

Analytical Method Development Validation and Degradation of Inosine Pranobex by Using RP-HPLC Method.

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ABSTRACT-

This study focuses on the development, validation, and degradation assessment of an analytical method for inosine pranobex using Reverse Phase High-Performance Liquid Chromatography (RP-HPLC). The objective was to establish a reliable RP-HPLC method for the accurate quantification of inosine pranobex in pharmaceutical formulations and to evaluate its stability under various stress conditions.

Method Development: The RP-HPLC method was optimized using a C18 column with a mobile phase composed of Methanol (with 0.1% Water) and OPA (45%).

The flow rate was set at 0.8 mL/min, and detection was performed at 259 nm. Sample preparation involved dissolving inosine pranobex in methanol, followed by filtration.

Method Validation: The developed method was validated for specificity, linearity, precision, accuracy, limit of detection (LOD), and limit of quantitation (LOQ). Specificity tests confirmed no significant interference from excipients or degradation products. The calibration curve demonstrated linearity over the concentration range of 10-50 µg/mL. Precision studies showed acceptable repeatability and reproducibility. Accuracy was confirmed through recovery studies, with results within acceptable ranges. The LOD and LOQ were determined to be 0.16 µg/mL and 0.49 µg/mL, respectively.

Degradation Studies: Stability of inosine pranobex was assessed under acidic, alkaline, oxidative, photolytic, and thermal conditions. Degradation products were identified and quantified using the RP-HPLC method. Significant degradation was observed under acidic and oxidative conditions, with multiple degradation products formed.

Conclusion: The RP-HPLC method developed is robust, accurate, and precise for the analysis of

inosine pranobex. It is suitable for routine quality control

and stability testing. The degradation studies provide valuable insights into the stability profile of inosine pranobex, ensuring its effective application in pharmaceutical formulations.

Keywords: Inosine pranobex, RP-HPLC, method development, validation, degradation studies, stability analysis.

I. INTRODUCTION-

Inosine pranobex, also known as inosine pranobex or inosine acedobendimepranol, is a pharmaceutical agent used primarily for its immunomodulatory effects. It is often utilized in the treatment of viral infections and certain chronic conditions due to its ability to enhance the immune response. Given its clinical significance, it is crucial to have accurate and reliable methods for the quantification and quality control of inosine pranobex in pharmaceutical formulations.

Reverse Phase High-Performance Liquid Chromatography (RP-HPLC) is a widely employed analytical technique known for its high resolution, precision, and accuracy. This method is particularly effective in separating and quantifying complex mixtures of compounds, making it suitable for analyzing pharmaceutical products. The development and validation of an RP-HPLC method for inosine pranobex are essential for ensuring the consistency, safety, and efficacy of its pharmaceutical preparations.

Objective

The primary objective of this study was to develop and validate an RP-HPLC method for the determination of inosine pranobex. The method needed to be capable of accurately quantifying inosine pranobex in its dosage forms and assessing its stability under various stress conditions.

Validation was essential to confirm that the method met regulatory requirements and was suitable for routine quality control.

Importance of Method Development and Validation

- 1. Accuracy and Precision:** An accurate and precise RP-HPLC method is critical for ensuring that the concentration of inosine pranobex in pharmaceutical formulations is within the specified limits, thereby guaranteeing the therapeutic efficacy of the drug.
- 2. Specificity:** The method must be specific to inosine pranobex, effectively distinguishing it from other compounds and potential impurities or degradation products.
- 3. Robustness:** The method should be robust enough to withstand variations in operating conditions, ensuring consistent performance under different conditions.
- 4. Regulatory Compliance:** Validation ensures that the method complies with regulatory standards, such as those set by the International Conference on Harmonisation (ICH) and the United States Pharmacopeia (USP).

Degradation Studies

Understanding the degradation profile of inosine pranobex is crucial for evaluating its stability and shelf life. Degradation studies simulate various stress conditions, such as acidic, alkaline, oxidative, photolytic, and thermal environments, to determine how the drug degrades over time and under different conditions. These studies provide insights into the stability of the drug and the formation of degradation products, which are essential for ensuring drug safety and efficacy.

Significance of the Study

This study's significance lies in its contribution to the development of a reliable analytical tool for inosine pranobex. The RP-HPLC method established will serve as a critical quality control measure in pharmaceutical manufacturing and stability testing, ensuring that inosine pranobex products maintain their quality throughout their shelf life. Furthermore, understanding the degradation behavior of inosine pranobex aids in optimizing formulation conditions and storage practices to enhance product stability and patient safety.

Drug Profile: Inosine Pranobex

1. General Information:

- **Chemical Name:** Inosine pranobex
- **Chemical Structure:** Inosine pranobex is a combination of inosine, a purine nucleoside, and the complex compound acedoben dimepranol. The chemical formula of inosine pranobex is typically represented as $C_{16}H_{20}N_4O_6$.
- **Molecular Weight:** Approximately 316.36 g/mol.
- **Synonyms:** Inosine acedoben dimepranol, inosine pranobex, immuno-inosine.

2. Pharmacological Classification:

- **Class:** Immunomodulator, antiviral agent.
- **Mechanism of Action:** Inosine pranobex is believed to enhance the immune response by stimulating the activity of T-cells and other components of the immune system. It also has potential antiviral effects, improving the host's ability to combat viral infections.

3. Therapeutic Uses:

- **Primary Indications:**
- **Viral Infections:** Used in the treatment of viral infections such as herpes simplex, influenza, and other viral conditions.
- **Chronic Conditions:** Sometimes used to manage chronic infections or conditions that involve immune system dysfunction.

Off-

Label Uses: Occasionally used in other immunological conditions or chronic diseases based on clinical experience and physician discretion.

4. Dosage Forms and Administration:

- **Dosage Forms:** Available in various formulations including tablets, oral powders, and injectables.
- **Administration Routes:** Typically administered orally in tablet form, but may also be available as an injectable form depending on the indication and formulation.
- **Common Dosage Regimens:**
- **Adults:** Often prescribed in doses ranging from 1 to 2 grams daily, divided into multiple doses.
- **Children:** Dosage is usually adjusted based on body weight and specific condition.

5. Pharmacokinetics:

- **Absorption:** Inosine pranobex is absorbed from the gastrointestinal tract, though the extent of absorption can be affected by food and formulation.

- **Distribution:** Once absorbed, it distributes throughout the body, including tissues and fluids where it exerts its therapeutic effects.
- **Metabolism:** Inosine pranobex is metabolized primarily in the liver, with its active components being processed and broken down.
- **Excretion:** Excreted through urine, primarily as metabolites.

6. Side Effects and Adverse Reactions:

- **Common Side Effects:**
- Gastrointestinal issues such as nausea, vomiting, or diarrhea.
- Mild headache or dizziness.

Serious Adverse Reactions:

- Allergic reactions including rash, itching, or swelling.
- Rarely, severe liver toxicity or hematological abnormalities.

7. Contraindications and Precautions:

- **Contraindications:**
- Hypersensitivity to inosine pranobex or any of its components.
- Severe liver dysfunction or pre-existing liver disease.

Precautions:

- Use with caution in patients with pre-existing kidney or liver conditions.
- Monitoring may be required for patients with a history of allergic reactions.

8. Drug Interactions:

- **Potential Interactions:**
- May interact with other immunosuppressive drugs or antiviral agents.
- Potential interactions with drugs affecting liver metabolism.

9. Stability and Storage:

- **Storage Conditions:**
- Typically stored at room temperature, away from light and moisture.
- Tablets should be kept in tightly closed containers to prevent moisture absorption.

Shelf Life:

- Usually has a shelf life of 2-3 years from the date of manufacture when stored under recommended conditions.

10. Regulatory Status:

- **Approval:** Inosine pranobex is approved for use in several countries, including various European nations and others, for the treatment of specific viral infections and immunological conditions.
- **Regulatory Classification:** It is classified as a prescription medication, and its use is regulated by national and international health authorities.

11. Summary:

Inosine pranobex is a notable immunomodulatory agent with applications in treating viral infections and managing immune system-related conditions. Its efficacy is supported by its ability to enhance immune responses and combat viral activity. Proper formulation, administration, and monitoring are essential to maximizing therapeutic benefits while minimizing potential side effects.

Materials and Methods

1. Materials:

1. Drug Substance:

- **Inosine Pranobex:** Pure reference standard obtained from Sun Pharmaceutical Industries Pvt. Ltd

Chemicals and Reagents:

- **Mobile Phase Components:**
- **Water:** High-purity grade, typically deionized or distilled.
- **Acetonitrile:** HPLC grade.
- **Ammonium Phosphate Buffer:** For pH adjustment, analytical grade.
- **Sample Solvents:**
- **Methanol:** HPLC grade, for sample dissolution.
- **Dimethyl Sulfoxide (DMSO):** If needed, HPLC grade.

Standards and Impurities:

- **Inosine Standard:** For method validation and calibration.

Equipment:

- **HPLC System:** Equipped with a UV detector, autosampler, and suitable RP-HPLC column (C18, 250 mm x 4.6 mm, 5 μ m).
- **Pipettes and Micropipettes:** For accurate measurement of reagents and samples.
- **Filtration Apparatus:** 0.45 μ m filters for sample and solution filtration.
- **pH Meter:** For adjusting and monitoring pH of mobile phases and sample solutions.

Software:

- **HPLC Data Analysis Software:** For peak integration and quantification.
- 2. Methods:**
- 1. Method Development**
- 1. Column Selection and Optimization:**
- Use a C18 column (250 mm x 4.6 mm, 5 μ m) for the RP-HPLC analysis. Optimize the column temperature (commonly at 30°C) for best performance.
- Mobile Phase Preparation:**
- **Mobile Phase A:** Prepare an aqueous phase by adding 0.1% phosphoric acid to deionized water.
 - **Mobile Phase B:** Prepare by using acetonitrile.
 - **Gradient Profile:** Develop a gradient elution method starting with 90% A and 10% B, transitioning to 50% A and 50% B over 20 minutes. Adjust as necessary for optimal separation.
- Detection Wavelength:**
- Set the UV detector at 259 nm for monitoring inosine pranobex.
- Sample Preparation:**
- **Stock Solution:** Dissolve an accurately weighed amount of inosine pranobex in methanol or DMSO to prepare a stock solution (e.g., 1 mg/mL). Filter through a 0.45 μ m filter.
 - **Standard Solution:** Prepare a series of standard solutions at different concentrations (e.g., 0.5 μ g/mL to 50 μ g/mL) from the stock solution for calibration.
- 2. Method Validation**
- 1. Specificity:**
- Inject standard solutions, placebo (excipients without drug), and spiked samples to confirm no interference with inosine pranobex peak.
- Linearity:**
- Plot the calibration curve using the standard solutions. Assess linearity by calculating the correlation coefficient (r^2) of the calibration curve.
- Precision:**
- **Repeatability:** Analyze multiple injections ($n=6$) of the same sample to determine intra-day precision.
 - **Intermediate Precision:** Perform the analysis on different days and/or by different analysts to evaluate inter-day precision.
- Accuracy:**
- Perform recovery studies by spiking known amounts of inosine pranobex into the placebo matrix. Calculate the percentage recovery and ensure it falls within the acceptable range (typically 98-102%).
- Limit of Detection (LOD) and Limit of Quantitation (LOQ):**
- Determine LOD and LOQ based on the standard deviation of the response and the slope of the calibration curve. LOD is calculated as $3.3 \times \sigma/S$ and LOQ as $10 \times \sigma/S$, where σ is the standard deviation and S is the slope.
- Robustness:**
- Test the method by varying parameters such as flow rate (± 0.1 mL/min), column temperature ($\pm 5^\circ\text{C}$), and pH of the mobile phase (± 0.2 units) to assess the impact on method performance.
- 3. Degradation Studies**
- 1. Acidic Degradation:**
- Expose a solution of inosine pranobex to hydrochloric acid (0.1 N) at 60°C for 2 hours. Neutralize, dilute, and analyze by RP-HPLC.
- Alkaline Degradation:**
- Expose the drug solution to sodium hydroxide (0.1 N) at 60°C for 2 hours. Neutralize, dilute, and analyze.
- Oxidative Degradation:**
- Treat the drug with hydrogen peroxide (3%) at room temperature for 2 hours. Analyze after dilution.
- Photolytic Degradation:**
- Expose the drug to UV light (365 nm) for 24 hours. Analyze the sample before and after exposure.
- Thermal Degradation:**
- Store the drug at elevated temperatures (e.g., 60°C) for 1 month. Analyze samples periodically.
- Analysis of Degradation Products:**
- Identify and quantify degradation products by comparing retention times and UV spectra with standards and using the developed RP-HPLC method.
- 4. Data Analysis:**
- Use HPLC software for data acquisition, peak integration, and quantification. Validate results by comparing them to known standards and evaluating method performance parameters such as resolution, peak symmetry, and retention time.

5. Documentation:

- Document all procedures, results, and observations meticulously. Include calibration curves, validation data, degradation study results, and any deviations from standard procedures.

This comprehensive approach ensures that the RP-HPLC method for inosine pranobex is robust, accurate, and suitable for routine analysis and stability testing.

6. MATERIAL AND METHODS

1. Selection and Procurement of Drug **Drug sample supplier**

Name of Drug	Drug Supplier
Inosine Pranobex	Sun pharmaceutical pvt. Ltd

Table 3: Drug and Drug Supplier

List of reagents & chemicals used

Table 4: List of Reagents and Chemicals used

Sr.No.	Name of chemicals	Manufacturer.
1.	Acetonitrile (HPLC grade)	Merck Ltd., India
2.	Methanol (HPLC grade)	Merck Ltd., India
3.	0.1% OPA (HPLC grade)	Loba Chemie
4.	Ammonium phosphate Buffer	Avantor performance material India Ltd
5.	water (HPLC grade)	Merck specialities private limited, Mumbai

2. Selection of formulation:

Marketed Preparation:

Table No. 5: List of brand names of combined formulations of Inosine Pranobex

Sr.No	Brand name	Formulation	Available strength	Address of manufacturer
1.	Viralex	Tablet	Inosine Pranobex 500 mg	Themis medicare ltd

The marketed preparation was obtained from local market and is referred here after in this thesis by the name as such.

3. Selection of Analytical Technique

HPLC was selected as an analytical technique for estimation of Inosine Pranobex.

• Instruments:

The analysis of the drug was carried out on Agilent Technology Gradient System with Auto injector, (DAD) & Gradient Detector. Equipped with Reverse Phase (Agilent) C18 column (4.6mm x 250mm; 5µm), a 20µl injection loop and DAD Absorbance detector and running chemstation 10.1 software.

• Stock preparations:

- Stock I: Standard Sample Preparation

Std. Inosine Pranobex

10mg in 10ml Methanol = 100µg/ml

- Stock II: Formulation solution Preparation:-

Take 13.54ml in 10ml Methanol i.e. = 100µg/ml

- For Accuracy Solution Preparations:-

- Take 10µg/ml FORMULATION FOR ACCURACY,

80% = 0.1ML FORMULATION and ADD 0.8ml STD Inosine Pranobex AND MAKE UP VOL 10 ML WITH Mobile Phase

100% = 0.1ML FORMULATION and ADD 0.1ml STD Inosine Pranobex AND MAKE UP VOL 10 ML WITH Mobile Phase

120% = 0.1ML FORMULATION and ADD 0.12ml STD Inosine Pranobex AND MAKE UP VOL 10 ML WITH Mobile Phase

4. Instruments and Equipments

Name of Instrument	Company Name
1 HPLC Instrument	Agilent Technology Gradient System with Auto injector (chemstation software 10.1)
2 UV-Spectrophotometer	Analytical Technologies Limited
3 Column (C18)	Agilent C18 (250mm X 4.6mm, 5µm)
4 pH meter	VSI pH meter (VSI1-B)
5 Balance	WENSAR™ High Resolution Balance.
6 Sonicator	Ultrasonic electronic instrument

Table No.6: Instrument (HPLC) Details used during Method Development

7. EXPERIMENTAL WORK

7.1. Preliminary characterization

Preliminary characterization involved determining colour, odour, and appearance, which are needed for Inosine pranobex analysis.

The following criteria were used to analyze common drugs.

A. Melting point

The purity and standard of a drug can be determined by the melting point. It is determined with

the aid of digital melting point equipment. The optimized Melting point of inosine pranobex is 143 compares to the standard is 140-145 in range.

B. Solubility Studies

This study was carried out to find an ideal solvent in which drugs are completely soluble. Various solvents were tried for checking solubility of Inosine pranobex. From solubility studies it was concluded that of Inosine pranobex is freely soluble in Methanol and poorly soluble in water.

C. UV-Spectrophotometer

UV Spectrophotometer was used to determine the λ_{max} and it was compared to the standard value acquired from books and research articles.

1. Preparation of standard stock solution

For the preparation of stock solution, weighed 10mg of drug Inosine pranobex transferred into 10ml volumetric flask, added 10 ml of methanol and sonicate to dissolve the standard completely and make up the volume with methanol (1000 μ g/ml). Further diluted from 0.1ml to 10 ml methanol (10 μ g/ml).

2. Selection of analytical wavelength

Methanol as a blank and drug standard solution (10 μ g/ml) was scanned from 400nm to 200nm & absorption maxima were determined for drug Inosine pranobex showed maximum absorbance at 259 nm. (Fig No:7).

3. HPLC:

1. Selection of Analytical Technique

HPLC was selected as an analytical technique for estimation of Inosine pranobex.

a. Instruments:

The analysis of the drug was carried out on Agilent Tech Gradient System with Auto injector DAD Detector Equipped with Reverse Phase C18 (Agilent) with 250mm x 4.6 (5 μ m), a SP930D pump, a 20 μ l injection

loop and DAD Absorbance detector and running chemstation software.

b. Selection of stationary phase:

The column used in this method C18 Agilent The configuration of the column is 4.6x250mm, particle size 5 μ m C18 column gives high non-polar retentive, symmetric peak shape, highly reproducible and stable ideal for HPLC method.

c. Mobile phase selection:

In order to produce a sharp peak with a spectrum the mobile phases of the Inosine pranobex were mixed at varying ratios and flow rate. Water (Ammonium Ammonium Phosphate Buffer PH-7.4 with 0.1% OPA), methanol, and other solvents consisting of one or more of the different mobile phases. Different ratios and Mobile phase were examined, but a suitable peak was produced by combining methanol and water (10mM Ammonium Ammonium Phosphate Buffer PH-7.4 with 0.1% OPA) as a solvent.

d. Preparation of Stock Solution: 10 mg of Inosine pranobex were dissolved in 10ml of methanol which gives concentration of stock solution was 1000 μ g/ml respectively.

e. Preparation of Standard Solution: Pipette out 0.2ml from stock solution and dilute with 9ml methanol & 1ml water to give 10ml solution.

7.4 METHOD DEVELOPMENT OF HPLC:

Following Mobile phase were tried

Table No.8: Selection of mobile Phase

Sr.No	MobilePhase
1.	Methanol+Buffer(90+10), 259nm, 20Mg, Flowrate 0.7ml/min.
2.	Methanol+Buffer(80+20), 259nm, Flowrate 0.7ml/min.
3.	Methanol+Buffer(70+30), 259nm, Flowrate 0.7ml/min

4.	Methanol+Buffer(60+40% v/v),259nm,Flowrate0.7ml/min
5	Methanol+Buffer(55+45% v/v),259nm,Flowrate0.7ml/min.
6	Methanol+10mmAmmoniumPhosphateBufferpH-7.4with0.1%OPA(45+55% v/v),259nm,andFlowrate0.8 ml/min.

7.4 Study on the chromatographic conditions of Inosine pranobex:

Accurately weigh and transfer 10mg Inosine pranobex working standard into 10ml volumetric flask as about dilute Methanol prepared in completely and make volume up to the mark with the same solvent to get 1000µg/ml standard (stock solution) and 15 min sonicate to dissolve it and from the resulting solution 0.1-0.5ml was transferred to 10 ml volumetric flask and the volume was made up to the mark with mobile phase Methanol:(10mM Ammonium Ammonium Phosphate Buffer pH adjusted 7.4 with OPA) Water solvent. The resulting 10-50µg/ml of solution was subjected to chromatographic analysis using mobile phases of different strengths with chromatographic conditions mentioned below: **(Table No: 11).**

Analytical column

: Agilent C18 Column (250mm x 4.6mm), 5µm particle size. Injection volume : 20µl

Flow rate: 0.8ml/min

Detection: 259nm

Run Time: 10min

1. Preparation of standard stock solution:-

• Inosine pranobex standard stock solution: (Stock I)

An accurately weighed quantity, 10 mg of Inosine Pranobex (ISP) was dissolved in Methanol in a 10 ml volumetric flask and volume made up to 10.0 ml to produce a solution of 1000 µg/ml.

a) Take 0.1-0.5 ml from stock solution and volume make up with mobile phase (4.5ml Methanol + 5.5ml Buffer). Solution contains 10-50 µg/ml of Inosine pranobex.

8. Studies of Calibration plot:-

1. Optimization of Chromatographic condition:

The following chromatographic conditions were established by trial and error and were kept constant throughout the analysis.

Analytical column

: Agilent C18 Column (250mm x 4.6mm) particle size: 5µm

Injection volume : 20µl

Flow rate: 0.8ml/min

Detection: 259nm

Run Time: 15min

Mobile phase: Methanol:(10mM Ammonium Phosphate Buffer pH adjusted 7.4 with OPA)(45%:55%)

9. Procedure for calibration curve of Inosine pranobex:

The mobile phase was allowed to equilibrate with stationary phase until steady baseline was obtained. From the freshly prepared standard stock solution, pipette out 10 mg Inosine pranobex in 10ml of volumetric flask and dilute with mobile phase. From it 0.1, 0.2, 0.3, 0.4 and 0.5ml of solution were pipette out in 10 ml volumetric flask and volume was made up to 10 ml with mobile phase to get final concentration 10, 20, 30, 40, 50 µg/ml of Inosine pranobex. Sample were injected and peaks were recorded at 259 nm as the graph plotted as concentration of drug versus peak areas depicted in **(fig. no.15)** respectively.

10. Study of system suitability parameters:

The system suitability is used to verify, whether the resolution and reproducibility of the chromatographic system are adequate for analysis to be done. The test was performed

bycollectingdata from tworeplicateinjections of standard solution.

1. CalibrationExperiment:

• **RP-HPLCMethod:**

a. PreparationofCalibrationcurvestandard:

The above standard stocksolution(1000µg/ml) of Inosine pranobexwasdiluted withmobilephaseto yieldFivecalibrationcurve(cc)standardswithconcentrationsof10,20,30,40,50 µg/ml of Inosine pranobex. The calibration curve of Inosine pranobex isdepictedin (FigNo.15).

b. SelectionofdetectionWavelength:

Standard solutions were scanned in the range of 200-400nm ,against 10 ml Methanol andvolumemakewithwatersolventsystemasreferenceInosinepranobexwereshowedabsorbancemaxima (lamdamax)at 259 nm(FigureNo:7).

c. Calibrationstandarddrugandregression equationdata:

FromthestandardstocksolutionofInosinepranobex,differentconcentrationwereprepared respectively in the range of 10-50 µg/ml for Inosine pranobex (Figure No:18) andmeasuredat259nm.The calibrationcurveswere plottedun Regressionequationdatapresentedin (Table No:22).

d. Calibrationrunsandregression analysis:

Thesecalibrationstandardsolutionswereanalyzedinth reeplicatesusingtheundermentionedchromatographic conditions.

Analyticalcolumn

:C18Column(250mmx4.6mm,5µmpartical size).Injectionvolume :20µl.

Flowrate:0.8ml/min.

Mobilephase

:Methanol:Water(10mMAmmoniumPhosphateBufferpHadjusted7.4

withOPA)(45:55%V/V).

Detection :259nm.

11. ValidationofmethodforanalysisofInosine pranobex:

• ThedevelopedmethodwasvalidatedasperICHguidelines.

1. Linearity:

Linearity of an analytical method is its ability to elicit test results that are directly or by a well defined mathematical transformation, proportional to the concentration of analyte insampleswithin agiven range, TheResult areshown in;(Table No:9).

Determination:

The linearity of the analytical method is determined by mathematical treatment of testresults obtained by analysis of samples with analyte concentrations across the claimedrange. Area is plotted graphically as a function of analyte concentration. (Fig No. 19-29)Percentage curve fittings are calculated. The Result are shown in; (Table No.34 and TableNo.35).

AcceptanceCriteria:

Theplotshouldbelinearpassingthroughtheorigin.

CorrelationCoefficient

shouldnotbelessthan0.999.TheResult areshownin;

Preparationofstandardstocksolutionforlinearity:

10mgofInosinepranobexwereweighedandtransferred to10mLvolumetricflask&diluentwasaddedtomakeup thevolume.Sonicatedfor10minwithoccasionalswirling.

0.1mlofthissolutiondilutedupto10mlvolumetric flaskwithdiluentwasaddedtomakeup the volume.

Preparationoflinearitysolution:

Aseriesofstandardpreparationsofworkingstandardof wereprepared.

TableNo.9:TableoflinearityforRp-HPLCMethod

LinearityofInosinepranobex HPLC	
Sr.No.	Concentratin(µg/mL)
1	10
2	20
3	30
4	40
5	50

2. Accuracy(recovery):

The accuracy of an analytical method is the closeness of test results obtained by that method to the true value. Accuracy may often be expressed as percent recovery by the assay of known added amounts of analyte. The accuracy of an analytical method is determined by applying the method to analyzed samples, to which known amounts of analyte have been added. The accuracy is calculated from the test results as the percentage of analyte recovered by the assay. The RP-HPLC Result are shown in; (Table No:39,40).

Acceptance Criteria:

Mean recovery should be in the range of 98-102%.

The Relative Standard Deviations should not be more than 2.0%.

Preparation of standard stock solution:

10 mg of Inosine pranobex working standards were weighed and transferred to 10 mL volumetric flask & diluent was added to make up the volume 0.2 ml of this solution diluted up to 10 ml with diluent.

Application of proposed method for analysis of tablet formulation: Accuracy

The accuracy was determined by Inosine pranobex (equivalent to 10 mg (80 %, 100 % and 120 % of the label claimed, respectively) to quantity equivalent to average weight of marketed tablets. This powder containing 10 mg of Inosine pranobex were triturated and then subjected to chromatographic analysis using the described method. The resulting was analyzed in triplicates over three days. The % recovery of added drug was taken as a measure of accuracy.

The Results are shown in; (Fig No:31,32,33).

Table No.10: Table of Accuracy for RP-HPLC Method

Sample	Amount Added (mg)
	Inosine pranobex
Accuracy 80%	8
Accuracy 100%	10
Accuracy 120%	12

3. Repeatability:

Precision of the system was determined with the sample of RP-HPLC for. Three replicates of sample solution containing 10 mg of Inosine pranobex were injected and peak areas were measured and %RSD was calculated. Was repeated for five times result are shown in; (Table No : 43) & (Fig No :34,35).

Application of proposed method for analysis of Repeatability:

10 mg was weighed and transferred to 10 mL volumetric flask & diluent was added to make up the volume. Sonicated for 10 min with occasional swirling. The above solution was filtered through 0.45µm membrane filter 0.5 ml of this solution diluted up to 10 ml with diluent.

4. Precision:

Precision of an analytical method is the degree of agreement among Individual test results when the procedure is applied repeatedly to multiple Samplings of a homogenous sample. Precision of an analytical method is usually expressed as standard deviation or relative standard deviation. Also, the results obtained were subjected to one way ANOVA and within-day means square and between-day means square was determined and compared using F-test. (Fig No:36).

Result of Intraday and Interday Precision studies on RP-HPLC method for Inosine pranobex

1. Intra-day precision:

Sample solutions containing 10 mg of Inosine pranobex three different concentration (10µg/ml, 30µg/ml, 50µg/ml Inosine pranobex were analyzed three times on the same day and %R.S.D

was calculated. The Result are shown in; (Table No.50)&(Fig No:36-38).

2. Inter-day precision:

Samplesolutionscontaining10mgofInosine pranobexthree different concentration (10µg/ml,30µg/ml,50µg/ml)inHPLCandthree different concentration in Inosine pranobex different days and % R.S.D was calculated. It is usually expressed as standard deviation or relative standard deviation. The Results are shown in; (Table No.50)&(Fig No:39-41).

- **Acceptance criteria:**

The Relative Standard Deviations should not be more than 2% for test

- **Preparation of standard stock solution:**

10 mg of Inosine pranobex working standards were weighed and transferred to 10 mL volumetric flask & diluent was added to make up the volume. 0.1 ml of this solution diluted up to 10 ml with diluent.

5. Robustness:

The mobile phase composition was changed in ($\pm 1 \text{ ml/ min}^{-1}$) proportion and the flow rate was (Fig No:42,43) of Methanol: Water (10mM Ammonium Phosphate Buffer pH adjusted 7.4 with OPA) in the mobile phase composition ($\pm 1 \text{ ml/ min}^{-1}$) and the change in detection wavelength ($\pm 1 \text{ ml/ min}^{-1}$) and the effect of the results were examined. (Fig No:44,45) and (Fig No:46,47) it was performed using 30µg/ml solution of Inosine pranobex in replicate. The Result are shown in; (Table No:57).

5. Detection Limit

Based on the S.D. of the response and the slope of calibration curve, the detection limit (DL) was calculated as,

$$DL = \frac{3.3\sigma}{S}$$

Where,

σ = the S.D. of the y-intercept of regression lines. S = the slope of the calibration curve.

The slope S may be estimated from the calibration curve and S.D. was used should be calculated from the y-intercept of regression line in calibration curve. The results are shown in; (chapter:8).

6. Quantitation Limit

Based on the S.D. of the response and the slope of calibration curve, the quantitation limit (QL) was calculated as,

$$QL = \frac{10\sigma}{S}$$

Where,

σ = the S.D. of the y-intercept of regression lines. S = the slope of the calibration curve.

The slope S may be estimated from the calibration curve and S.D. was used should be calculated from the y-intercept of regression line in calibration curve.

The results are shown in (chapter:8)

7.12 Analysis of marketed formulation

Weigh Equivalent weight of tablet 13.54 mg. add about 10 ml of diluent and sonicate to dissolve it completely and make volume up to the mark with diluent. Mix well and filter through 0.45 µm filter. Further pipette 0.3 ml of the above stock solution into a 10

ml volumetric flask and dilute up to the mark with diluent (30µg/ml). The simple chromatogram of test Inosine pranobex shown in (Fig No: 48) the amounts of Inosine pranobex per tablet were calculated by extrapolating the value of area from the calibration curve. Analysis procedure was repeated five times with tablet formulation. Tablet Assay for % Label claim for % RSD Calculated, Result was shown in (Table No. 59).

Brand Name: Viralex (500mg) [Themis Medicare Ltd]

Total weight of 20 tab Powder wt. = 13.54 gms

$$\frac{\text{Avg Powder Weight} \times 10 \times \text{Tab Eq. Weight}}{10 \times \text{Avg. Wt.}} = \frac{0.677 \times 10 \times 677}{10 \times 677} = 13.54 \text{ mg Label claim}$$

Take 13.54 mg tablet powder wt. in 10 ml MEOH = 1000 µg/ml

Ruggedness

The degree of reproducibility of test result obtains by the analysis of same sample under variety of Condition. Such as different analyst, laboratory Different instrument. Result was shown in (Table No. 59), (Fig No:48).

Forced degradation studies:

Stress degradation of the method was performed to measure the analyst response in the presence of its potential impurities. Stress testing of the individual drug substance and the combination was performed to measure the resolution factors of the drug peak from its nearest resolving peak and also from all other peaks. The drugs were subjected to acidic, alkaline, oxidizing and photolytic conditions. For acidic degradation, the drugs were subjected to 0.1 N hydrochloric acid for 2 hr. for the alkaline degradation the drugs treated with 0.1 N sodium hydroxide for 1 hr. Oxidative studies were carried out using 3% hydrogen peroxide for 2 hr. Thermal degradation the drug treated with 1 hr. Photodegradation was performed by exposing the solution to light for 24h. After the completion of the treatment, the solutions were left to return to room temperature, neutralized (for acidic and basic degradation) and diluted with the mobile phase to obtain the final concentration of 30 µg/mL.

Degradation behavior

Forced degradation studies of both the drugs namely Inosine pranobex were carried out individually and in under different stress conditions like acid hydrolysis, alkaline hydrolysis, hydrogen peroxide oxidation and photolysis. The results are shown in (Table no: 60).

1. Acid hydrolysis:

The acid hydrolysis performed using 0.1N HCl for 2 hr for Inosine pranobex indicated degradation 7.20 % and degradation was observed 2 hr. (Figure: 53).

2. Alkaline hydrolysis:

The alkaline hydrolysis condition was performed using 0.1N NaOH for 1 hr and 2

hr Inosine pranobex. Degradation of Inosine pranobex 2 hr was found to be 5.44%. (Figure: 50).

3. Oxidation:

In the oxidation condition with 3% H₂O₂ for 1 hr and 2 hr Inosine pranobex show any oxidative stress degradation peak in the chromatogram. Degradation of Inosine pranobex were observed 3.16% for 2 hr respectively (Figure no: 56).

4. Neutral:

There was no major degradation observed for Inosine pranobex and hence they were not sensitive to light for 2hr. (Figure no: 59).

II. RESULT AND DISCUSSION:

1. Preliminary studies on Inosine Pranobex

Table No. 7 Preliminary studies on Inosine Pranobex

x	
1 Appearance	Solid
2 Color	White to Offwhite Powder
3 Taste	Bitter Taste

1. Melting point

The procured reference standard of Inosine Pranobex was found to melt in the range of 140-145 °C.

2. Solubility

The drug was found to be

- Freely soluble in Methanol, Ethanol, DMSO.
- Practically insoluble in Ether.

8.1.3. UV Spectroscopy

Standard solutions were scanned in the range of 200-400nm, against 10 ml methanol and volume make with water solvent system as reference Inosine Pranobex in Methanol was found to be selected wavelength is 259 nm (Figure No: 07).

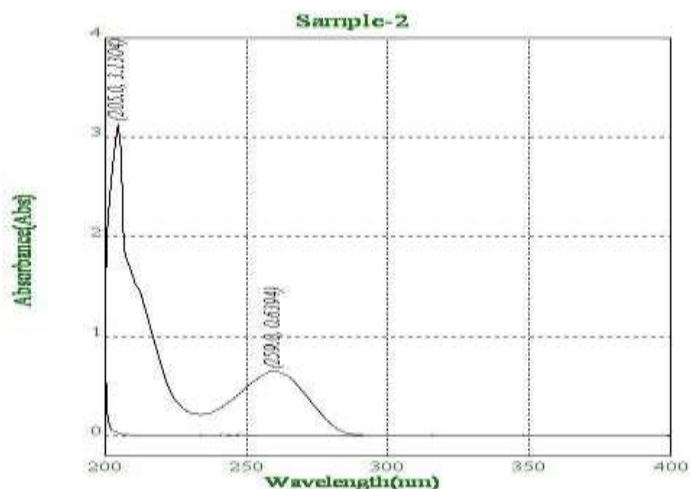


Fig.7:UVSpectrum of Inosine Pranobex

Table No.8: Chromatographic behavior of Inosine Pranobex mobile phase of various.

Fig No	Column used	Mobile phase, Flow Rate and Wavelength	Inj. Vol.	Observation	Conclusion
1.	C18 (Agilent) (4.6 mm x 250 mm), 5 μm)	90% Methanol + 10% Ammonium Phosphate buffer pH-7.4 with 0.1% OPA 10 Mcg, Flow rate 0.7 ml/min	20 μl	Sharp Peaks were not obtained	Hence rejected
2.	C18 (Agilent) (4.6 mm x 250 mm), 5 μm)	80% Methanol + 20% Ammonium Phosphate buffer (80+20) pH-7.4 with 0.1% OPA Water, Ph-3 259 nm 20 Mcg, Flow rate 0.7 ml/min	20 μl	Sharp Peaks were not obtained	Hence rejected
		70% Methanol +			

3.	C18(Agilent)(4.6mmx250mm), 5µm)	30% Ammonium Phosphate buffer (70+30), pH-7.4 with 0.1% OPA, 259nm, Flowrate 0.7ml/min	20µl	Sharp Peaks were not obtained	Hence rejected
4.	C18(Agilent)(4.6mmx250mm), 5.0µ)	60% Methanol + 40% Ammonium Phosphate buffer (60+40), pH-7.4	20µl	Sharp Peaks were not obtained	Hence rejected

		with 0.1% OPA 259nm, Flow rate 0.7ml/min.			
5	C18(Agilent)(4.6mmx250mm), 5µm)	Methanol + Ammonium Phosphate buffer pH-7.4 with 0.1% OPA (60+40% v/v), 259nm, Flowrate 0.7ml/min	20µl	Sharp Peaks were not obtained	Hence rejected

C18(Agilent)(4.6mmx250mm),5.0μ	Methanol + Ammonium Phosphate buffer pH-7.4 with 0.1% OPA (45+55% v/v), 259nm, Flowrate 0.8ml/min	20μl	Sharp and well resolved Peaks were obtained Hence selected
--------------------------------	---	------	--

Thus, from the above, it has been observed that using mobile phase of Methanol + Ammonium Phosphate buffer pH-7.4, with 0.1% OPA water

(45:55 % v/v) 259nm, Flowrate 0.8 ml/min gave adequate retention time is 5.555 min with good shape peak will be obtained.

Chromatogram of Trial 1:

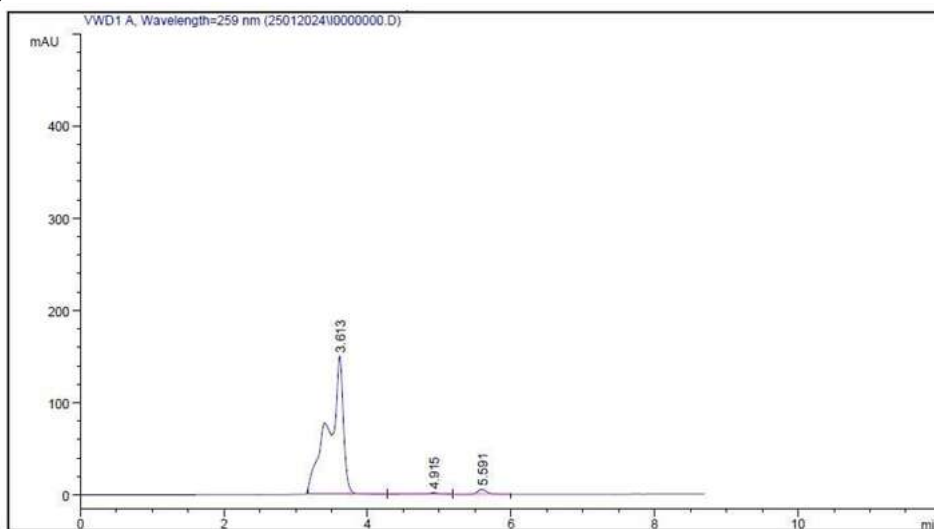


Fig.8: Chromatogram of Trial 1

No.	RT [min]	Area [mV*s]	TP	TF	Resolution
1	3.613	2170.05469	928	2.94	--
2	4.915	20.59516	9092	1.96	3.82
3	5.591	48.1525	10083	0.90	3.14

Table No 9 Result of chromatogram of trial 1

Observation: sharp peak were not obtained, so this method was rejected. Unsatisfactory result is obtained.

Chromatogram of Trial 2:

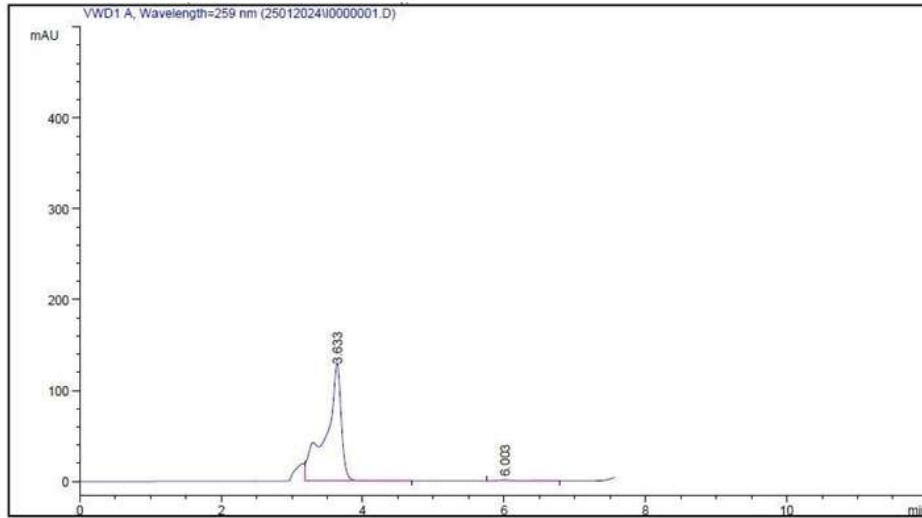


Fig.9:Chromatogram of Trial 2

No.	RT [min]	Area [mV*s]	TP	TF	Resolution
1	3.633	1969.28125	3031	2.37	--
2	6.003	14.44654	7783	0.40	8.83

Table No.10 Result of chromatogram of trial 2

Observation: sharp peak were not obtained, peaks splitting are observed, so this method was rejected. Unsatisfactory result is obtained.

Chromatogram of Trial 3:

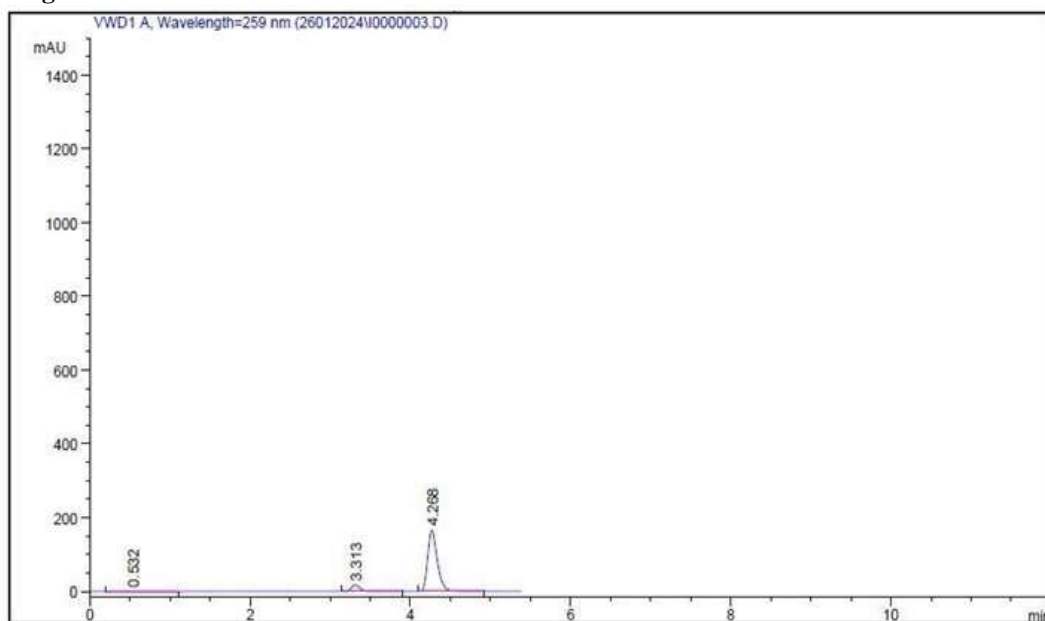


Fig.10:Chromatogram of Trial 3

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	0.532	29.02412	5308	0.71	--
2	3.313	152.35164	9261	2.31	2.29
3	4.268	120.81380	-	4.30	-

TableNo.11Resultofchromatogram oftrial 3

Observation:sharppeakwerenotobtained,peaktelling areobserved,sothismethodwasrejected. Unsatisfactory result is obtained.

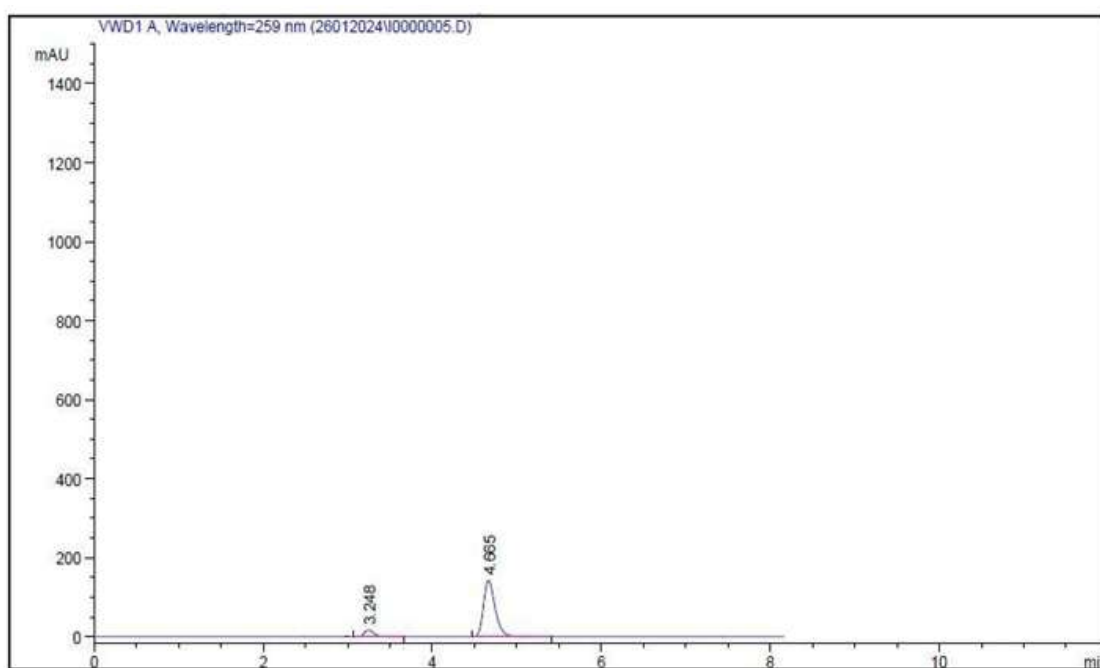


Fig.11:ChromatogramofTrial 4

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	3.248	137.13821	3969	0.79	-
2	4.665	1373.50842	5883	0.71	6.30

TableNo.12Resultofchromatogram oftrial 4

Observation:sharppeakwerenotobtained,peaksplittingareobserved,sothismethodwasrejected. Unsatisfactory result is obtained.

Chromatogram of Trial 5:

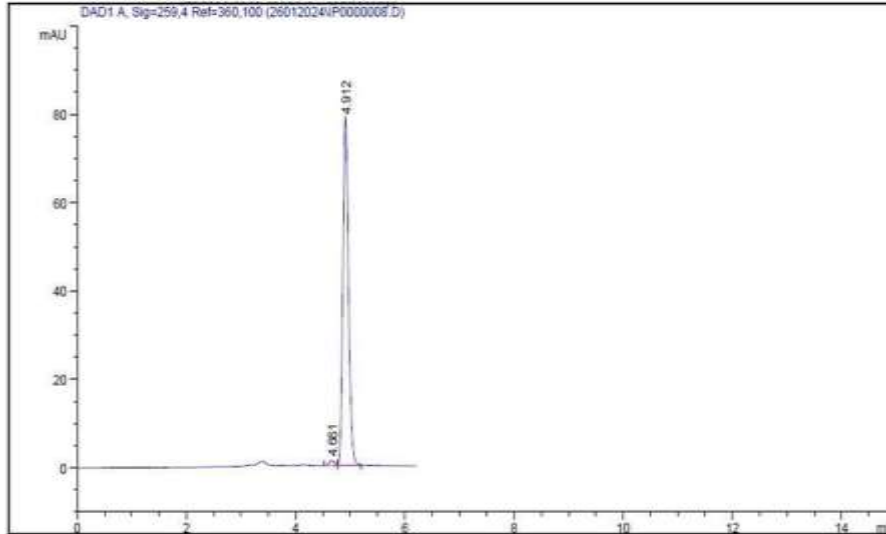


Fig.12: Chromatogram of Trial 5

No.	RT [min]	Area [mV*s]	TP	TF	Resolution
1	4.661	8.39332	9370	0.91	-
2	4.912	558.01556	11746	0.79	1.34

Table No.13 Result of chromatogram of trial 5

Observation: sharp peak were not obtained, peak splitting are observed, so this method was rejected. Unsatisfactory result is obtained.

Chromatogram of final Trial 6:

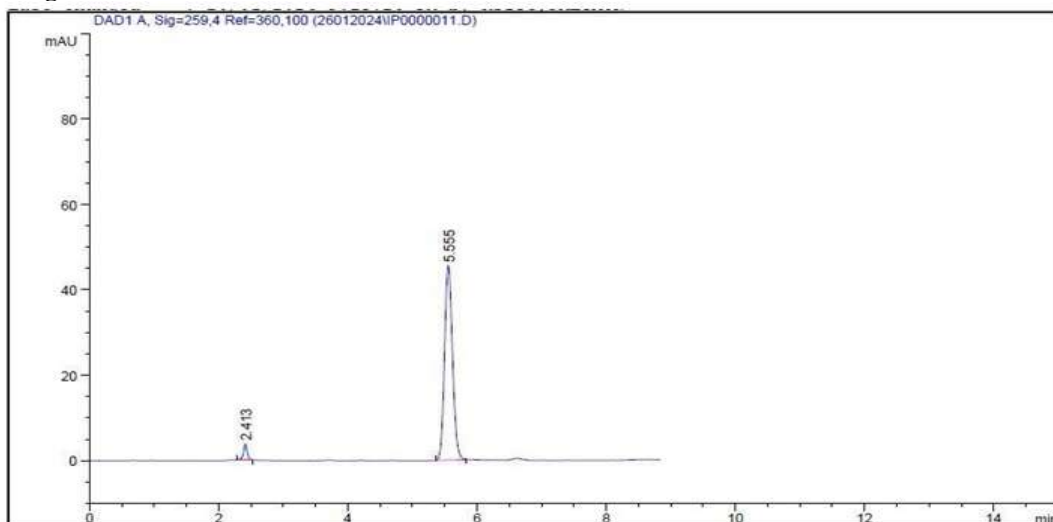


Fig.13: Chromatogram of Trial 6

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	2.413	16.44549	9216	1.03	--
2	5.555	391.08365	10115	0.85	19.51

TableNo.14Resultofchromatogram oftrial 6

Observation: Sharp peak was obtained so this method was selected.

• **The final chromatographic conditions selected were as follow:**

- **Analytical Column :** Agilent C18 Column (250mm x 4.6mm, 5µm particle size).
- **Injection volume :** 20µl.
- **Flow rate :** 0.8ml/min.

• **Mobile phase**

: Methanol: (Phosphate buffer pH-7.4 with 1% OPA) Water (45:55% V/V)

• **Detection wavelength :** 259nm.

• **Run Time :** 15min.

• **Preparation of Standard chromatogram of Inosine Pranobex**

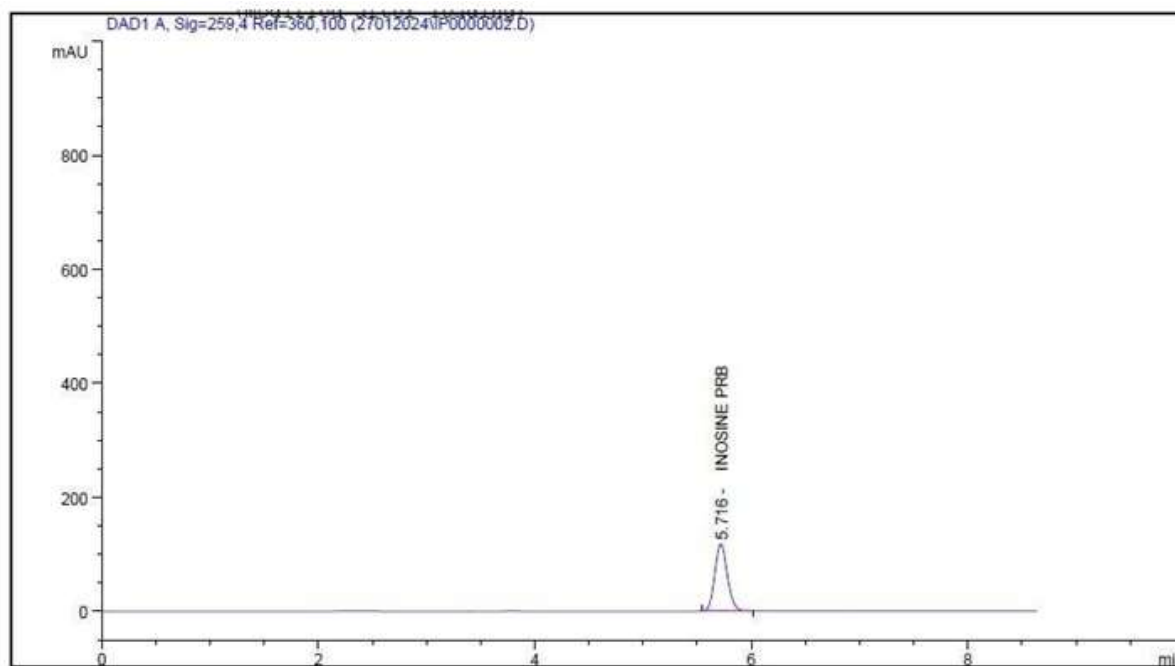


Fig.14: Chromatogram of standard Inosine Pranobex

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.716	956.42035	12031	0.84	--

Table.No15DetailsofchromatogramofstandardInosinePranobex

In the standard of Inosine Pranobex theoretical plates were found above 2000 i.e. for Inosine Pranobex is 12031 at minimum RT 5.716.

5. Calibration experiment

• RP-HPLC Method:

The data obtained in the calibration experiment when subjected to linear regression analysis showed

a linear relationship between peak areas and concentrations in the range 10-50 µg/ml for Inosine Pranobex (Table No: 16) depict the calibration data of Inosine Pranobex. The respective linear equation for Inosine Pranobex was $Y = 93.82x - 9.564$ where x is the concentration and y is the area of peak. The correlation coefficient was 0.999. The calibration curve of Inosine Pranobex is depicted in (Fig No. 15).

Method	Conc µg/ml	Peak area (µV.sec)		Average peak area (µV.sec)	S.D. of Peak Area	%RSD of Peak Area
		1	2			
RP-HPLC Method	10	956.0263	956.450	305.685	0.77	0.25
	20	1894.0279	1896.92	718.485	1.82	0.25
	30	2791.7861	2799.79	1140.14	2.88	0.25
	40	3746.9580	3767.88	1606.42	1.44	0.09
	50	4716.0210	4716.87	2036.725	6.26	0.31
Equation		Y=93.82x-9.564				
R ²		0.999				

Table No 16 Linearity data for Inosine Pranobex

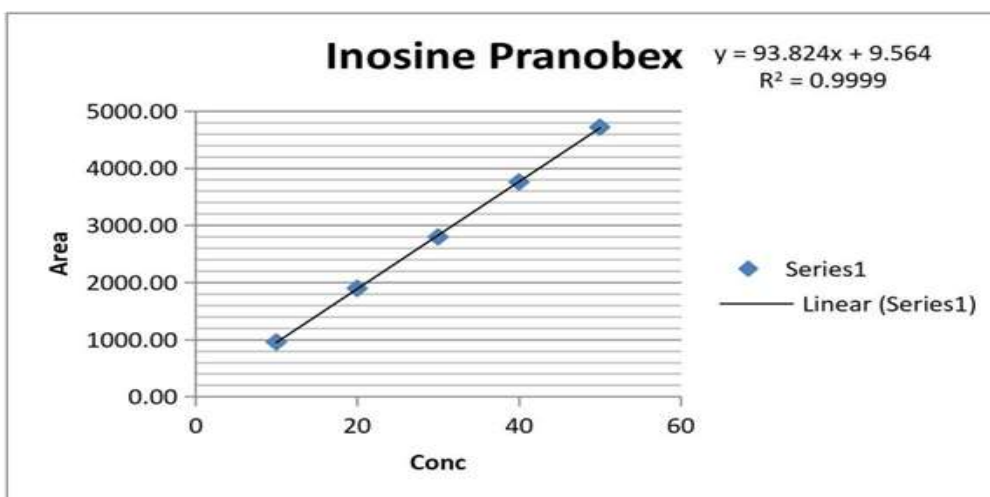


Fig.15: Calibration curve of Inosine Pranobex (HPLC)

The RP-HPLC Method for respective linear equation for Inosine Pranobex was $y = 93.8x - 9.564$ where x is the concentration and y is area of peak. The correlation coefficient was 0.999. The calibration curve of Inosine Pranobex is depicted in Fig 15.

2. Analytical of Method Validation:

1. Linearity:

From Inosine Pranobex standard stock solution, different working standard solution (10-50 µg/ml) were prepared in mobile phase 20 µl of sample solution was injected into the chromatographic system

using mixed volume loop injector Chromatograms were recorded. The areas for each concentration were recorded (Table No. 17) The Calibration curves are shown in [Fig. No. 16].

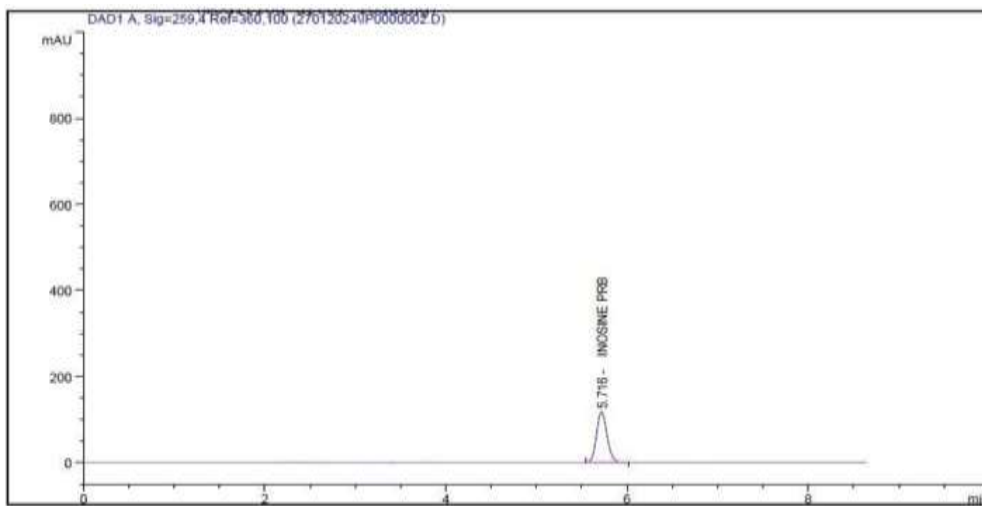


Fig.16:Chromatogram of linearity

No.	RT [min]	Area [mV*s]	TP	TF	Resolution
1	5.716	956.42035	12031	0.84	--

Table No. 17 Detail of chromatogram of Linearity

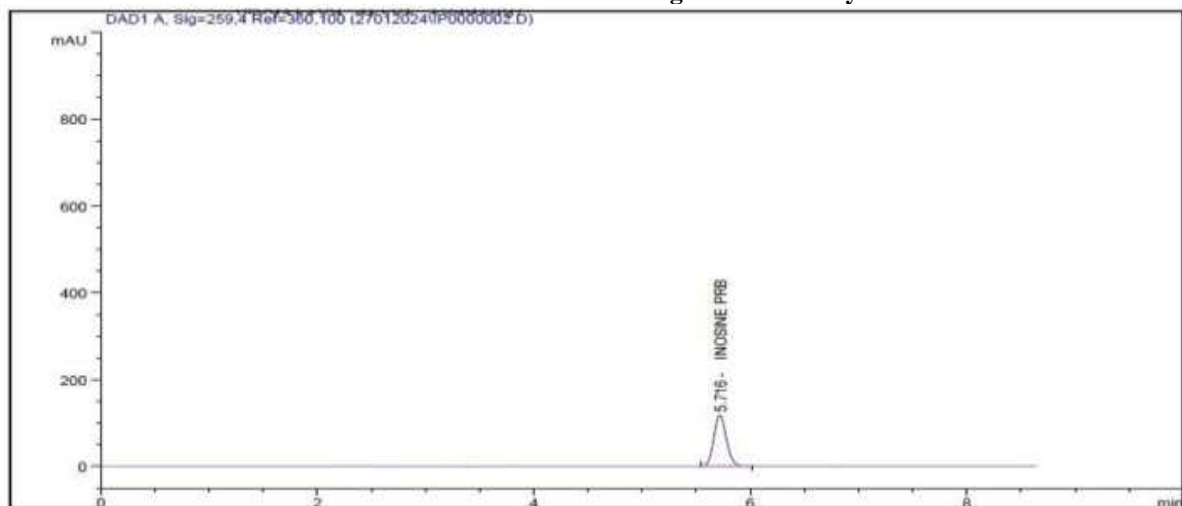


Fig. 17 Chromatogram of linearity (10mcg)-01

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.716	956.42035	12031	0.84	--

TableNo.18Detailsofchromatogram ofLinearity(10mcg)-01

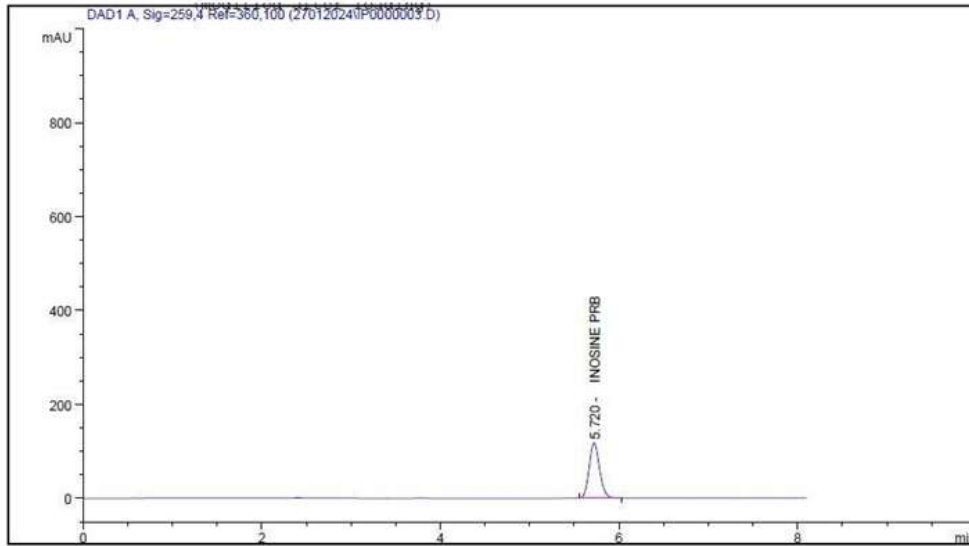


Fig.18Chromatogramoflinearity(10mcg)-02

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.720	957.35382	11788	0.84	--

TableNo.19DetailsofchromatogramofLinearity(10mcg)-02

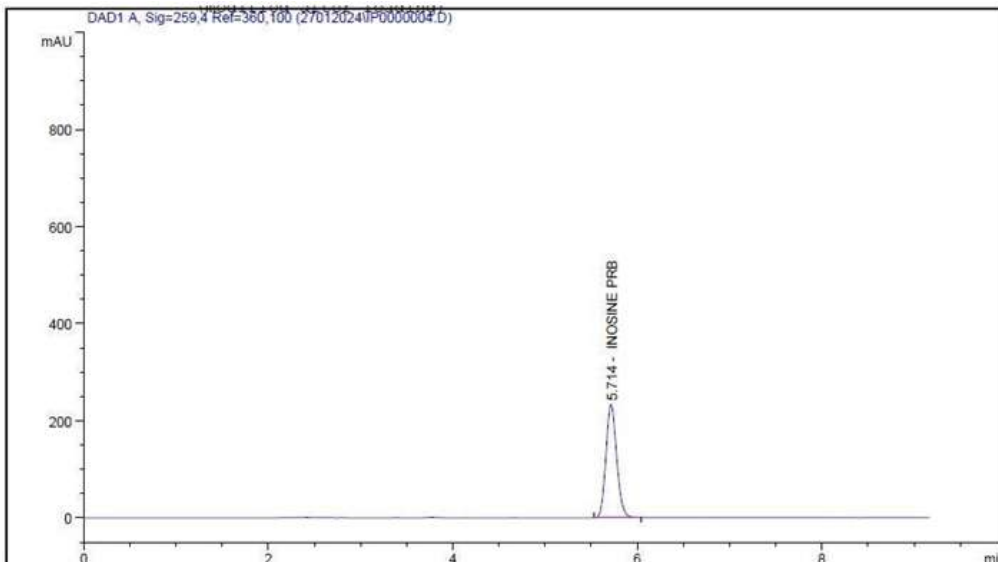


Fig.19Chromatogramoflinearity(20mcg)-01

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.714	1894.02795	11762	0.84	--

TableNo.20Detailsofchromatogram ofLinearity(20mcg)-01

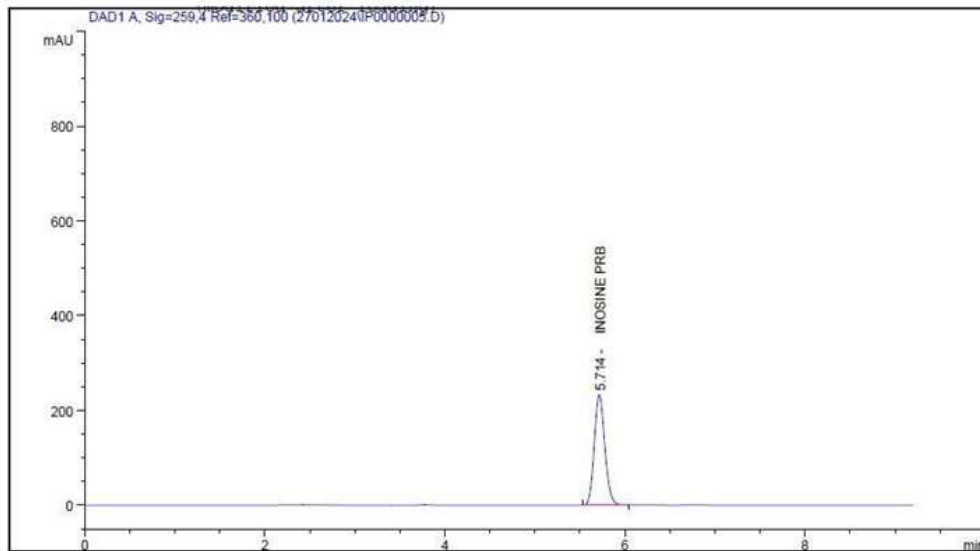


Fig.20Chromatogramoflinearity(20mcg)-02

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.714	1896.92249	11762	0.84	--

TableNo.21Detailsofchromatogram ofLinearity(20mcg)-02

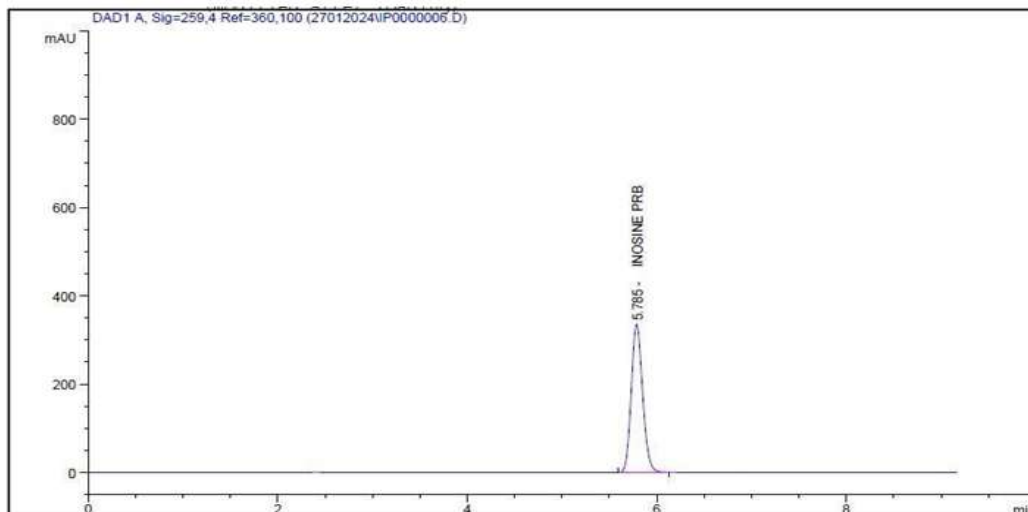


Fig.21Chromatogramoflinearity(30mcg)-01

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.785	2791.78613	11557	0.84	--

TableNo.22Detailsofchromatogram ofLinearity(30mcg)-01

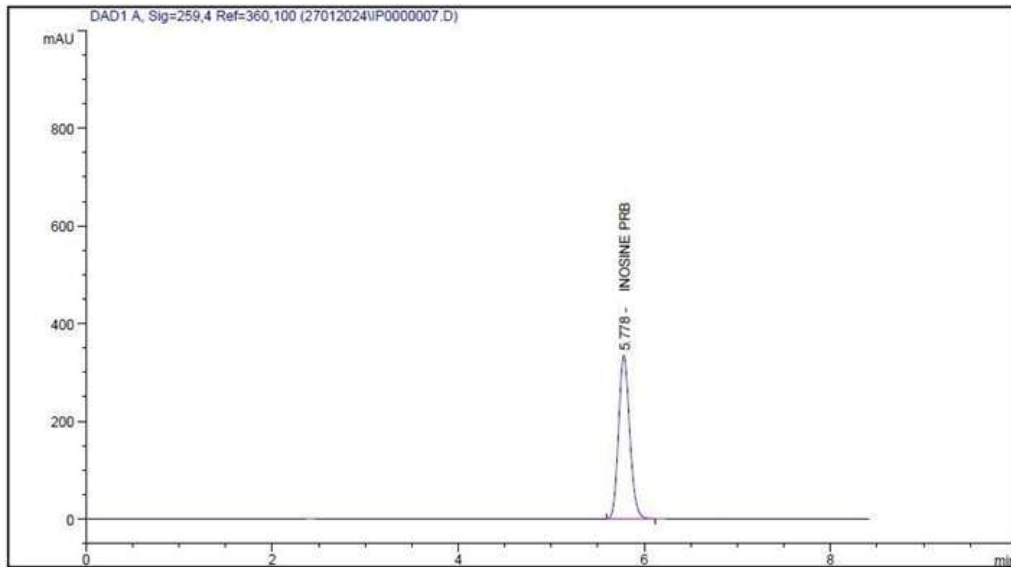


Fig.22Chromatogramoflinearity(30mcg)-02

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.778	2789.79639	11526	0.84	--

TableNo.23Detailsofchromatogram ofLinearity(30mcg)-02

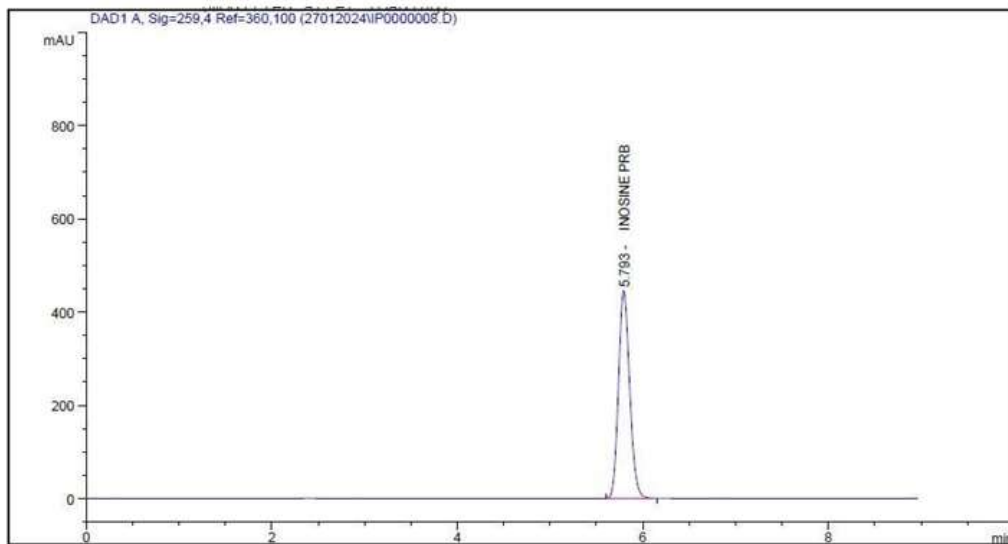


Fig.23Chromatogramoflinearity(40mcg)-01

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.793	3746.95801	11587	0.83	--

TableNo.24Detailsofchromatogram ofLinearity(40mcg)-01

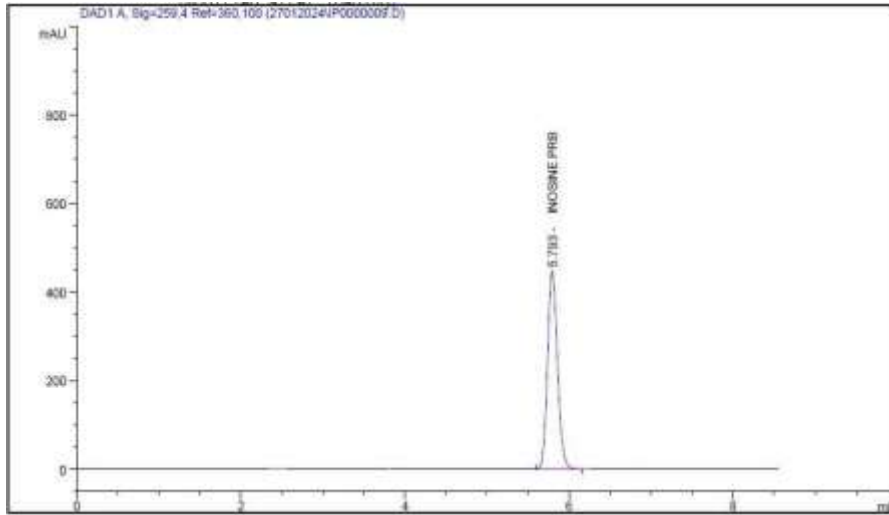


Fig.24Chromatogramoflinearity(40mcg)-02

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.793	3767.88013	11587	0.83	--

TableNo.25Detailsofchromatogram ofLinearity(40mcg)-02

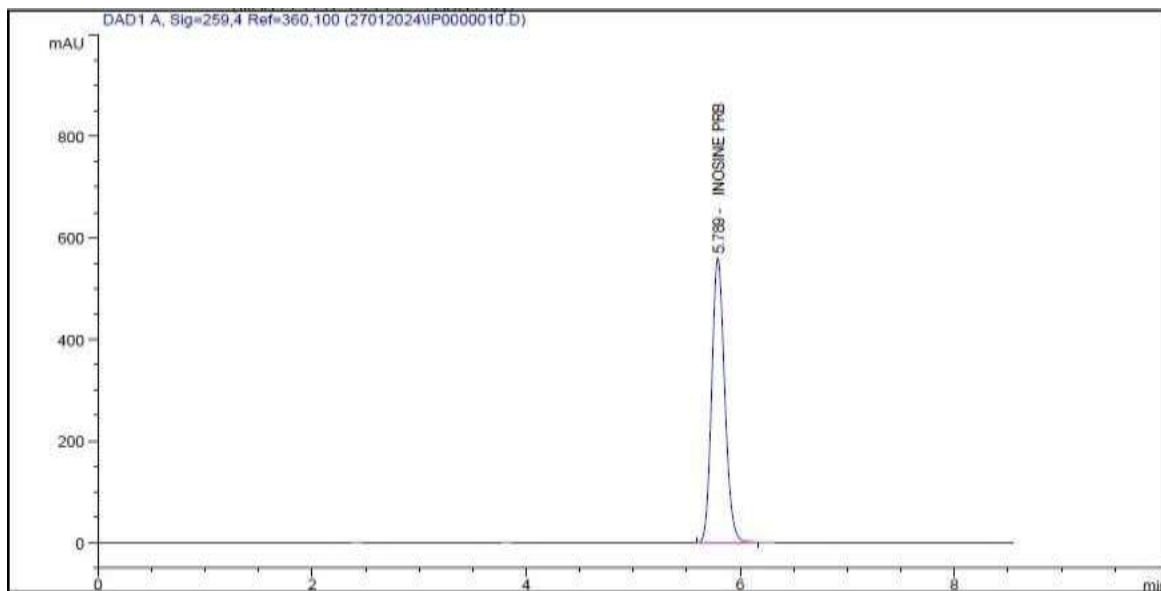


Fig.25Chromatogramoflinearity(50mcg)-01

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.789	4716.02100	11271	0.82	--

TableNo.26Detailsofchromatogram ofLinearity(50mcg)-01

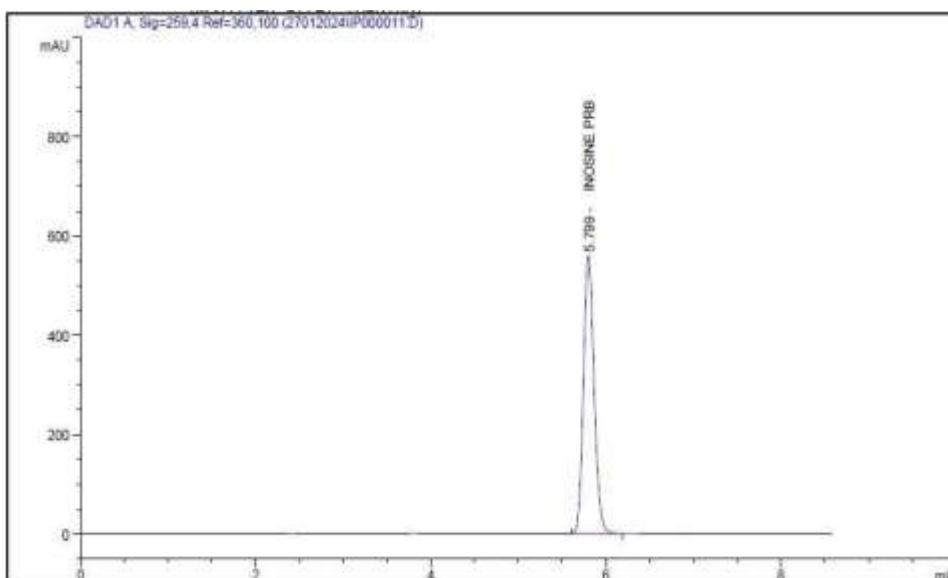


Fig.26Chromatogramoflinearity(50mcg)-02

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.799	4716.87402	11313	0.83	--

TableNo.27Detailsofchromatogram ofLinearity(50mcg)-02

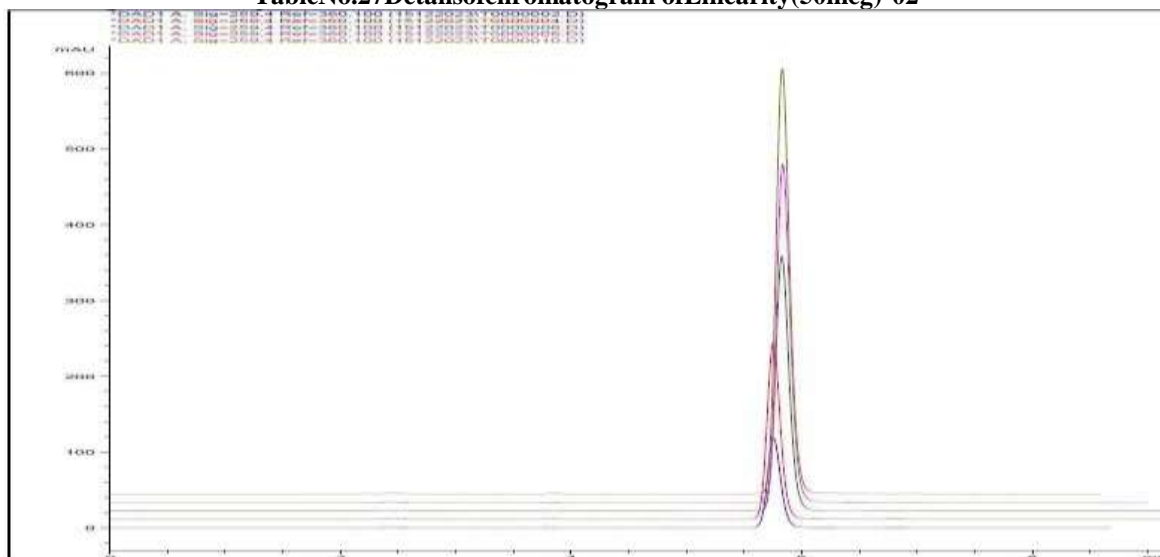


Fig.27Chromatogramofoverlaylinearity

*mean of each 5 readings for RP-HPLC method blue, red, green, pink, Peanut colours denoted by concentration 10, 20, 30, 40, 50 mcg.

Sr.No.	Concentration $\mu\text{g/ml}$ Inosine Pranobex	Area Inosine Pranobex
10	956.0263	956.4506
20	1894.0279	1896.9224
30	2791.7861	2799.7963
40	3746.9580	3767.8801
50	4716.0210	4716.8740

Table No. 38 Linearity of Inosine Pranobex

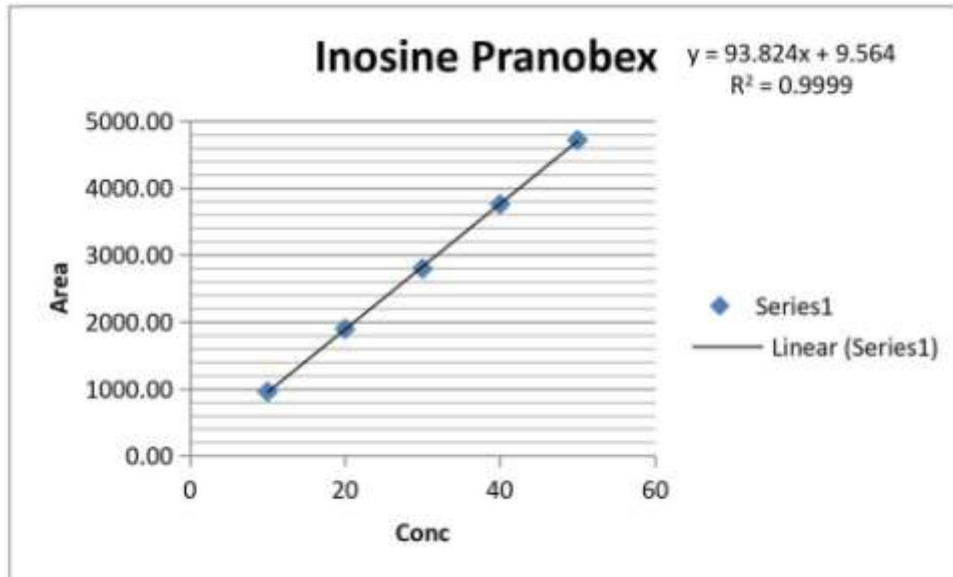


Fig. 28 Calibration curve of Inosine Pranobex for HPLC method

Linearity of Inosine Pranobex was observed in the range of 10-50 $\mu\text{g/ml}$. Detection wavelength used was 259 nm.

The plot should be linear passing through the origin; Correlation Coefficient should not be less than 0.999. that concluded. (Table No. 29).

RegressionEquationDataY=mx+c	
Slope(m)	93.82
Intercept(c)	9.564
CorrelationCoefficient	0.999

TableNo.29RegressionequationdataforInosinePranobex

2. Accuracy:-

Recovery studies were performed to validate the accuracy of developed method. Topre analyzed tablet solution, a definite concentration of standard drug (80%, 100%, and120%) was added

and then its recovery was analyzed (**Table No.32**). Statistical validationofrecovery studies shown in(**Table No. 34**).
Accuracy80%

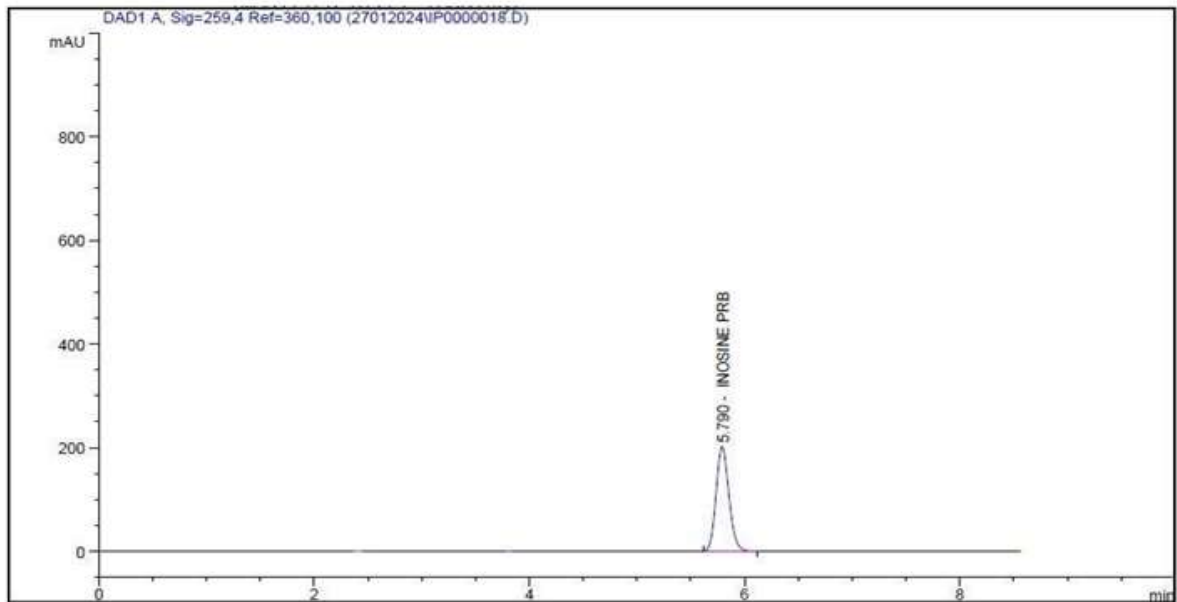


Fig.29ChromatogramofAccuracy80%

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.790	1695.55920	11822	0.84	--

TableNo.30DetailsofchromatogramofAccuracy80%

Accuracy100%

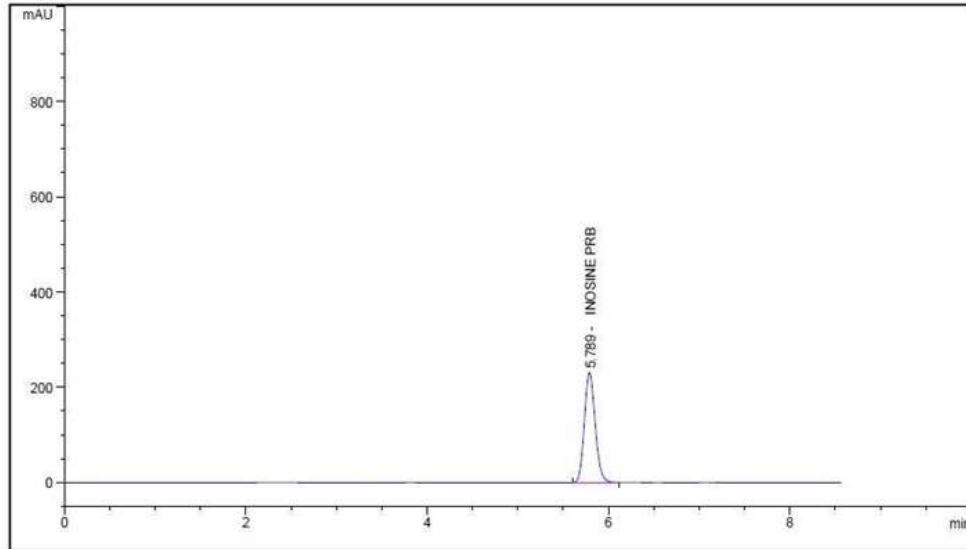


Fig.30ChromatogramofAccuracy100%

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.789	1895.37378	11571	0.84	--

TableNo.31DetailsofchromatogramofAccuracy100%

Accuracy120%

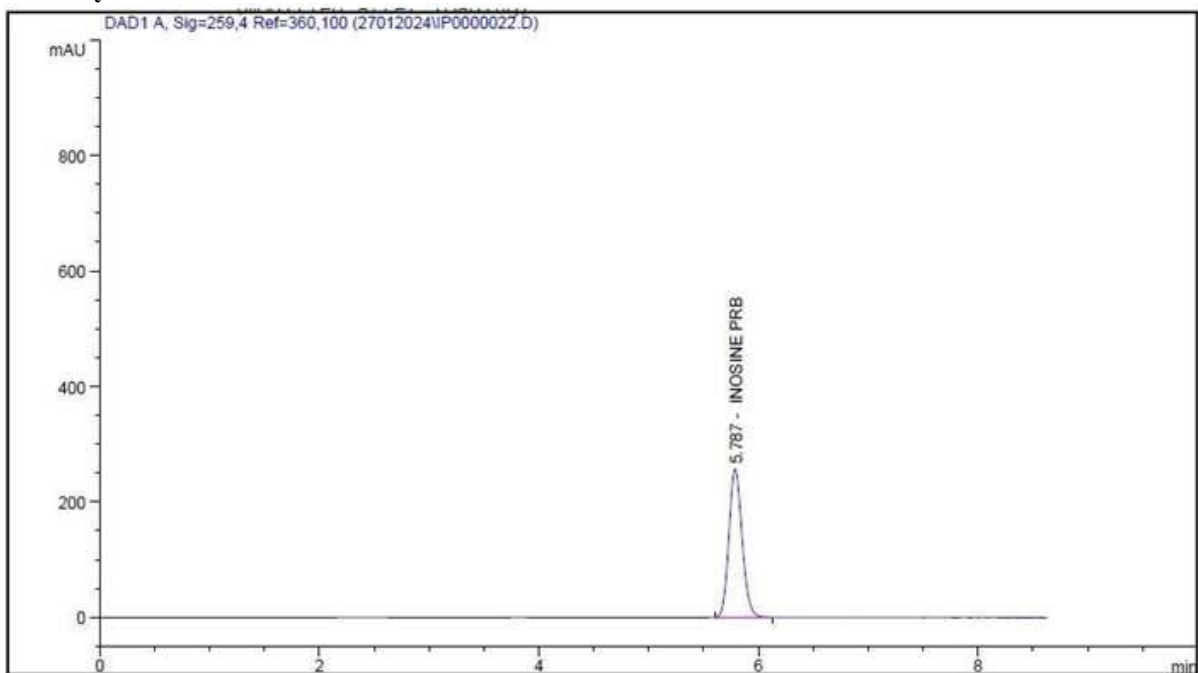
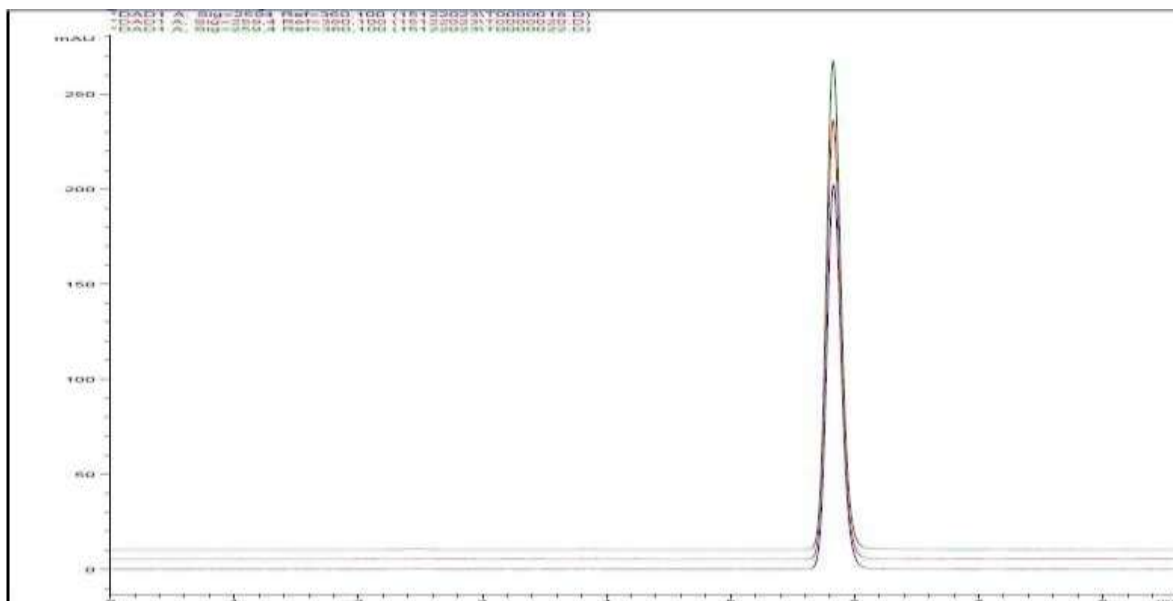


Fig.31ChromatogramofAccuracy120%

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.787	2068.91992	11563	0.84	--

TableNo.32DetailsofchromatogramofAccuracy120%



Figno:32ChromatogramofOverlayAccuracy

*meanofeach3readingforRP-HPLCmethodblue,red,greencoloursdenotedbyconcentration 80%,100%120% forISP.

METHOD	Drug	Level(%)	Amt.taken(µg/ml)	Amt.Added(µg/ml)	A.foundMean ±S.D.	Amt.recoveredMean ±S.D.	%RecoveryMean ±S.D.
RP-HPLC Method	ISP	80%	10	8	17.94±0.03	7.94±0.03	99.29±0.48
		100%	10	10	20.08±0.03	10.08±0.03	100.75±0.36
		120%	10	12	21.93±0.02	11.93±0.023	99.45±0.19

*meanofeach 3readingforRP-HPLCmethod.

TableNo.33ResultofRecoverydataforInosinePranobex

METHOD	Drug Level(%)	Mean %Recovery	StandardDeviation*	%RSD
RP-HPLC ISP Method	80%	99.29	0.48	0.49
	100%	100.75	0.36	0.35
	120%	99.45	0.19	0.19

*Denotes average of three determinations for RP-HPLC method Table No.34 Statistical Validation of Recovery Studies Inosine Pranobex

Accuracy of RP-HPLC method is ascertained by recovery studies performed at different levels of concentrations (80%, 100% and 120%). The % recovery was found to be within 98-

101% (Table No. 33,34).

3. System suitability parameters: (Repeatability)

To ascertain the resolution and reproducibility of the proposed chromatographic system for estimation of Inosine Pranobex system suitability parameters were studied. The result shown in below (Table No.35)

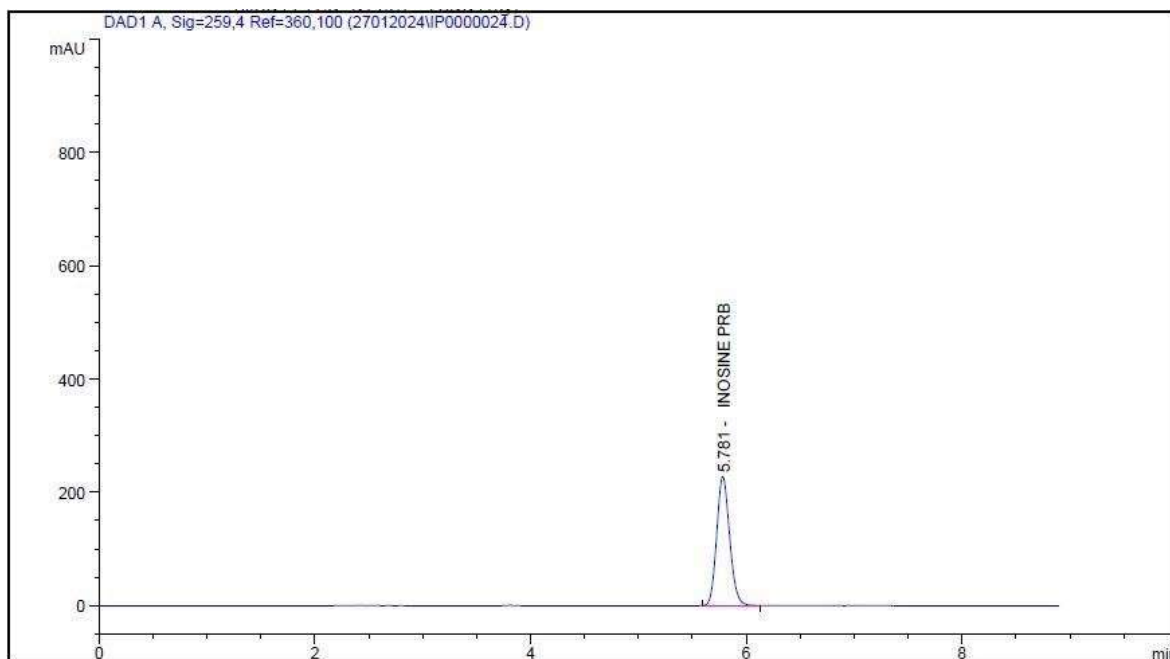


Fig.34 Chromatogram of System suitability-1

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.781	1902.08423	11540	0.84	--

TableNo.35Detailsofchromatogramofsystemsuitability-1

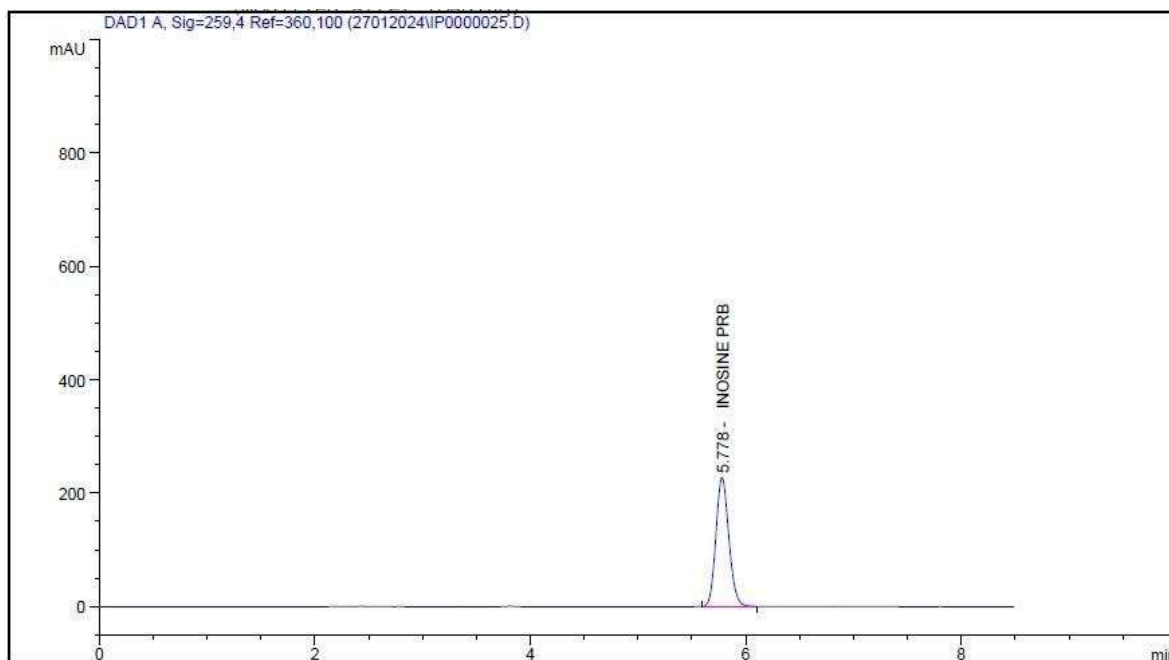


Fig.35ChromatogramofSystemsuitabilityNo-2

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.778	1902.49988	11529	0.84	--

TableNo.36Detailsofchromatogramofsystemsuitability-2

Sr.No.	ConcentrationofInosinePranobex(mg/ml)	Peakarea	Amountfound(mg)	% Amountfound
1	20	1902.084	20.09	100.46
2	20	1902.500		
		Mean	20.09	100.46
		SD	0.29	0.29
		%RSD	0.02	0.02

TableNo.37:RepeatabilitystudiesonRP-HPLCforInosinePranobex

Repeatability studies on RP-HPLC method for Inosine Pranobex was found to be The %RSD was less than 2%, which shows high percentage amount found in between 98% to 102% indicates the analytical method that concluded (Table No. 37).

The method was established by analyzing various replicates standards of Inosine Pranobex. All the solution was analyzed thrice in order to record any intra-day & inter-day variation in the result that concluded. The result obtained for intraday is shown in (Table No. 44) respectively.

4. Precision:-

Chromatogram of Intra-Day Precision:

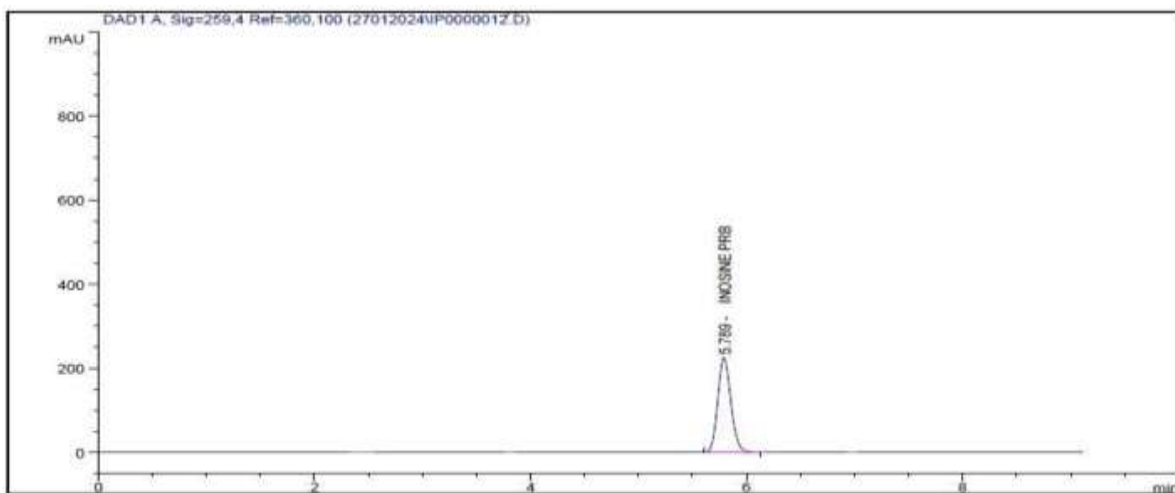


Fig.36 Chromatogram of Intra-Day Precision (20mcg)

No.	RT [min]	Area [mV*s]	TP	TF	Resolution
1	5.789	1898.54761	11573	0.84	--

Table No.38 Details of chromatogram of Intra-Day precision (20mcg)

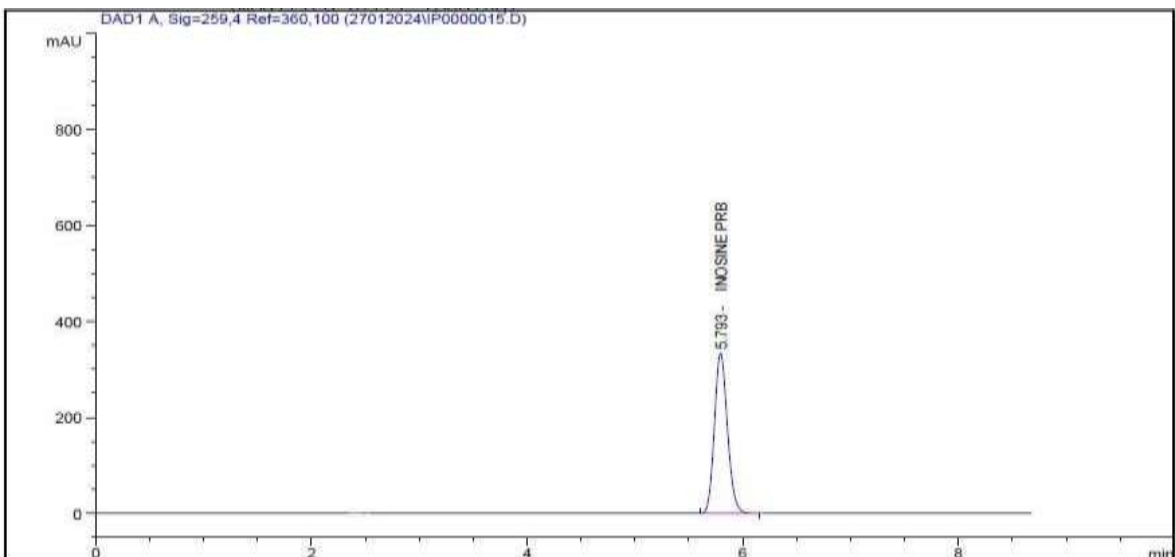


Fig.37 Chromatogram Intra-day precision-(30mcg)

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.793	2802.84131	11588	0.84	--

TableNo.39DetailsofchromatogramofIntra-Dayprecision (30mcg)

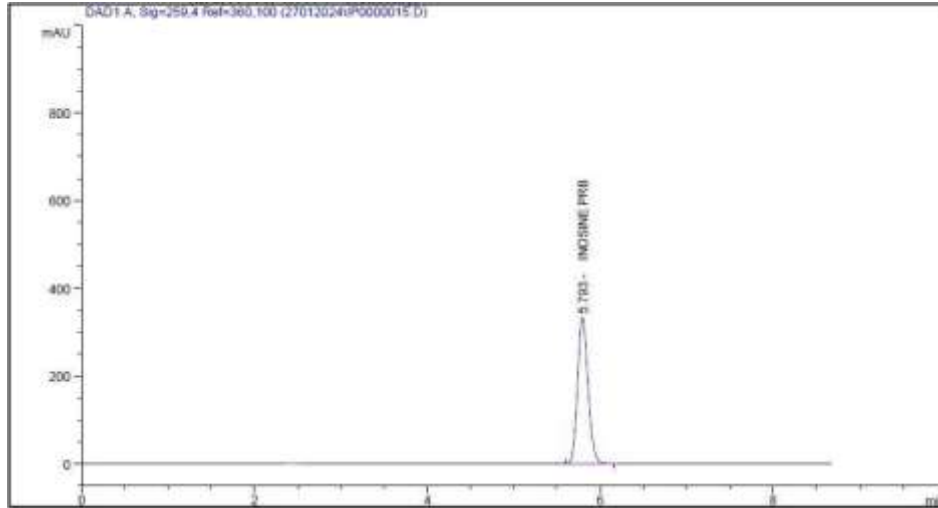


Fig.38ChromatogramIntra-dayprecision-(40mcg)

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.793	2802.84131	11588	0.84	--

TableNo.40DetailsofchromatogramofIntra-Dayprecision (40mcg)

ChromatogramofInter-DayPrecision:

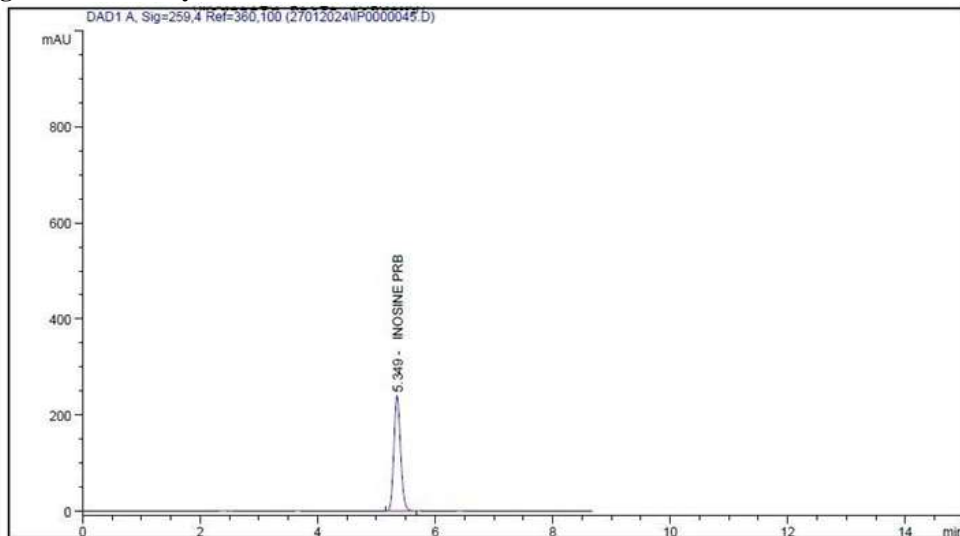


Fig.39ChromatogramInter-dayprecision(10mcg)

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.349	1897.52820	11009	0.83	--

TableNo.41DetailsofchromatogramofInter-Dayprecision(10mcg)

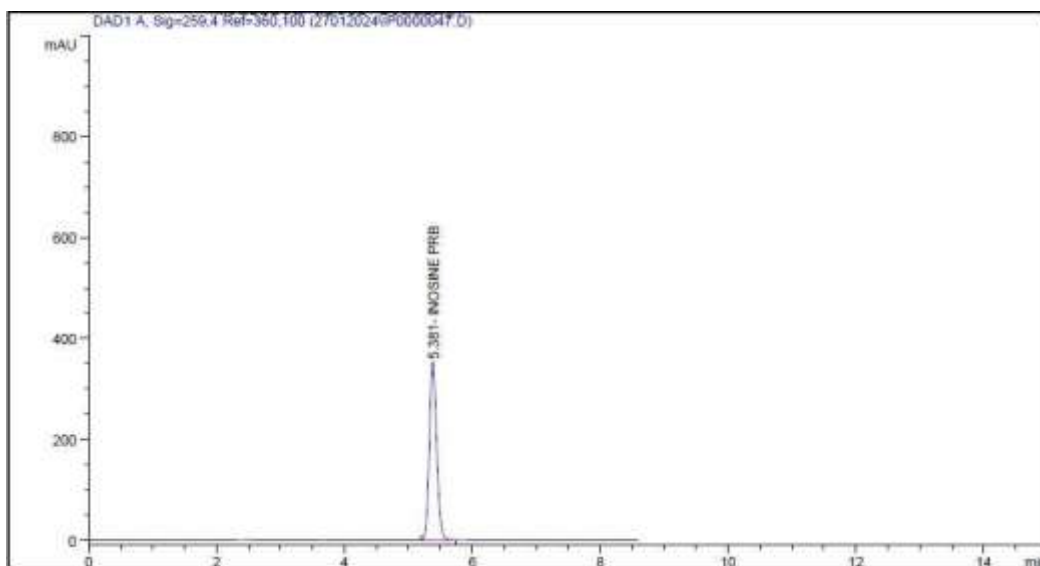


Fig.40ChromatogramInter-dayprecision(30mcg)

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.381	2802.16553	11142	0.83	--

TableNo.42DetailsofchromatogramofInter-Dayprecision(30mcg)

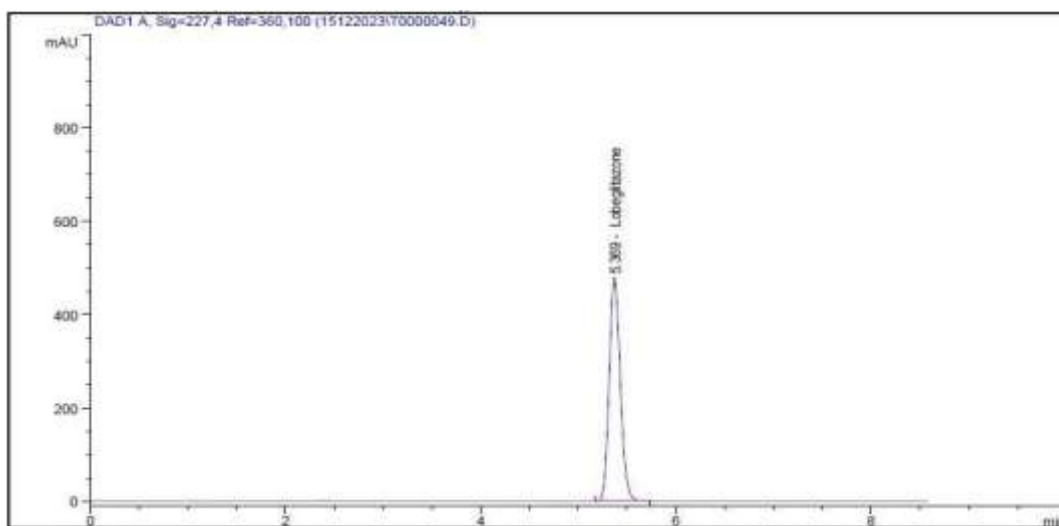


Fig.41ChromatogramInter-dayprecision(40mcg)

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.369	3783.32275	11088	0.83	--

Table No.43 Detailsof chromatogram of Inter-Day precision (40mcg)

METHOD	Drug	Conc ⁿ (µg/ml)	Intraday Precision		Interday Precision	
			Mean±SD	%AmtFound	Mean±SD	%AmtFound
HPLC METHOD Rp-	ISP	20	1899.4±1.24	100.72	1897.97±0.63	100.64
		30	2801.37±2.08	99.19	2802.49±0.46	99.23
		40	3779.43±1.18	100.45	3784.03±1.01	100.58

*Mean of each 3 reading for RP-HPLC method

Table No .44: Result of Intraday and Inter day Precision studies on RP-HPLC method for Inosine Pranobex

Intra day and Inter day Precision studies on RP-HPLC method for Inosine Pranobex which shows the high precision % amount in between 98% to 102% indicate to analytical method that concluded.

5. Robustness:

The Robustness of a method is its ability to remain unaffected by small deliberate changes in parameters. To evaluate the robustness of the proposed method, small but deliberate variations in the optimized method parameters were done. The

effect of changes in mobile phase composition and flow rate, wavelength on retention time and tailing factor of drug peak was studied.

The mobile phase composition was changed in (±1 ml/min⁻¹) proportion and the flow rate was varied by (±1 ml/min⁻¹), and wavelength change (±1 ml/min⁻¹) of optimized chromatographic condition. The results of robustness studies are shown in (Table No.45). Robustness parameters were also found satisfactory, hence the analytical method would be concluded.

1. Flow Rate Change 0.9 ml

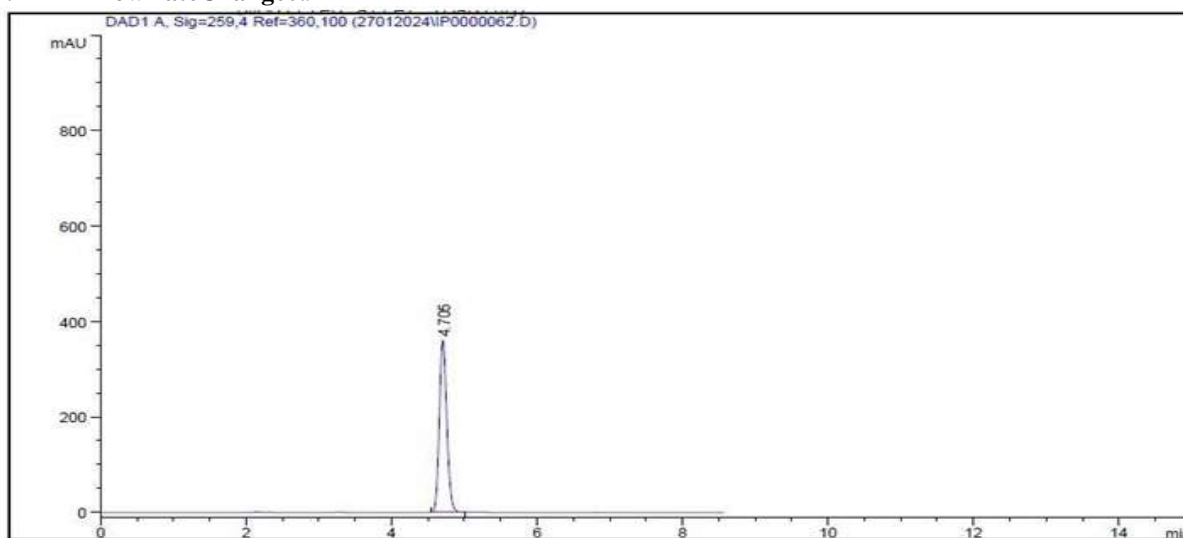


Fig.42 Chromatogram of Flow rate change 0.9ml

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	4.705	2581.92017	10258	0.84	--

Table No.45 Detailsof chromatogram of Flow rate change 0.9ml

2. FlowRateChange0.7 ml

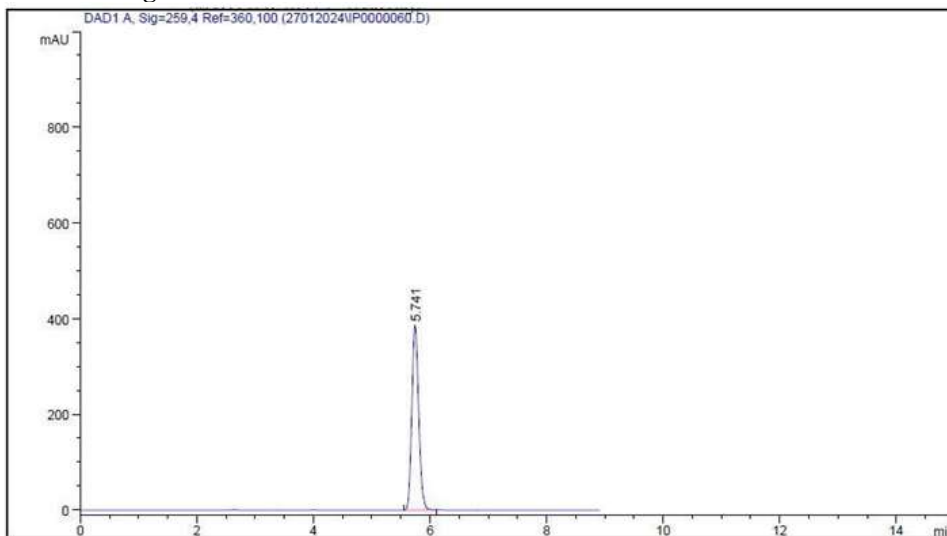


Fig.43.ChromatogramofFlowratechange0.7ml

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	4.705	2581.92017	10258	0.84	--

TableNo.46DetailsofchromatogramofFlowratechange0.7ml

3. MobilephasecompositionChange:44mlMeoH+56mlWater

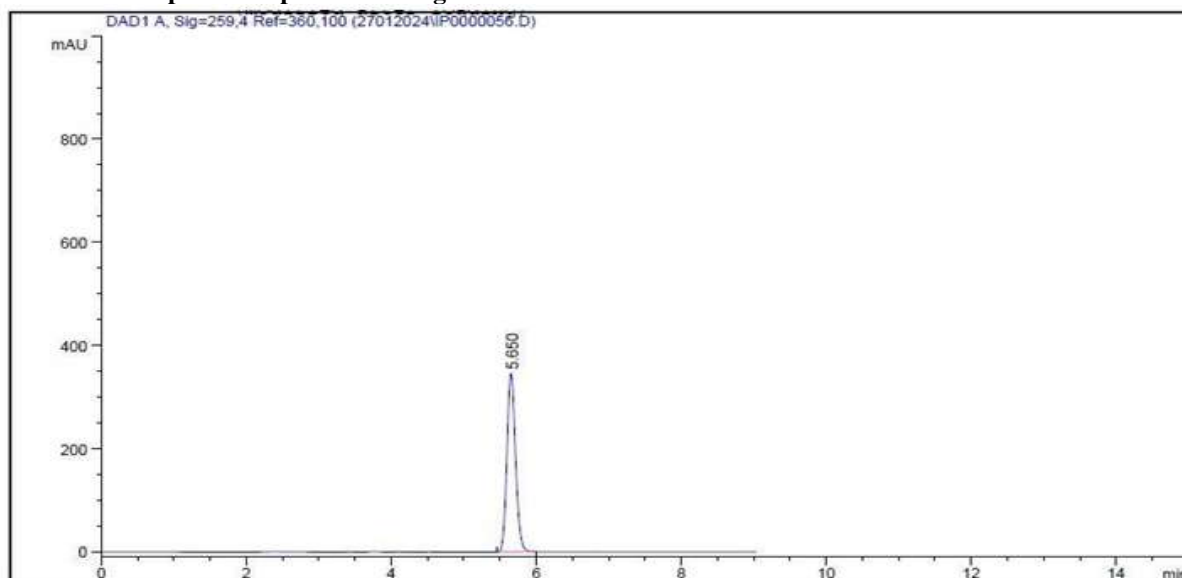


Fig.44ChromatogramofMobilephasecompositionchange:44mlMeoH+56mlWater

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.650	2848.25928	11258	0.84	--

TableNo.47DetailsofchromatogramofMobilephasecompositionchange

4. MobilephasecompositionChange:46mlMEOH+54mlWater

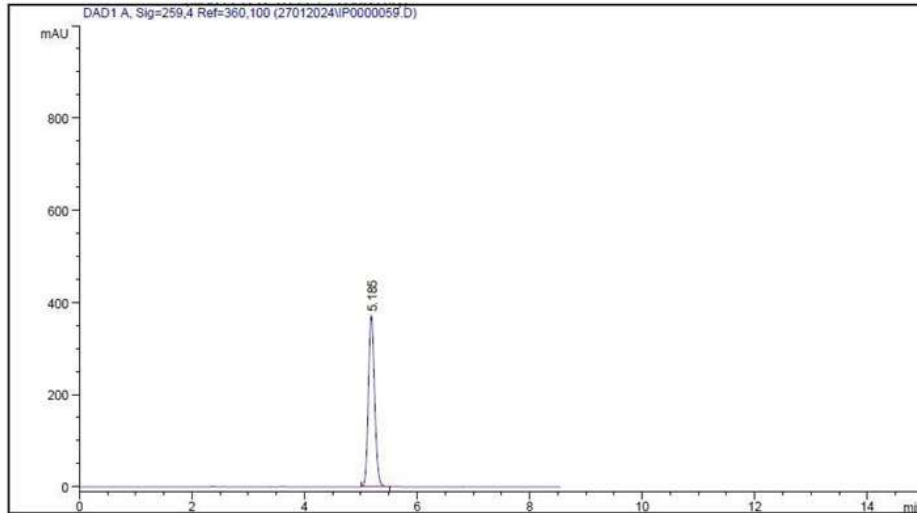


Fig.45ChromatogramofMobilephasecompositionchange46mlMEOH+ 54 ml water

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.185	2844.2246	11070	0.84	--

TableNo.48DetailsofchromatogramofMobilephasecompositionchange

5. WavelengthChange258nm

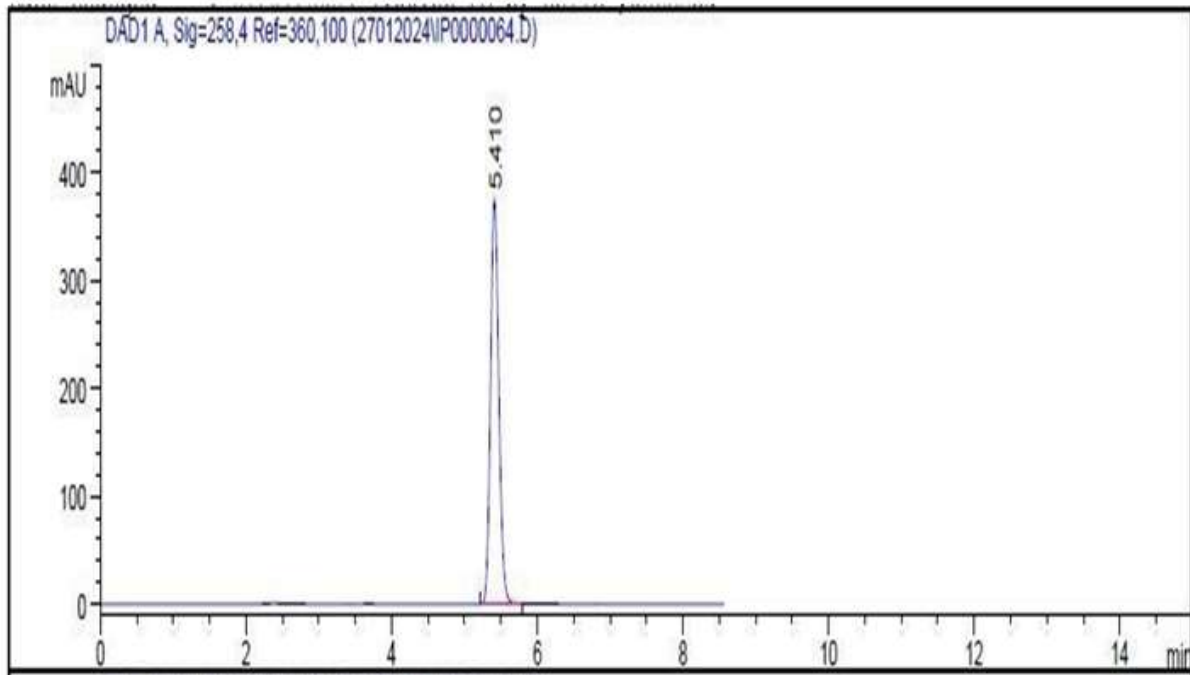


Fig.46Chromatogramofcompchangewavelengthchange258nm

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.410	2975.02930	11014	0.84	--

TableNo.49Detailsofchromatogramofwavelengthchange258nm

6. WavelengthChange228nm

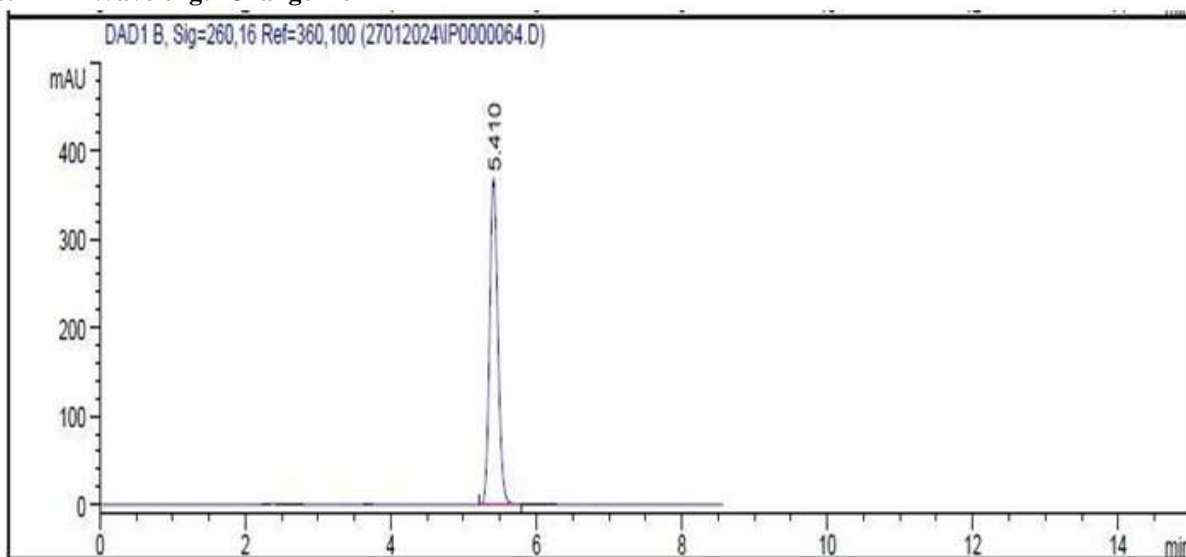


Fig.47Chromatogramofcompchangewavelengthchange260nm

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.410	2921.20142	11014	0.84	--

TableNo.50Detailsofchromatogramofwavelengthchange260nm

Parameters	Conc.	Amount ofdetected(mean±SD)	%RSD
Mobilephasecomposition-(44+56)	30	2849.4230±1.64	0.06
Mobilephasecomposition-(46+54)	30	2845.03±1.14	0.04
Wavelengthchange258nm	30	2974.2±1.12	0.04
WavelengthChange260nm	30	2920.70±0.71	0.02
Flowratechange(0.7ml)	30	3165.55±1.48	0.05
Flowratechange(0.9ml)	30	2583.00±1.52	0.06

TableNo.51Resultof Robustness StudyofInosine Pranobex

Robustness Study of Inosine Pranobex:

The changes were did flow rate ($\pm 1 \text{ ml/min}^{-1}$), PH of mobile phase composition ($\pm 1 \text{ ml/min}^{-1}$), and Wavelength ($\pm 1 \text{ ml/min}^{-1}$). %RSD for peak area was calculated which should be less than 2%. the result shown in analytical method that concluded. (Table No.57).

6. Limit Detection

The LOD is the lowest limit that can be detected. Based on the S.D. deviation of the response and the slope the limit of detection (LOD) may be expressed as:

$$\text{LOD} = 3.3 \times \text{Avd. SD} / \text{Slope}$$

$$= 3.3 \times 4.68 / 93.82$$

$$= 0.1646$$

Where, SD = Standard deviation of Y intercept S = Slope

- The LOD of Inosine Pranobex was found to be 0.1646 ($\mu\text{g/mL}$) analytical method that concluded.

8. Limit Quantification

The LOQ is the lowest concentration that can be quantitatively measured. Based on the S.D. deviation of the response and the slope,

The quantitation limit (LOQ) may be expressed as:

$$\text{LOQ} = 10(\text{SD}) / \text{S}$$

$$= 10 \times 4.68 / 93.82$$

$$= 0.4990$$

Where, SD = Standard deviation Y intercept
S = Slope

- The LOQ of Inosine Pranobex was found to be 0.4990 ($\mu\text{g/mL}$) analytical method that concluded.

8.3 Analysis of tablet formulation:- Procedure:

Weigh equivalent weight of Inosine Pranobex 13.54 mg in 10 ml volumetric flask. Add about 10 ml of diluent and sonicate to dissolve it completely and make volume up to the mark with diluent. Mix well and filter through 0.45 μm filter. Further pipette 0.3 ml of the above stock solution into a 10 ml volumetric flask and dilute up to the mark with diluents (30 $\mu\text{g/mL}$). The simple chromatogram of test Inosine Pranobex Shown in (Fig No:48) the amounts of Inosine Pranobex per tablet were calculated by extrapolating the value of area from the calibration curve. Analysis procedure was repeated two times with tablet formulation. Tablet Assay for % Label claim for % RSD Calculated, Result was shown in (Table No. 52).
Brand Name: Viralex (500MG) [Themis medicare Ltd]
13.54 mg SAMPLE IN 10 ML METHANOL = 1000 $\mu\text{g/mL}$ INOSINE PRANOBEX STOCK - II
Take 0.3 ml in 10 ml Mobile Phase i.e. = 30 $\mu\text{g/mL}$ absolute

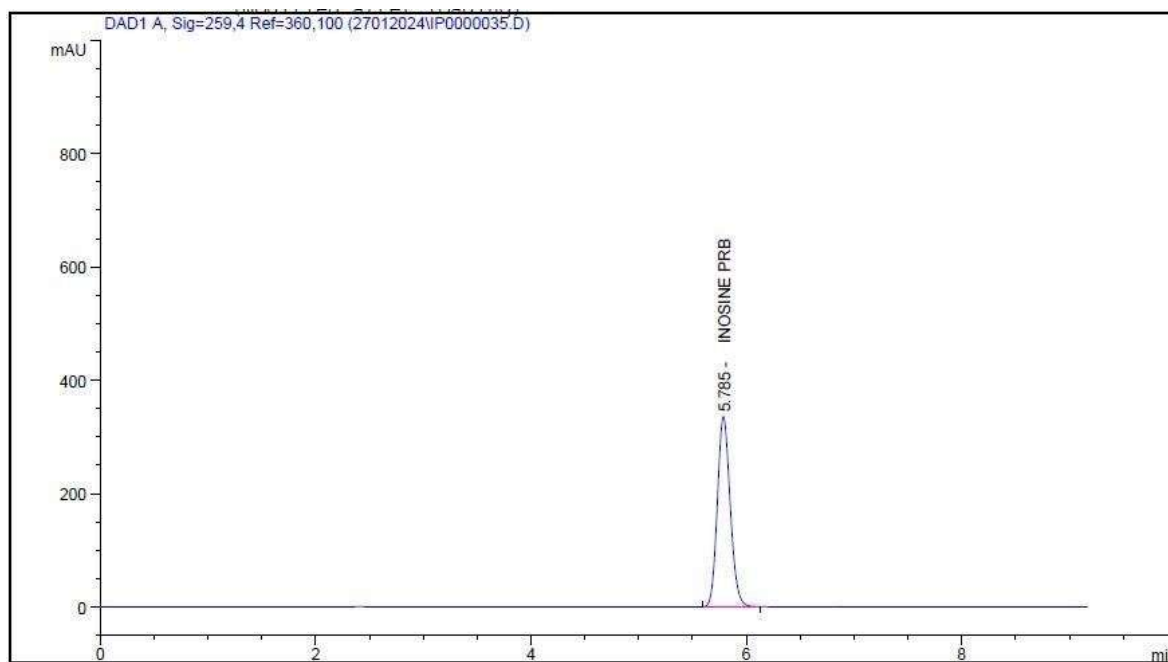


Fig.48 Chromatogram for Marketed Formulation

No.	RT[min]	Area[mV*s]	TP	TF	Resolution
1	5.785	2801.11613	11597	0.89	--

TableNo.52DetailsofchromatogramofMarketedFormulation

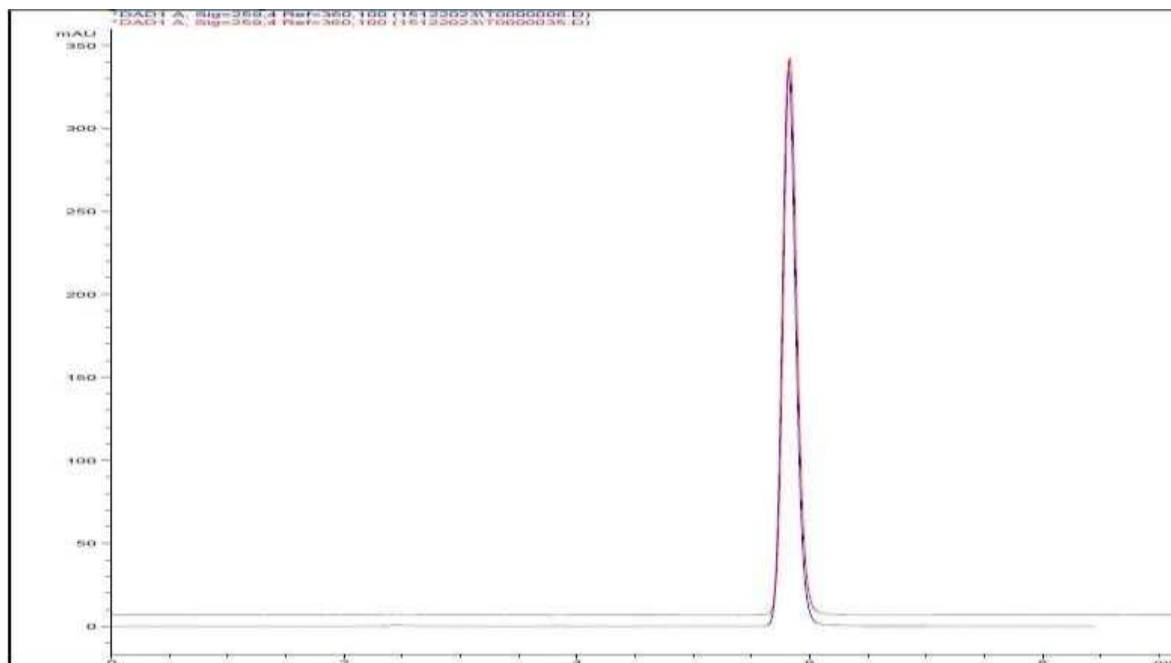


Fig.49ChromatogramforoverlayofMarketedFormulationandSTD.

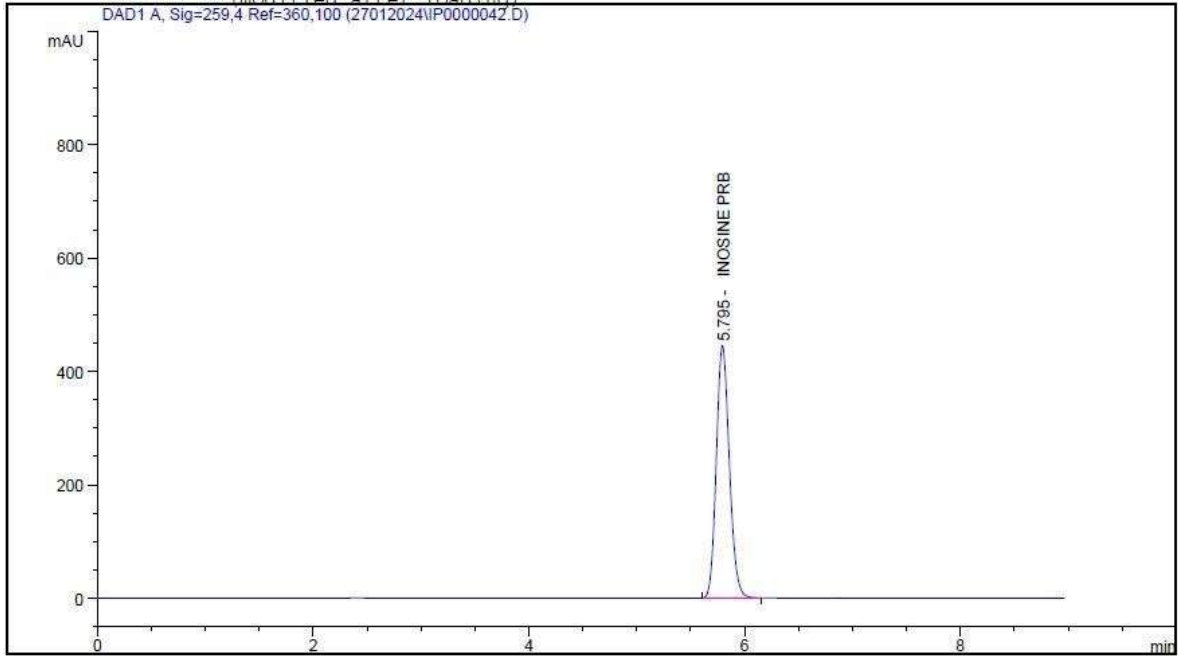
Assy	Drug	Lable Claimed	Amt. Found	% Lable Claim	SD	%RSD
Rp-HPLC Method	ISP	30	29.6723	98.91	0.11	0.35
		30	29.8215	99.41	0.35	0.35

TableNO.53Analysisofmarketedformulation

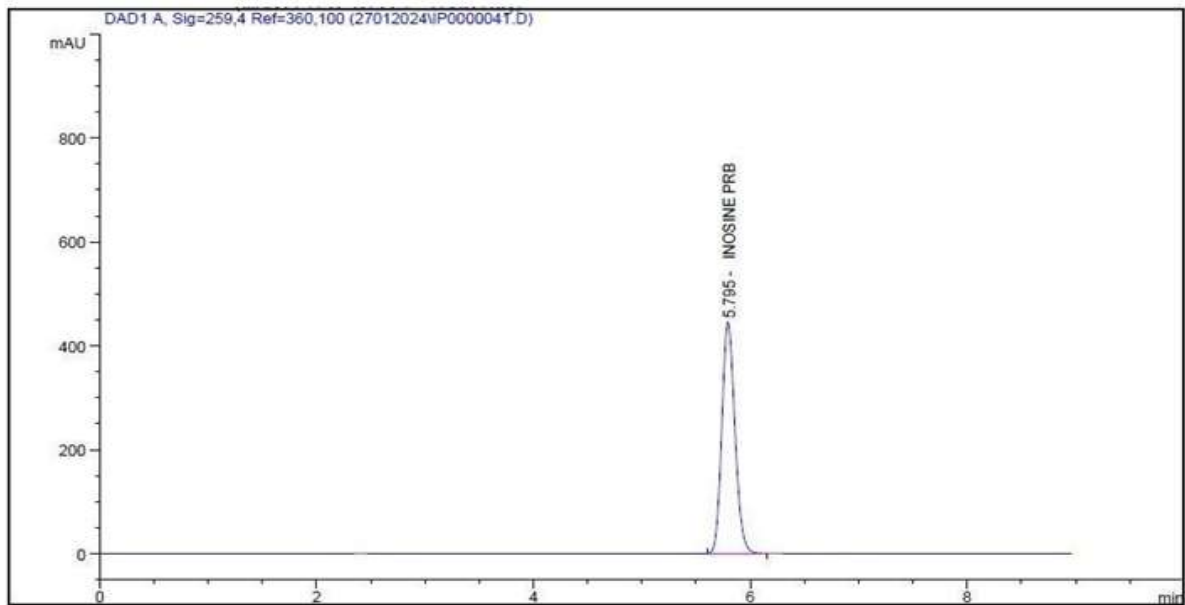
Analysisofmarketedformulationwerealso%LableCl aimwasfoundtobe98-102% Satisfactoryareconcluded.(Table No.53).

9. Ruggedness

The degree of reproducibility of test results obtained by the analysis of a sample under variety of Condition. Such as different analyst, laboratory Different instrument.



FigNo.50:ChromatogramforAnalyst-1(40mcg)



FigNo.51:ChromatogramforAnalyst-II(40mcg)

Drugname	R.T	AREA	TH.PLATES	SYMM
Analyst-I	5.795	3740.1210	12956	0.80
Analyst-II	5.795	3745.98001	12456	0.84

Table.no.55.AnalysisofAnalyst-1(40mcg)

Force of Degradation:

Degradation behavior:

Forced degradation studies of the drugs namely Inosine Pranobex were carried out individually and in combination under different stress conditions like acid hydrolysis, alkaline hydrolysis, hydrogen peroxide oxidation and photolysis. The results are shown in (Figure No: 52, 53, 54, 55 and Table no: 56)

ion under different stress conditions like acid hydrolysis, alkaline hydrolysis, hydrogen peroxide oxidation and photolysis. The results are shown in (Figure No: 52, 53, 54, 55 and Table no: 56)

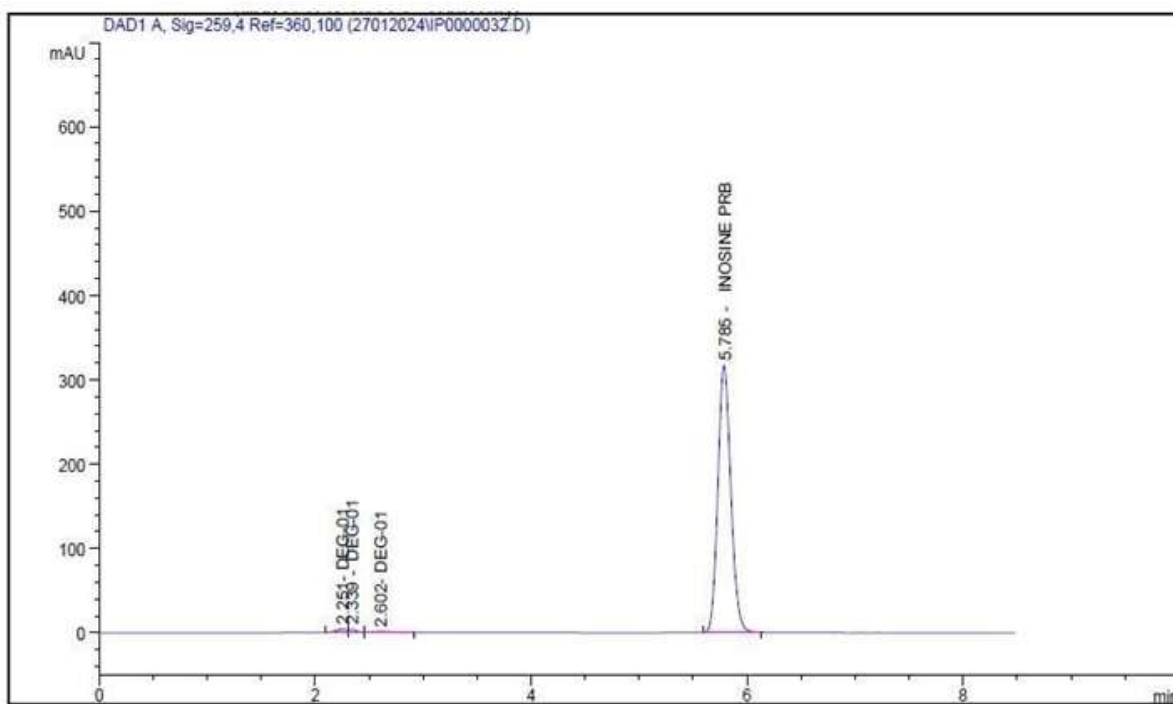
Sr no Degradation parameter		% Degradation
		2hr
1	Alkali DEG. 0.1 N NaOH- AFTER 1hr 30mcg	5.44
2	Acid DEG. 0.1 N HCL- AFTER 1hr (30mcg)	7.20
3	3% H ₂ O ₂ DEG AFTER 1hr- 30mcg	3.16
4	Neutral After 1hr- 30mcg	6.23
5	Photolytic 24Hr.	7.35

Table no: 56 Degradation of different stress condition

1. Alkaline hydrolysis:

The alkaline hydrolysis condition was performed using 0.1 N NaOH for 2hr Inosine Pranobex. Degradation of Inosine Pranobex. The major

degradation products for Inosine Pranobex were observed at 2 hr % degradation 5.44% respectively.



FigNo: 52 Degradation for 0.1 N NaOH at 2hr

Acid hydrolysis:

The acid hydrolysis performed using 0.1 N HCl 1hr min for all three Inosine

Pranobex indicated degradation. The major degradation products for Inosine Pranobex were observed at 2 hr, % degradation 7.20% the

degradation product was observed at RRT of 2 hr. These impurities were also

in Inosine Pranobex.

detected

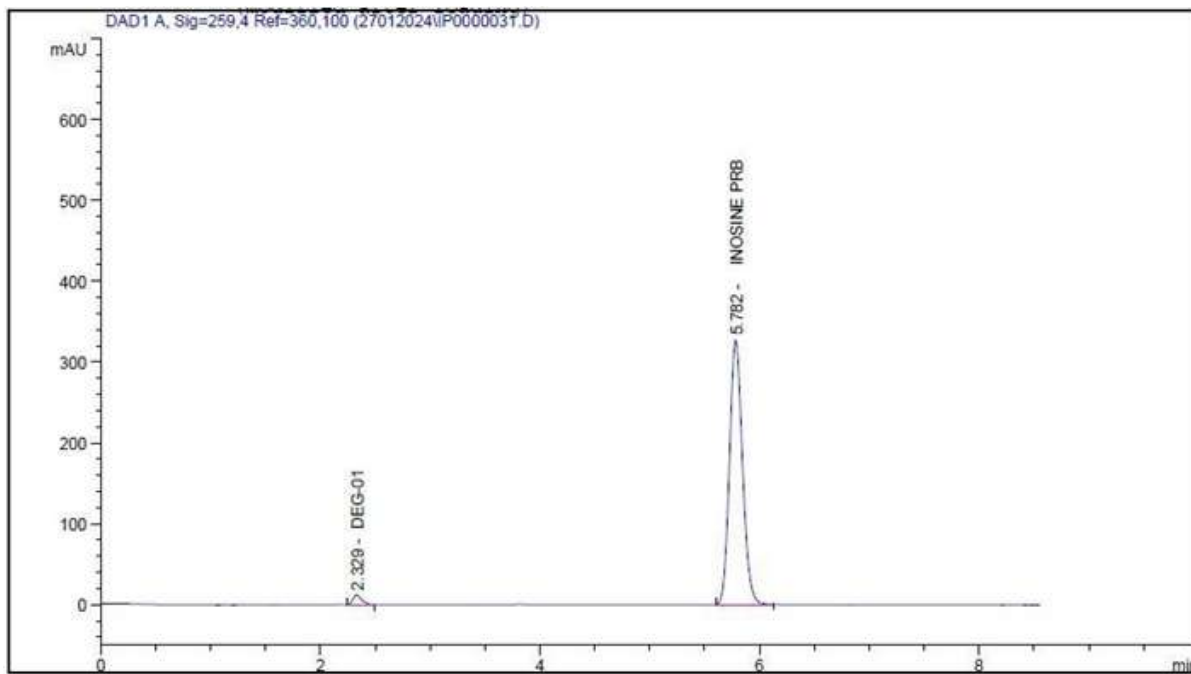


Fig.No:53 Degradation for 0.1NHCL at 2Hr

3. Oxidation

In the oxidation condition with 3% H₂O₂ for 2hr Inosine Pranobex show

oxidative stress degradation peak in the chromatogram observed at 2hr % degradation (2hr for 3.16%).

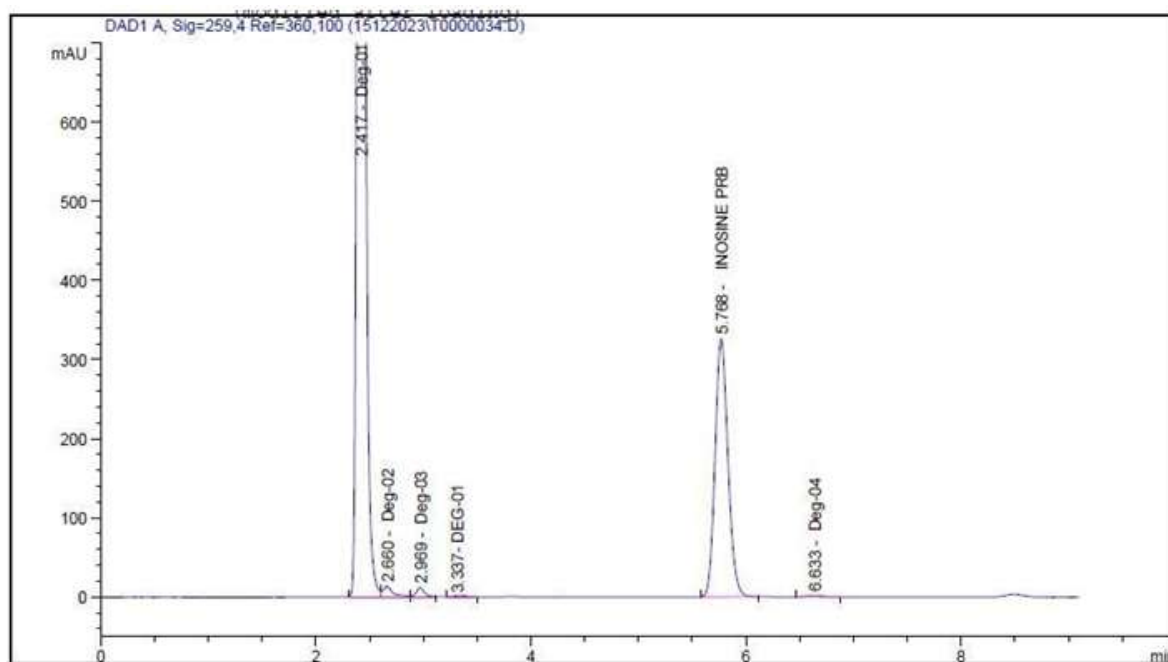
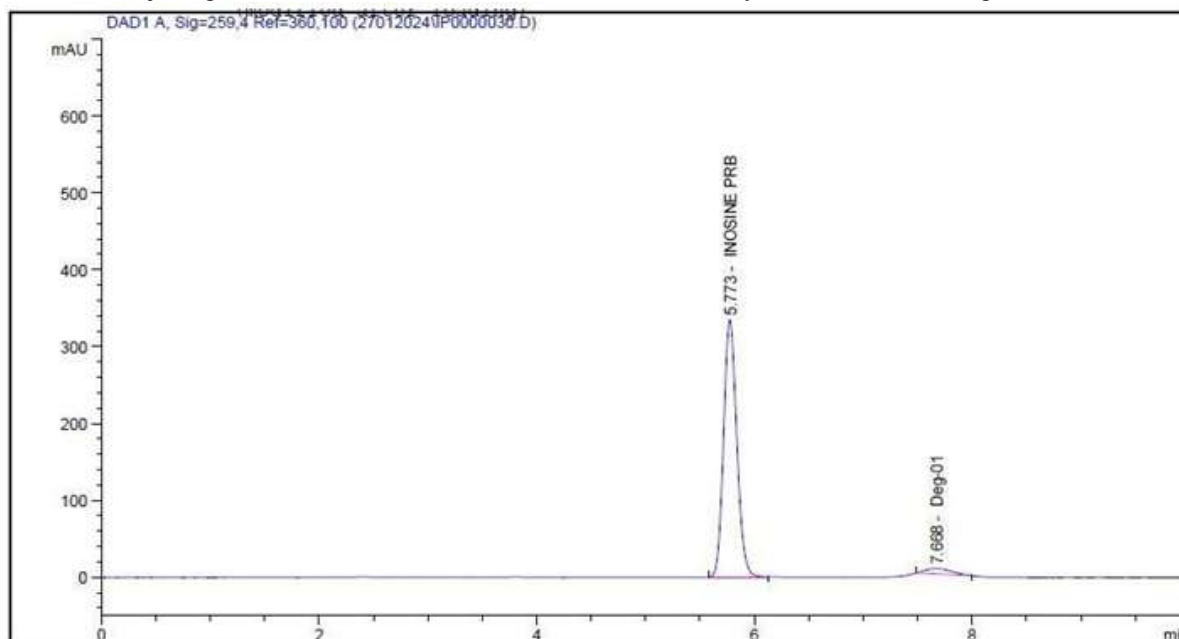


Fig.No.54 Degradation for 3% H₂O₂ at 2hr.

4. Neutral Studies:

There was no major degradation observed for Inosine Pranobex and hence they were not sensitive to light.



FigNo.55: Degradation for neutral at 2hr.

III. SUMMARY AND CONCLUSIONS

The present work deals with the Development and validation of RP-HPLC using for determination of Inosine Pranobex by pure and Tablet dosage form.

Summary of RP-HPLC method

Attempts were made to develop RP-HPLC for estimation of Inosine Pranobex from tablet. For the RP - HPLC Agilent (S.K) method Gradient System UV Detector and C18 column with 250mm x 4.6 mm I'd and 5µm particle size methanol: 0.1% water with OPA (45% : 55% v/v) was used as the mobile phase for the method. The detection wavelength was 259 nm and flow rate was 0.8 ml/min. In the developed method, the retention time of Inosine Pranobex were found to be 3.613 and 5.555 min

The developed method was validated according to the ICH guidelines. The linearity, precision, range, robustness was within the limits as specified by the ICH guidelines. Hence the method was found to be simple, accurate, precise, economic and reproducible. So, it is worthwhile that, the proposed method can be successfully utilized for the routine quality control analysis of Inosine Pranobex in bulk drug as well as in formulations.

Conclusion

Simple, rapid, accurate and precise RP-HPLC have been developed and validated for the routine analysis of Inosine Pranobex in API and tablet dosage forms. Both methods are suitable for the simultaneous determination of Inosine Pranobex in Single-component formulations without interference of each other. The developed methods are recommended for routine and quality control analysis of the investigated drugs in two-component pharmaceutical preparations. The amount found from the proposed method was in good agreement with the label claim of the formulation. Also the value of standard deviation and coefficient of variation calculated were satisfactorily low, indicating the suitability of the proposed methods for the routine estimation of tablet dosage forms.

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