Vancomycin-resistant Enterococci (VRE), carbapenem-resistant Enterobacteriaceae (CRE), and Methicillin-resistant *Staphylococcus aureus* (MRSA) are notable examples [11]. In clinical settings, these bacteria provide serious difficulties since they frequently cause serious infections with little available treatments. The emergence of pan-resistant bacterial strains, which are resistant to every known antibiotic, emphasizes even more how urgently creative strategies to counteract AMR are needed [13].

AMR's growing threat calls for quick fixes and creative solutions to protect public health. This work intends to support the worldwide effort to prevent antimicrobial resistance and guarantee the sustained effectiveness of antimicrobial therapies for future generations by investigating cutting-edge approaches including genomics, CRISPR, and new medicines [14].

Understanding Antimicrobial Resistance

Antimicrobial resistance (AMR) in microorganisms is a complex process that is mostly caused by horizontal gene transfer and genetic changes [15]. Genetic mutations can alter the target locations of antimicrobial medicines, making them less effective or useless altogether. They happen spontaneously during DNA replication. For example, resistance may result from changes in the genes encoding bacterial ribosomal RNA, which hinder the efficient binding of antibiotics such as tetracyclines.

The spread of resistance is further aggravated by horizontal gene transfer (HGT). This process allows for the transfer of genetic material between bacteria, often via plasmids, transposons, or integrins, which can carry multiple resistance genes [16]. Conjugation, transformation, and transduction are the main mechanisms of HGT. Conjugation involves the direct transfer of DNA between bacteria through pilus formation, while transformation involves the uptake of free DNA from the environment. Transduction, on the other hand, is mediated by bacteriophages that transfer DNA between bacteria [18].

These processes greatly contribute to the worldwide AMR epidemic by enabling the rapid spread of resistance traits among bacterial

populations. Apart from HGT and genetic changes, bacteria use a variety of different tactics to avoid the effects of antimicrobial drugs. One such mechanism is efflux pumps, which actively remove antibiotics from the bacterial cell, lowering the drug's intracellular concentration to levels below the fatal threshold [19]. Multidrug resistance may result from these pumps' ability to expel a variety of antimicrobial drugs or from their specificity to a single antibiotic.

Another important resistance tactic is the creation of biofilms. Complex bacterial colonies known as biofilms are coated in an extracellular matrix that the bacteria manufacture on their own and stick to surfaces. By serving as a physical barrier, this matrix prevents antimicrobial chemicals from penetrating and shields the bacterial cells inside. Furthermore, the transmission of resistance genes is facilitated by the close contact of bacteria within biofilms.

The synthesis of enzymes that break down or alter drugs, such as beta-lactamases, which hydrolyse beta-lactam antibiotics like cephalosporins and penicillin's, is another approach. In order to avoid the effects of antibiotics, bacteria can potentially change their metabolic processes, which guarantees their survival even when antimicrobial drugs are present [20].

Globally, there is increasing worry about the predominance of bacteria that are resistant to antibiotics [21]. The prevalence of resistant illnesses has significantly increased in both community and clinical settings, according to surveillance statistics. Methicillin-resistant *Staphylococcus aureus* (MRSA) and Vancomycin-resistant Enterococci (VRE), for instance, are common in hospitals and cause serious illnesses linked to healthcare [22]. Treatment of sexually transmitted diseases and urinary tract infections is becoming increasingly difficult due to the prevalence of resistant strains of pathogens such *Neisseria gonorrhoeae* and *Escherichia coli* in the population [24].

Each location has a different prevalence of resistance; in some, misuse of antibiotics, insufficient infection control methods, and a lack of regulatory frameworks contribute to higher rates. Because they have fewer resources and less access to healthcare, developing nations in particular have a difficult time controlling antimicrobial resistance (AMR).

Because of their high levels of resistance and the seriousness of the diseases they cause, a few bacteria are especially concerning. Priority pathogens have been designated by the World

Health Organization (WHO), and they include: • Methicillin-resistant Enterobacteriaceae (CRE), which are resistant to carbapenems and are frequently used as a last resort to treat infections that are resistant to several drugs; Multiple antibiotic-resistant *Staphylococcus aureus* (MRSA) is a leading source of infections acquired in hospitals and the community [26].

- Vancomycin-resistant Enterococci (VRE): these bacteria can cause severe infections, especially in those with weakened immune systems.
- The two most effective TB medications, isoniazid and rifampicin, cannot cure multidrug-resistant *Mycobacterium tuberculosis* (MDR-TB), which poses a serious risk to public health [27]. Developing effective methods to address this global health catastrophe requires an understanding of the epidemiology and mechanisms of antimicrobial resistance (AMR). A comprehensive strategy that incorporates surveillance, infection management, and the creation of novel antimicrobial drugs and treatments is needed to stop the spread of resistance [29].

Diagnostic Techniques for Antimicrobial Resistance (AMR)

Rapid and precise diagnostic methods must be developed in light of the global increase in antimicrobial resistance (AMR). Despite being commonly utilized, traditional methods have drawbacks such as complicated procedures and significant turnaround times (TAT). Among them are phenotypic techniques that track bacterial growth in the presence of antibiotics to ascertain susceptibility, such as disk diffusion, gradient testing, and broth microdilution. Sensitiser ARISTM 2X and VITEK®2 are examples of automated systems that increase throughput but are still too time-consuming for clinical usage [30].

More sensitivity and quicker findings are provided by molecular diagnostics. Assays based on PCR are frequently used to find resistance genes. For widespread monitoring, variants such as multiplex PCR, real-time PCR, and high-throughput qPCR have been used [31]. Because they do not require temperature cycling, isothermal amplification methods like RPA (Recombinase Polymerase Amplification) and LAMP (Loop-mediated isothermal amplification) are appropriate for point-of-care testing [32].

AMR diagnostics have been completely transformed by whole genome sequencing (WGS) and metagenomics, which enable thorough identification of resistance genes without the need

for previous culture. Both known and new ARGs are characterized using short-read (like Illumina) and long-read (like Oxford Nanopore) technologies. Interestingly, real-time sequencing data is provided by the Minion instrument, which allows for quick identification straight from clinical specimens [33]. MALDI-TOF MS is unique among non-traditional methods since it uses protein fingerprinting to quickly identify bacteria and detect resistance. Nevertheless, it has drawbacks with mixed samples and expensive implementation [34]. Additionally gaining traction are microfluidics-based systems that combine phenotypic and genotypic tests for quicker detection with less sample preparation. Rapid antimicrobial susceptibility testing (AST) from blood cultures is available through some commercial systems, such as BD Phoenix™ and Accelerate Pheno™ [35].

Antimicrobial Therapies and Novel Strategies to Control AMR

Modern medicine and public health are still seriously threatened by the worldwide antimicrobial resistance (AMR) dilemma. In order to successfully manage and control AMR, new measures must be put into place due to the declining effectiveness of current antibiotics and the concerning emergence of multidrug-resistant (MDR) pathogens. Antimicrobial stewardship, novel medication development, repurposing of current medicines, improving diagnostics, and the use of unconventional tactics like phage therapy, nanotechnology, and artificial intelligence (AI) are all necessary components of an integrated strategy to combat antimicrobial resistance (AMR).

Enhancing Antibiotic Stewardship and Drug Development

One important tactic to increase the potency of already available antibiotics is antibiotic stewardship. It is crucial to invest in the research and development of new antibiotics as well as to encourage appropriate usage in all facets of healthcare. Stewardship initiatives seek to decrease the overuse of antibiotics, improve treatment results, and slow the emergence of resistance. [36]

The majority of freshly licensed antibiotics, however, belong to classes that already have established resistance mechanisms. Recent data indicates that the discovery of novel antibiotics has stalled, with over 80% of newly licensed antibiotics coming from known classes. [37] One of the few novel antibiotics that works against multidrug-resistant Gram-negative bacteria, such as those that

produce AmpC β -lactamases and extended-spectrum β -lactamases (ESBLs), is cefiderocol, a new siderophore cephalosporin. [38] Regretfully, the existing pipeline is still unable to satisfy the pressing needs of the worldwide AMR crisis.

The Role of Vaccines in Combating AMR

By lowering the requirement for antibiotics, vaccination acts as a preventative measure that indirectly aids in the management of AMR. Vaccines are a useful tool in public health measures against AMR since, in contrast to antibiotics, they do not encourage resistance. To lower transmission and illness rates, immunization campaigns that target resistant bacteria—such as *Acinetobacter baumannii* and carbapenem-resistant Enterobacterales—are essential. [39]

Use of Antibiotic Adjuvants

Adjuvants are substances that increase the effectiveness of antibiotics, frequently by focusing on the processes behind bacterial resistance. The most prevalent kinds include membrane permeabilizers, efflux pump inhibitors, and β -lactamase inhibitors. Although clavulanic acid, sulbactam, and tazobactam are efficient β -lactamase inhibitors, they do not work against Metallo- β -lactamases, for which there are presently no therapeutic inhibitors. [40]

BLI-489 and LK-157, two of the more recent generations of β -lactamase inhibitors, have shown encouraging in vitro action against microbes that produce ESBL. [41] In order to restore antibiotic effectiveness against resistant bacteria, Melander and Melander stressed the significance of adjuvants in lowering the minimum inhibitory concentration (MIC) of antibiotics. [42] This is further supported by Laws et al., who show how adjuvants help maintain current therapy choices. [43]

Augmentin®, a combination of amoxicillin and clavulanic acid, is the quintessential example. By blocking β -lactamase enzymes, clavulanic acid prevents amoxicillin from breaking down and extends its useful life. [44] Despite the effectiveness of these combinations, resistance still arises, according to Worthington and Melander, which is why next-generation adjuvants are required. [44] Ceftazidime and avibactam work together to block the hydrolytic activity of AmpC β -lactamase, which gives *Pseudomonas aeruginosa* significant resistance. [45] Other non-antibiotic compounds, such as those enhancing the action of minocycline, have shown broad-spectrum potential against MDR organisms, including MRSA and *P. aeruginosa*. [46]

Efflux Pump Inhibitors (EPIs)

Antibiotics are expelled from bacterial cells by efflux pumps, which greatly increases resistance. Antibiotics can continue to function well inside the bacterial cell while EPIs block these pumps. EPIs are straightforward, strong, and well-tolerated substances that have the ability to reverse resistance, according to Lomovskaya and Watkins. [47] In *E. coli*, *K. pneumoniae*, and *S. aureus*, EPIs such as thioridazine, PA β N, and arylpiperazine NMP have demonstrated effectiveness in decreasing biofilm formation and regaining antibiotic sensitivity. [48] They are a crucial area of study for AMR because of their function in interfering with bacterial defense systems, particularly in illnesses linked to biofilms.

Combination Antibiotic Therapy

Antibiotic combinations are another tried-and-true method of combating resistance. By blocking distinct routes or specific locations within a single pathway, synergistic combinations might boost effectiveness and postpone the emergence of resistance. [49] For example, streptogramins block the same target by distinct ways, whereas trimethoprim and sulfamethoxazole inhibit successive stages in folate production. When it comes to treating complicated illnesses like malaria, HIV, and TB, combination treatment has proven very successful. [50] However, Coates et al. warn that in order to prevent antagonistic effects or unfavorable pharmacodynamic and pharmacokinetic interactions, such combinations need to be carefully chosen. [51]

Targeting Quorum Sensing (QS) and Biofilms

Quorum sensing (QS) is a process that bacteria employ to coordinate biofilm development and pathogenicity. One effective tactic to fight AMR is to disrupt QS. Cyclic-di-GMP and other nucleotide signaling molecules are essential for the formation of biofilms and bacterial adaptability. [52] Wu and Kali et al. highlighted how QS inhibitors lower bacterial virulence and biofilm durability. [53] [54] Inhibitors of diguanylate cyclase, which is in charge of c-di-GMP production, have demonstrated effectiveness in breaking up biofilms in *P. aeruginosa* and *A. baumannii*. [55] The chemical HAM improved *S. aureus* biofilm susceptibility to many antibiotic classes, as shown by Brackman et al. [56] In rat models, RNAIII-inhibiting peptides also successfully decreased MRSA graft infections, highlighting the potential of QS inhibitors as anti-biofilm therapies. [57] [58]

Disruption of Amyloid Structures in Biofilms

Curli are examples of bacterial amyloids that serve as structural elements in biofilms. Targeting these fibers compromises the integrity of the biofilm. In mice models of urinary tract infections, FN075, a 2-pyridone analog, has been shown to have decreased virulence and suppress curli formation in *E. coli*. [59] Romero et al. and further research demonstrated that substances like parthenolide and AA-861 can prevent the production of biofilms in *B. subtilis*. By destabilizing preexisting biofilms or preventing amyloid production, these substances offer a fresh way to boost antibacterial activity. [60] [61]

Phage Therapy and Adjunctive Approaches

Bacteriophages are used in phage therapy to target particular bacterial infections. Brives and Pourraz emphasized the low ecological impact, safety, and specificity of phage therapy. [62] Modified phages are used in Phico Therapeutics' SASPject™ technology to eradicate infections while protecting the natural microbiome. [63] Phage and antibiotic combinations have shown successful in eliminating MDR biofilms by utilizing two different methods. [64] Phages may also serve as adjuncts to probiotics and prebiotics, enhancing microbiome balance and therapeutic success. [65] Townsend et al. demonstrated that pre-treating biofilms with phages improves antibiotic penetration and efficacy, offering a valuable adjunctive therapy for stubborn infections. [66]

Nanotechnology and Targeted Drug Delivery

A new method for effectively delivering antibiotics and avoiding resistance mechanisms is offered by nanoparticles (NPs). Metallic, polymeric, and lipid-based nanoparticles (NPs) can transport medications, protect them against deterioration, and allow for prolonged release. [67] Additionally, nanocarriers lessen toxicity and increase bioavailability.

Conjugates of siderophores and drugs greatly improve intracellular delivery. Using ampicillin conjugates, Mollmann et al. showed a 1000-fold improvement in antimicrobial efficiency against Gram-negative bacteria. [68] The dual function of NPs as carriers and active antimicrobials was highlighted by Dey et al. and Chamundeeswari et al. [69] [70]

NPs cause membrane disruption, DNA replication inhibition, and reactive oxygen species generation. To further aid in the disintegration of biofilms, parthenolide and AA-861 both inhibit

bacterial amyloid formations. [71] When it comes to encapsulating antibiotics, improving their access into bacteria, and breaking down resistance barriers, liposomes are very useful. [72] According to Ferreira et al., their physicochemical flexibility accounts for their performance against resistant diseases. [73]

Optimizing Dosing Regimens

To optimize antibiotic efficacy and reduce resistance, appropriate dosage techniques are crucial. In order to reduce selective pressure, Negri et al. and Baquero and Negri promoted brief, high-dose regimens. [74] [75] Treatments should be customized according to the type of infection and patient-specific characteristics, according to Guillemot et al. [76] As advised by antimicrobial stewardship programs, practical uses include aminoglycoside interval dosing, prolonged β -lactam infusions, and modified fluoroquinolone dosages for the treatment of pneumonia. [77]

Artificial Intelligence in AMR Management

AI and machine learning (ML) are revolutionizing AMR diagnostics and drug development. Liu et al. discovered "abaucin," a novel antibiotic, by screening over 7,500 compounds using neural networks. [78] ML algorithms predict synergistic drug interactions, aiding in combination therapy design. [79] AI also enhances AMR surveillance by analyzing genomic and epidemiological data to predict emerging resistance patterns. Rabaan et al. and Amsterdam stress AI's utility in outbreak prediction and antimicrobial policy planning. [80][81] ML models improve public health responses by identifying high-risk populations and informing antimicrobial use strategies. These predictive insights can significantly reduce the burden of AMR through timely interventions. [82]

Genomics in Combating Antimicrobial Resistance

Genomics plays a pivotal role in understanding and combating antimicrobial resistance (AMR) [83]. Through whole-genome sequencing (WGS), scientists can identify pathogens at the strain level and detect genetic determinants of resistance, including mutations and novel resistance genes. This detailed genetic insight enhances our knowledge of how microbes adapt to antimicrobial agents. The advent of next-generation sequencing (NGS) has made genomic analysis faster and more

cost-effective, enabling quick detection of pathogens even in complex or low-abundance samples. Such data are critical in informing targeted therapies and improving infection control measures.

The tracking of resistance genes across populations and environments is also supported by genomics [84]. The mapping of the spread of resistance traits is aided by comparative genomic analysis of samples from various geographic regions. Metagenomics, which analyses DNA from entire microbial communities, is particularly helpful when traditional culture techniques are ineffective [85][86]. This approach identifies resistance elements in human microbiomes or environmental sources, guiding public health responses and antimicrobial stewardship. Advances in genomics have also transformed diagnostics [87]. Traditional culture-based tests are slow and occasionally fail to detect all resistance mechanisms. In contrast, genomic diagnostics, using tools like PCR and NGS, can quickly and accurately identify pathogens and resistance genes directly from clinical samples [88][89]. These technologies shorten the time to diagnosis, allowing for quicker and more effective treatment decisions.

Case studies highlight genetics' clinical significance. For example, WGS has greatly reduced the time required to start targeted therapy by speeding up the identification of drug-resistant TB. Similar to this, during hospital outbreaks, genomic technologies have helped restrict transmission and safeguard susceptible individuals by tracking the origin and spread of carbapenem-resistant Enterobacteriaceae (CRE) [90][91].

Another promising use of genomics in AMR management is the idea of personalized medicine [92], where clinicians can customize therapies to maximize efficacy while minimizing side effects and the risk of resistance by knowing the pathogen's genome as well as the patient's genetic profile [93]. Host genomics also helps predict treatment responses and optimize antimicrobial dosing.

The broad application of genomics in AMR is constrained by a number of obstacles, despite its potential. These include the creation of clinical recommendations, the integration of genomes into healthcare systems, and the requirement for reliable databases. Furthermore, deployment in environments with limited resources may be hampered by high prices and infrastructure requirements [94]. To guarantee that all areas profit from these developments, equitable access to genomic technologies is necessary. The key to fully

use genomics in the fight against AMR worldwide is sustained investment in infrastructure and research [95].

CRISPR Technology in Antimicrobial Resistance Management

With major ramifications for the control of antimicrobial resistance (AMR), CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats) technology has become a ground-breaking technique in genome editing (96). The CRISPR-Cas9 technology, which was initially developed from a bacterial immune system, allows for precise DNA editing and targeting. Bacteria convert stored viral sequences into guide RNAs when a virus infiltrates, guiding the Cas9 enzyme to the viral DNA and causing double-strand breaks. This technique has been modified to make very accurate genome edits in a wide range of species (97).

Various CRISPR systems, including CRISPR-Cas9, CRISPR-Cas12, and CRISPR-Cas13, have distinct functions. For example, Cas12 is known for its selectivity and decreased off-target effects, but Cas13 targets RNA and may have uses in the treatment of RNA viruses (98,99). Because of this diversity, CRISPR provides a flexible approach for addressing the intricate problem of AMR, where resistance mechanisms frequently incorporate both plasmid-borne genes and chromosomal abnormalities.

Targeted destruction of resistance genes is a promising use of CRISPR in AMR. CRISPR-Cas9 may introduce cuts that inactivate these genes by using guide RNAs that match resistance sequences. This makes bacteria vulnerable to drugs that were previously ineffective (100). For instance, beta-lactam antibiotic action can be restored by mutating the genes encoding beta-lactamases in resistant organisms. By removing plasmids that contain resistance, CRISPR can also reduce horizontal gene transfer between bacteria.

Additionally, a new class of antimicrobials is being created using CRISPR technology. By using sequence-specific targeting, CRISPR-based antimicrobials eliminate just the harmful bacteria while leaving the beneficial microbiota unharmed. This precise method lowers the danger of dysbiosis and prevents the side effects linked to broad-spectrum antibiotics (101,102). In environments like the gut microbiome, where microbial equilibrium is essential, such focused killing can be very beneficial.

CRISPR has the ability to fight important resistant infections, according to lab tests. Notably,

the *mecA* gene in MRSA, which confers methicillin resistance, has been knocked out using CRISPR-Cas9. As a result, beta-lactam antibiotics were once again effective against MRSA strains (103). In different research, the incidence of resistant bacteria in treated populations was considerably decreased by using CRISPR-Cas3 to break down plasmids containing several resistance genes in *E. coli*.

These discoveries are currently being translated into clinical practice. CRISPR's potential to alter the microbiomes of individuals who have recurring infections because of multidrug-resistant organisms is being investigated in trials. These treatments seek to prevent infections and enhance patient outcomes by specifically eradicating resistant strains (104). Furthermore, in order to enable prompt and accurate antimicrobial treatment, CRISPR-based diagnostics are being developed to quickly detect resistance genes in clinical samples (105).

The promise of CRISPR is enormous, despite several obstacles in its application to clinical practice, like as delivery methods, possible off-target effects, and regulatory barriers. To fully fulfil CRISPR's potential as a pillar of the worldwide campaign against AMR, further research and funding are needed (106).

Future Directions and Research in AMR

Antimicrobial resistance (AMR) is a growing threat, but the future of its management is promising due to advances in genomic technologies. Next-generation sequencing (NGS) is now faster and cheaper, enabling detailed detection of resistance genes and mutations (107). Innovations like single-cell sequencing offer insights into bacterial heterogeneity, while metagenomics helps map resistomes across various ecosystems. Artificial intelligence (AI) tools are being integrated with bioinformatics to interpret large-scale genomic data and develop targeted interventions (108). CRISPR systems, including Cas12 and Cas13, are expanding the toolkit for precise DNA and RNA editing (109). Base editing and gene drives represent novel applications that can reduce resistant strains or spread susceptibility traits among pathogens (110). Synthetic biology and CRISPR combine to enable engineered microbes to produce targeted antimicrobials (111). Although promising, these technologies raise ecological and safety concerns, such as potential off-target effects and gene transfer (112). Long-term studies are essential to evaluate their impact. The complexity of resistance mechanisms, from biofilm formation to efflux

pumps, necessitates further research into microbial interactions and genetics (113). The role of the microbiome in resistance spread also warrants deeper investigation (114). The development of new antimicrobials is still a top priority. Natural compounds combined with AI-driven screening provide a route to novel treatments (115). Innovative AMR therapies and diagnostics may be informed by cross-disciplinary insights from fields such as immunology and oncology (116).

II. Conclusion

Beyond the conventional development of antibiotics, the worldwide antimicrobial resistance challenge necessitates a swift and concerted response. Fighting resistant infections requires a multipronged strategy that includes genomics, quick diagnostics, innovative treatments, and precise genome editing using CRISPR. Advances in diagnostics, including WGS and PCR, facilitate timely and accurate treatment decisions. Innovative therapies such as phage therapy, adjuvant-enhanced antibiotics, and nanocarrier-based drug delivery offer promising alternatives to conventional antibiotics. CRISPR technology, with its ability to disrupt resistance genes and selectively eliminate resistant strains, represents a transformative tool in AMR management. Looking ahead, integrating artificial intelligence, synthetic biology, and in order to stop the development of AMR, tailored treatment combined with current tactics will be essential. To guarantee safe implementation, however, ethical, environmental, and regulatory issues need to be resolved. To protect present and future generations from the rising threat of antibiotic resistance, an international, multidisciplinary effort is required.

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